

Modelling cells as contractile matter

Ulrich Schwarz

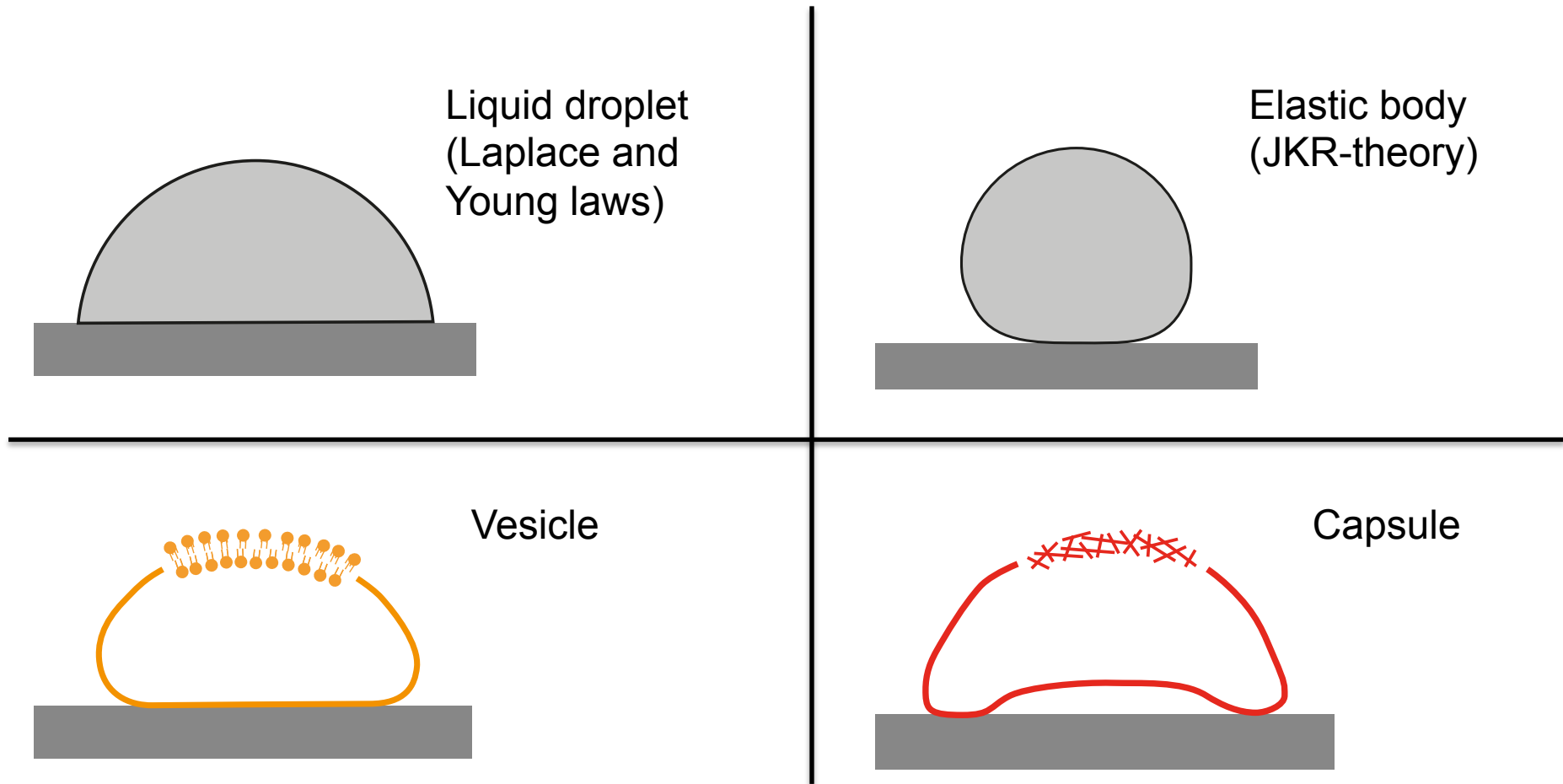
University of Heidelberg

Institute for Theoretical Physics and BioQuant



Introduction

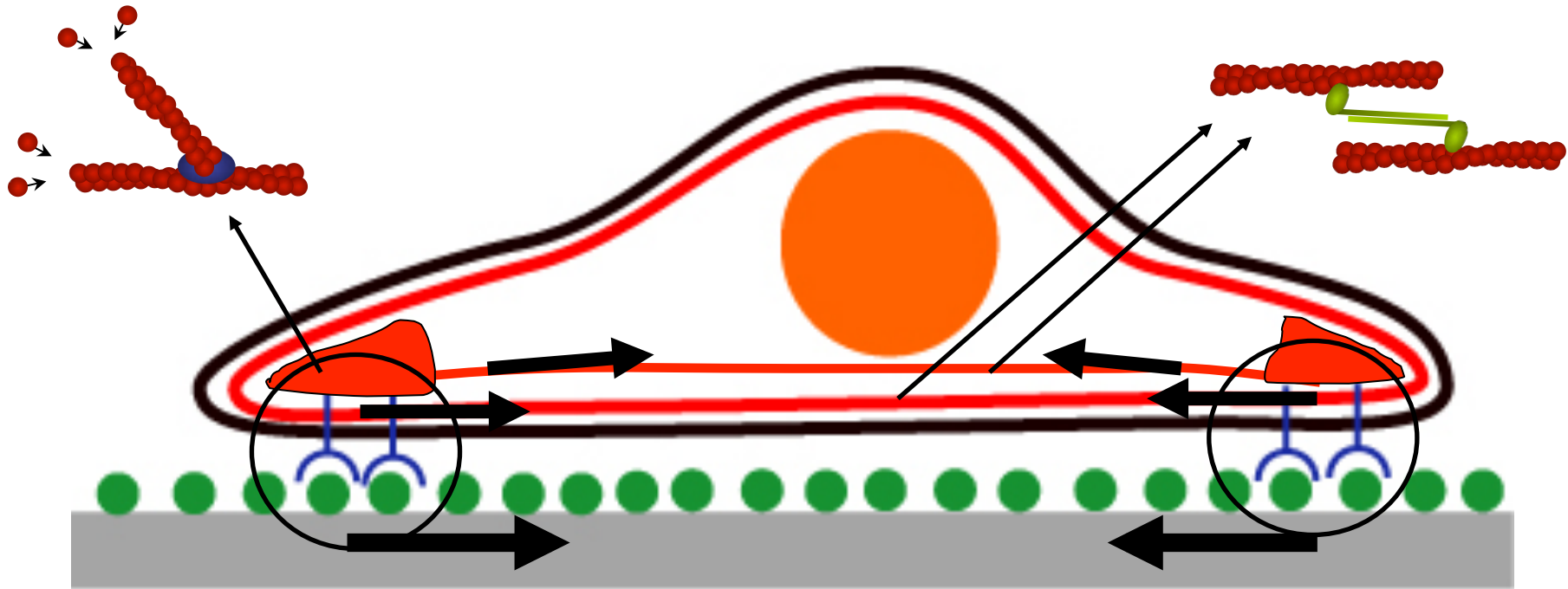
Soft matter models for cell adhesion



Cell adhesion is different because it is characterized by large contact area, tangential traction forces and localized adhesions

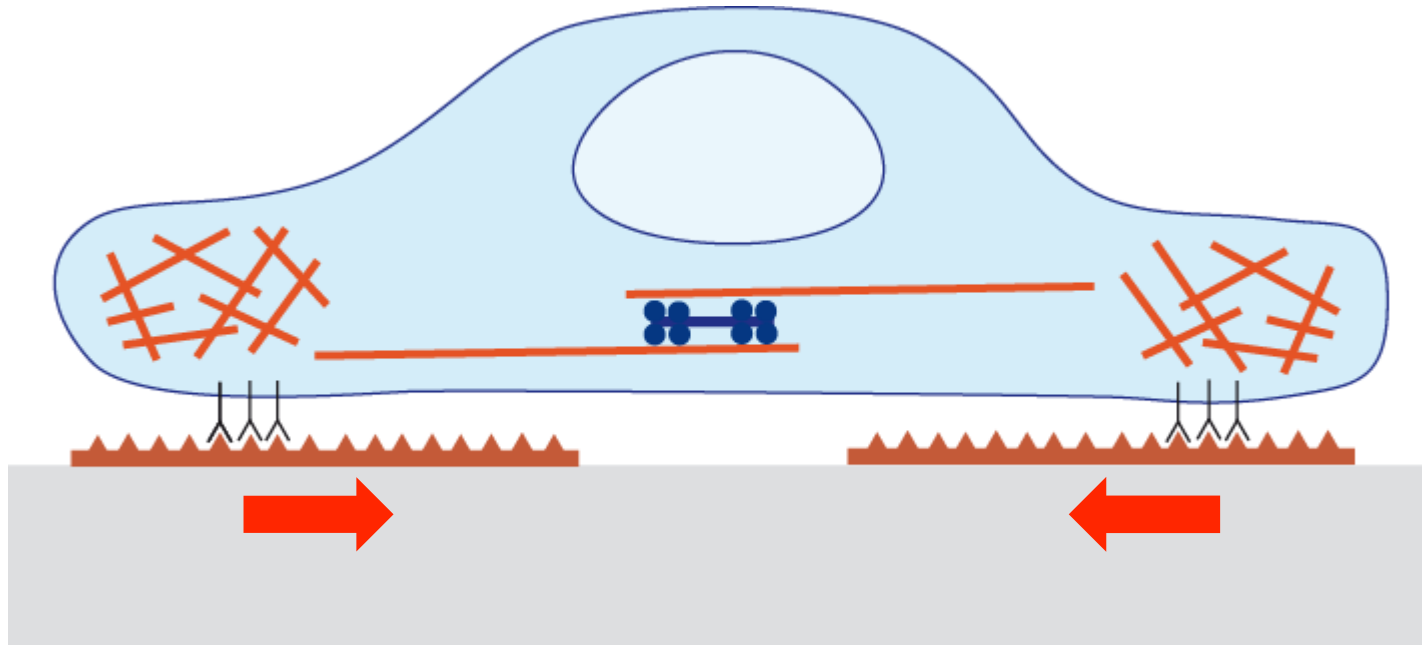
[Schwarz and Safran RMP 2013]

Active processes during cell adhesion



- actin polymerization pushes out **lamellipodia**
- contractile forces generated in **stress fibers** and **actin networks**
- force is transmitted to substrate through **focal adhesions**

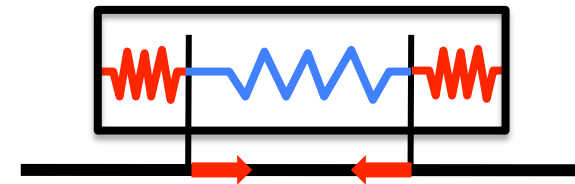
Overall force balance in the cell



Network polymerization: compressed spring

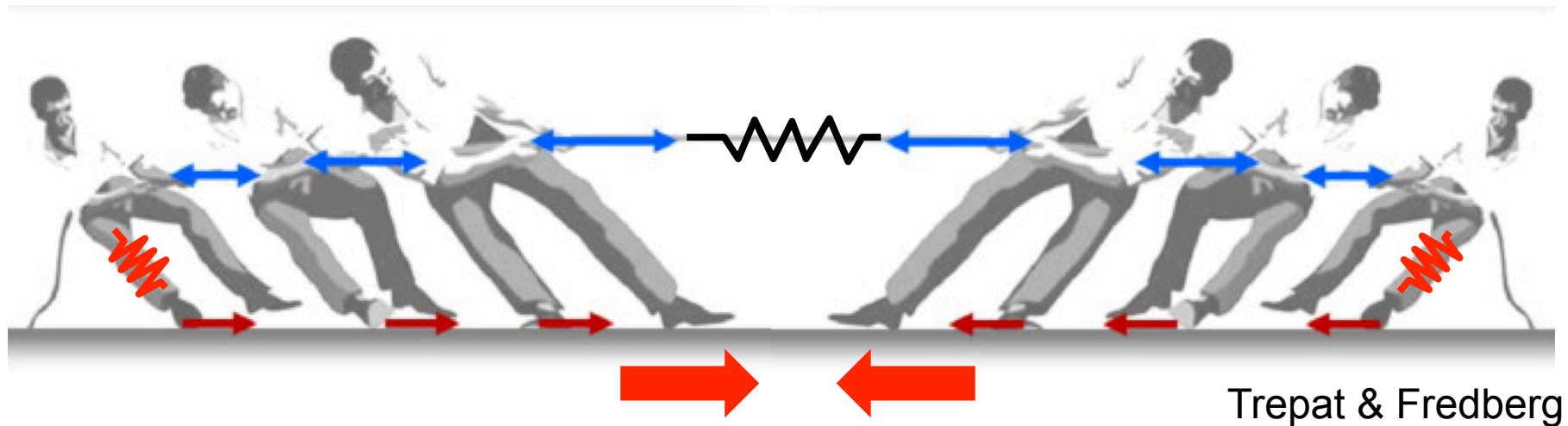
Motor contraction: tensed spring

Whole system: contraction force dipole



[Schwarz and Safran RMP 2013]

Both contraction and polymerization contribute to traction force

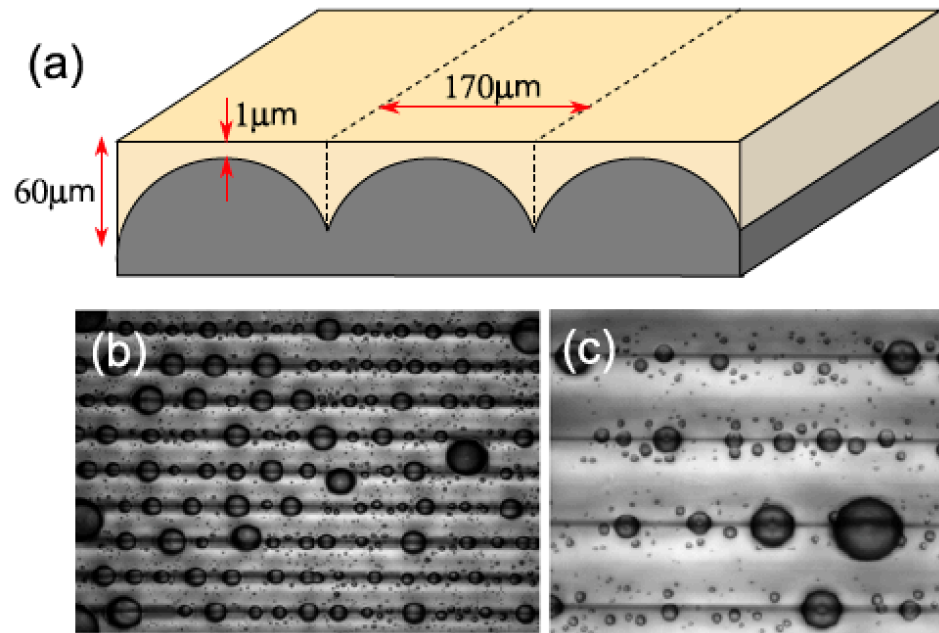


Motor contraction: tensed spring

Network polymerization: compressed spring

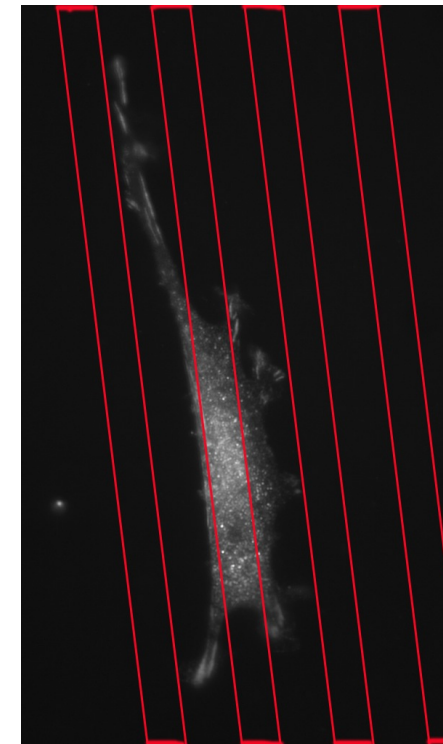
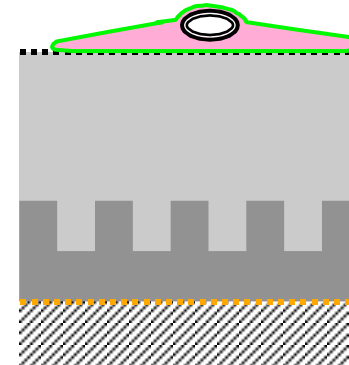
Whole system: contraction force dipole

Durotaxis of droplets versus cells



Droplets move to the positions of minimal effective stiffness

[Style et al. PNAS 2013]



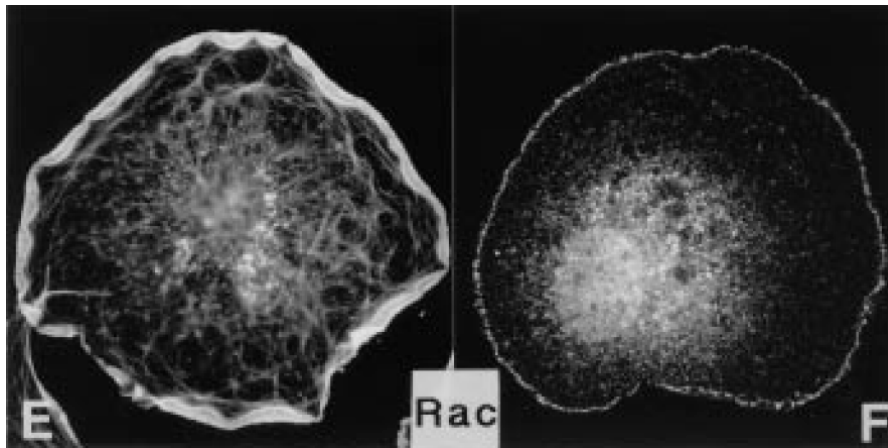
Cells move to positions of maximal effective stiffness (durotaxis)

[Mathis Riehle]

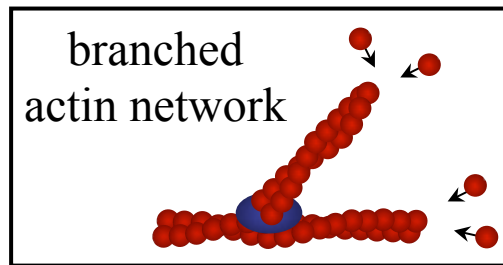
Regulation of the actin cytoskeleton through small Rho-GTPases

Ridely and Hall and coworkers Cell 1992

Rac: spreading and migration

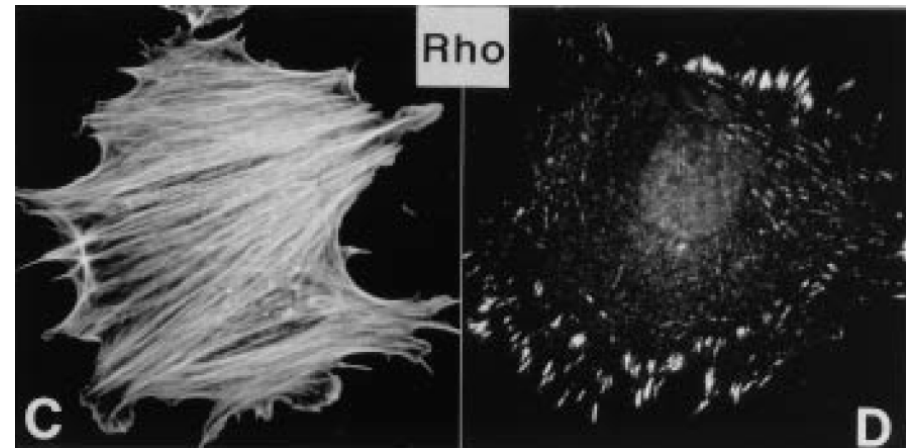


lamellipodia and focal complexes

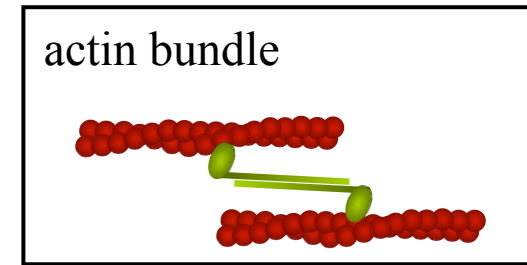


Arp2/3

Rho: mature adhesion



stress fibers and focal adhesions

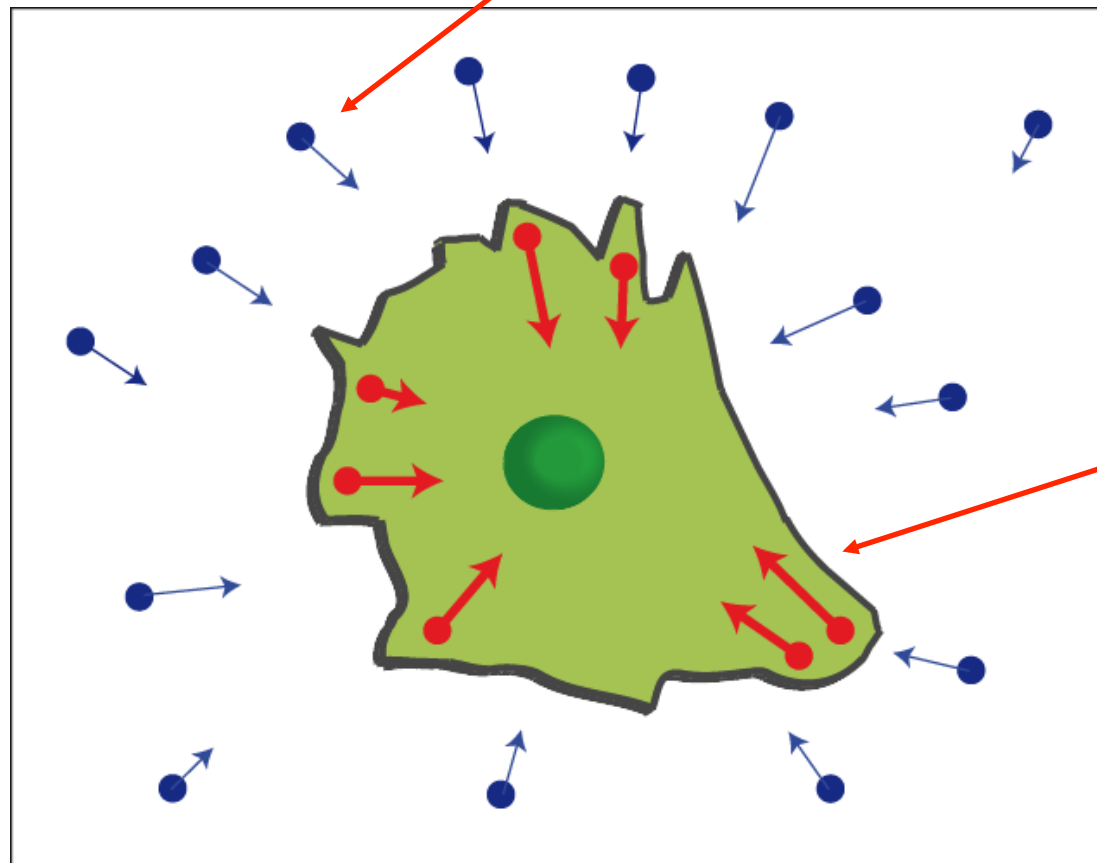


myosin II

Traction force microscopy

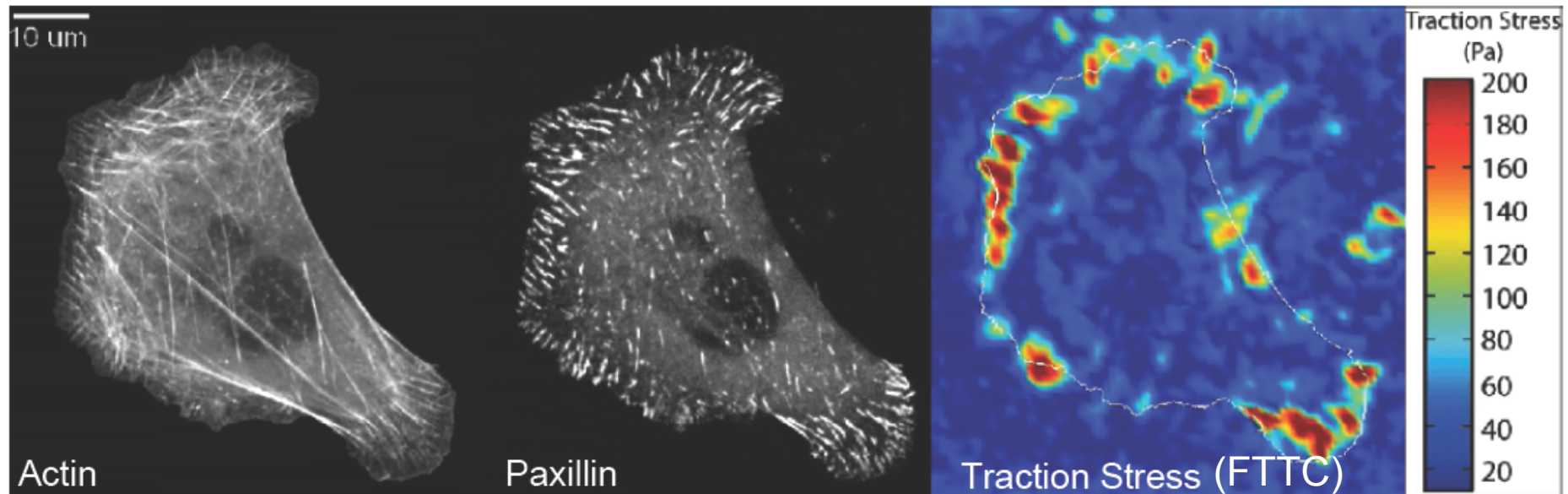
Traction force microscopy on soft elastic substrates

Pick up deformations by extracting movement of fiducial markers with image processing



Reconstruct cellular forces by solving the inverse problem of elasticity theory

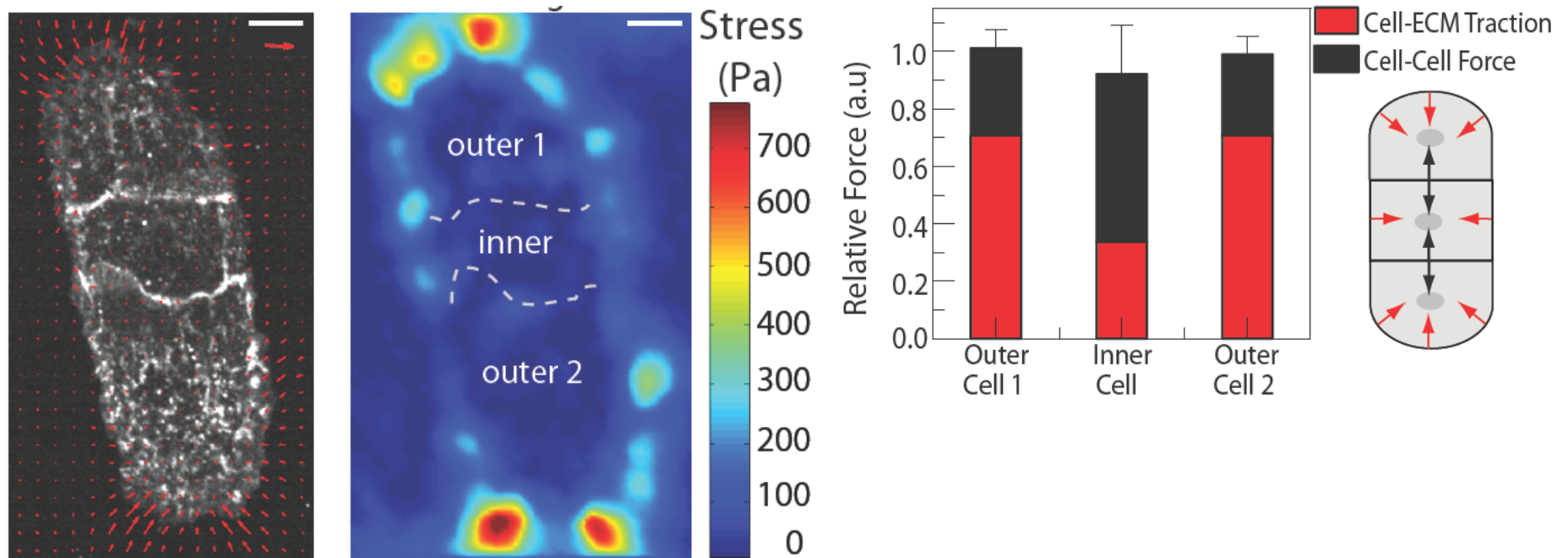
Traction map single U2OS cell



Traction forces correlate strongly with the organization of the actin cytoskeleton and the adhesion structure

[Sabass et al. BPJ 2008, Stricker et al. JPCM 2010, Schwarz and Gardel JCS 2012]

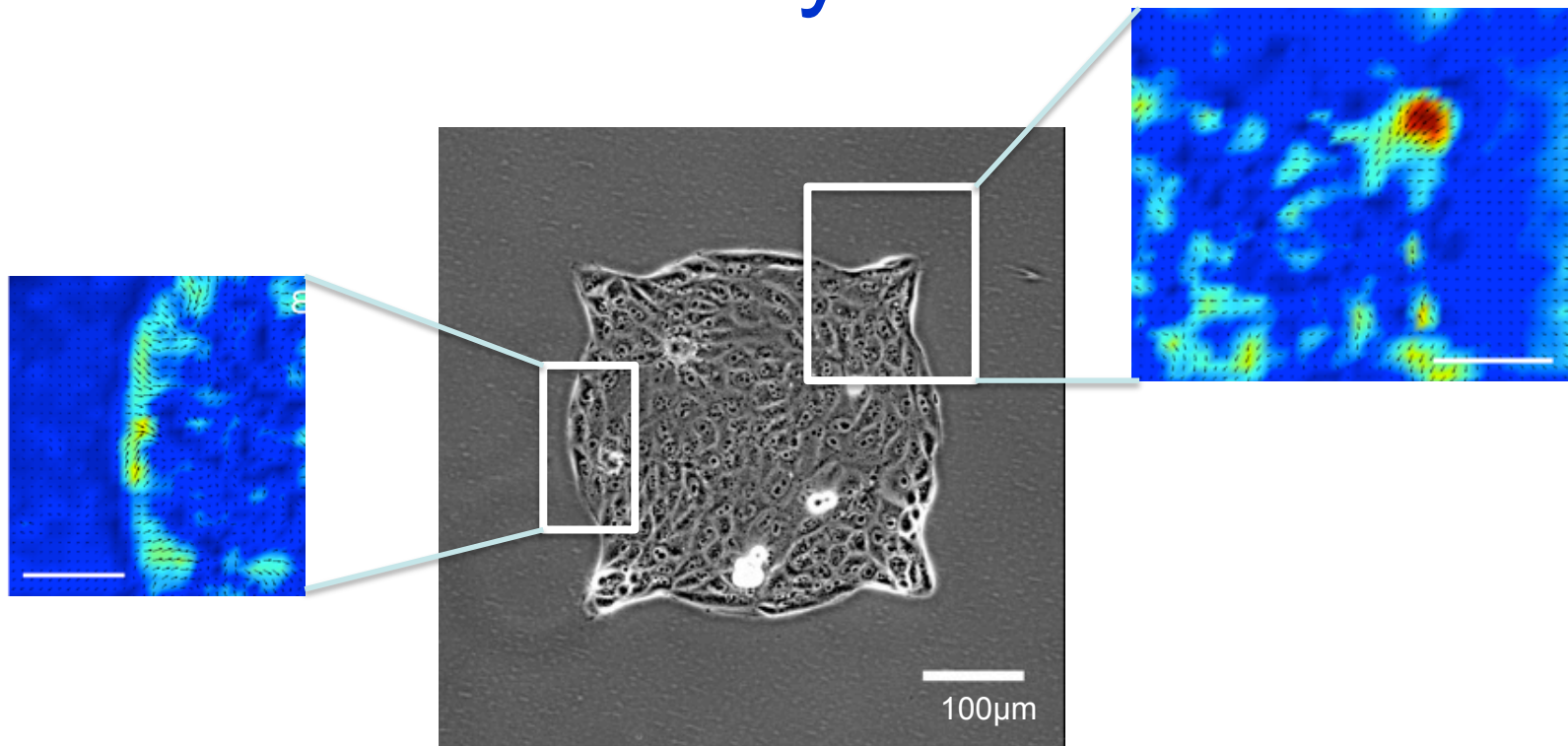
Traction force imbalance method



Cell-cell forces can be calculated from force balance.

[Maruthamuthu et al., PNAS 2011]

Traction forces of patterned cell monolayers



Geometry determines stress distribution
Leader cells emerge from traction hotspots

[Rausch et al., Biointerphases 2014]

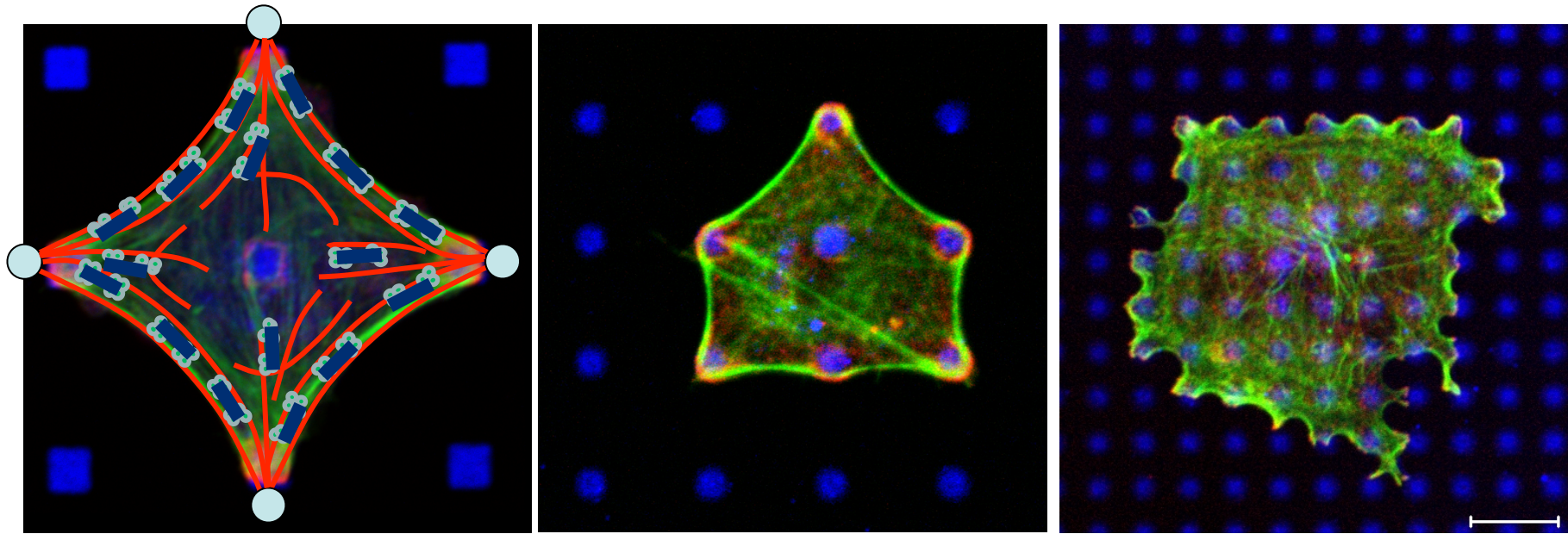
Modelling cell shape and forces

Different modelling approaches to contractile matter

- Mechanical (polymer) networks
- Active cable networks
- Contour models
- Active gels
- Active elasticity
- Cellular Potts Model
- Phase field or level set models

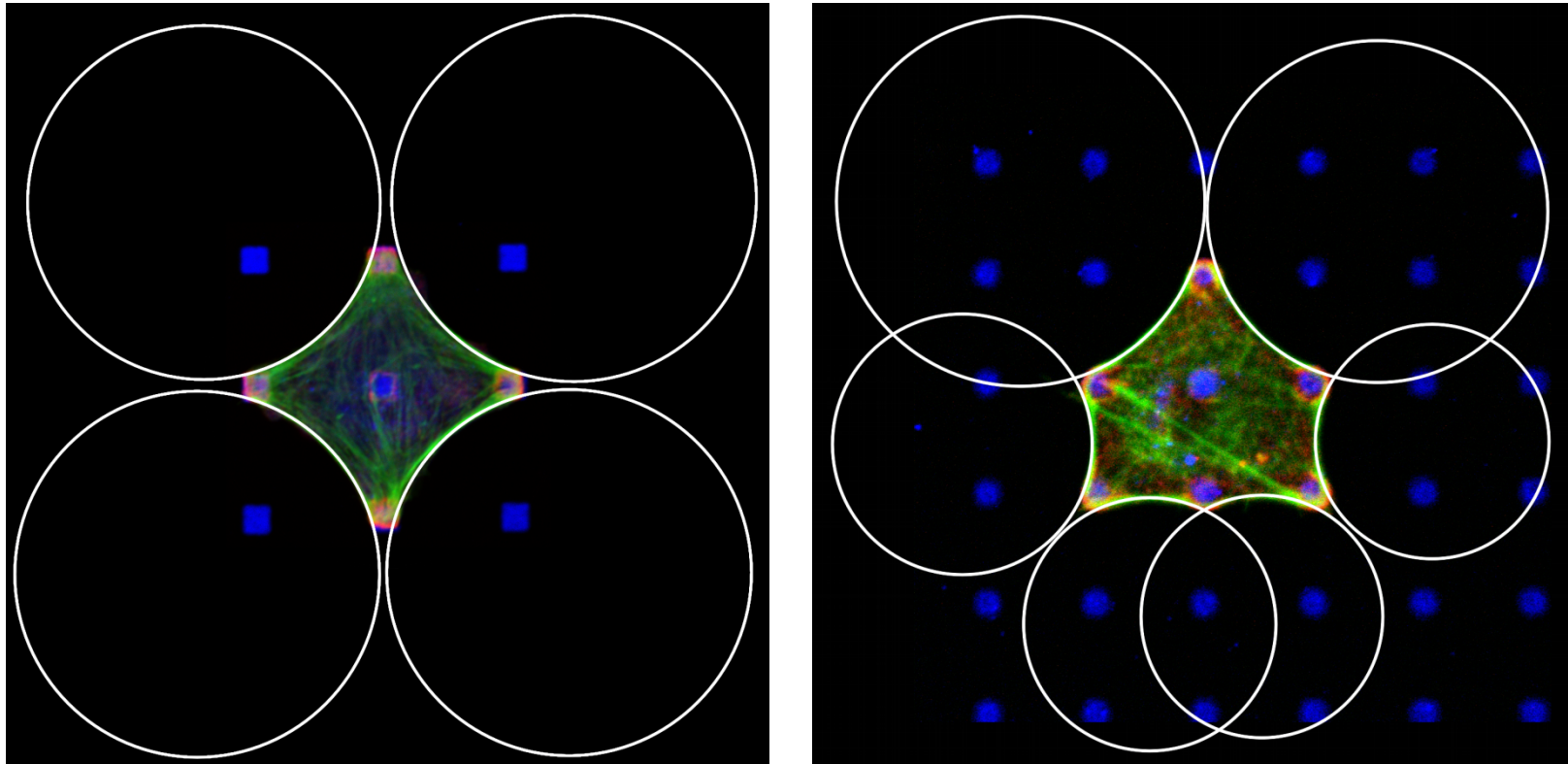
Active cable networks

Cell shape on micropatterns



[Lehnert et al. JCS 2003]

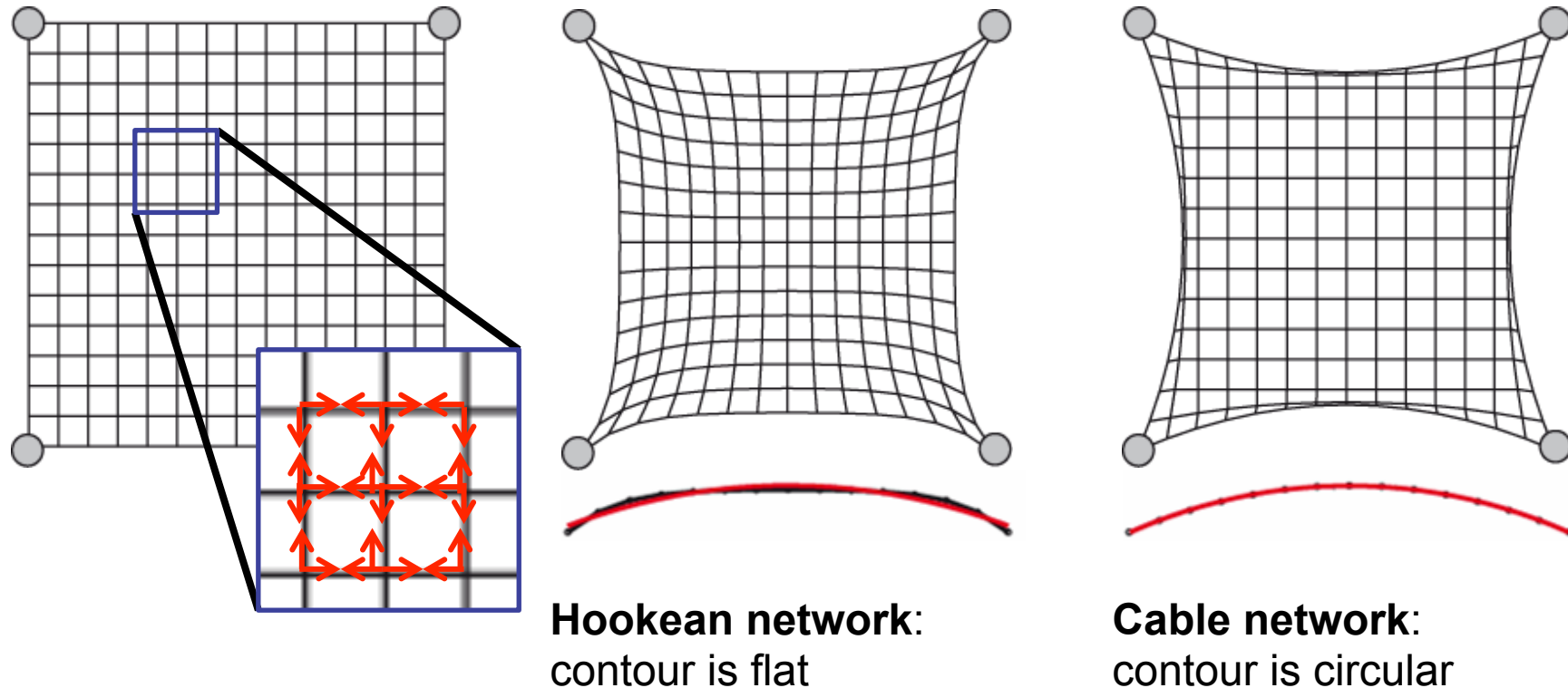
Quantitative analysis of cell shape



Cell shape on spatially constrained ligand patches resembles a sequence of circular arcs.

[Bischofs et al. BPJ 2008]

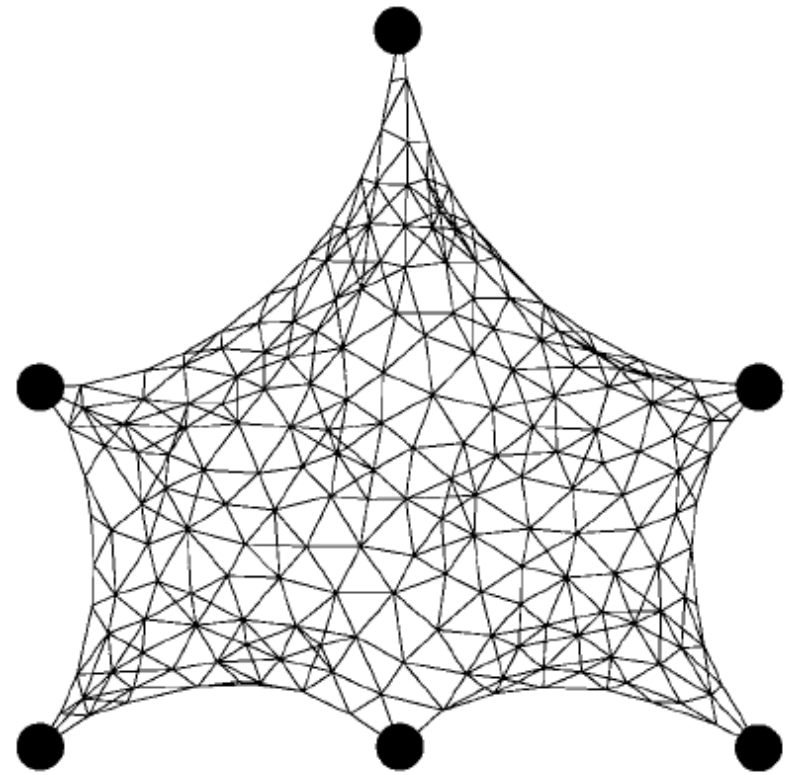
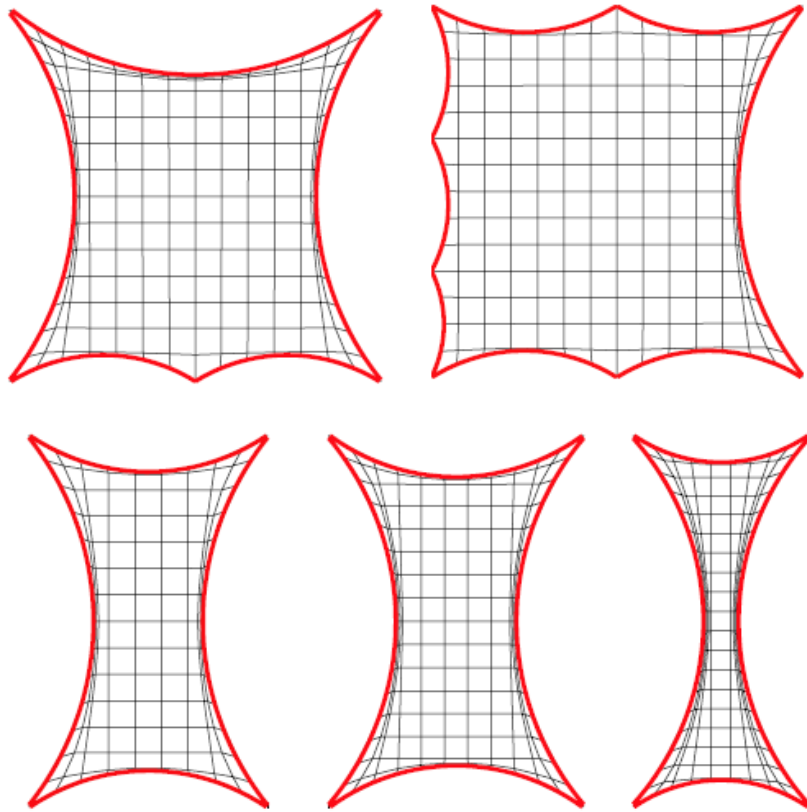
Actively contracting cable networks



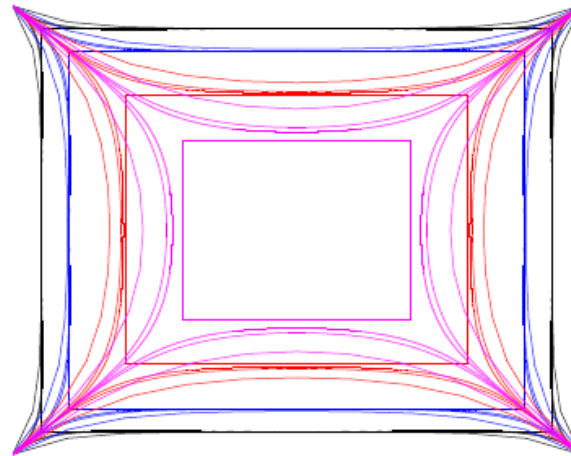
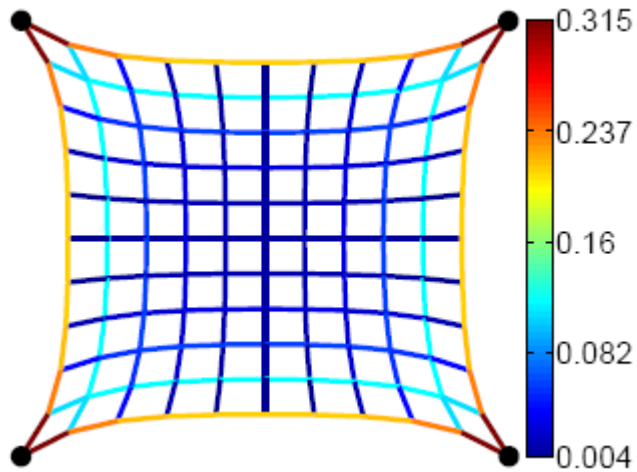
Actively contracting cable networks result in
circular arc morphology

[Bischofs et al. BPJ 2008, Guthardt Torres et al. PRE 2012]

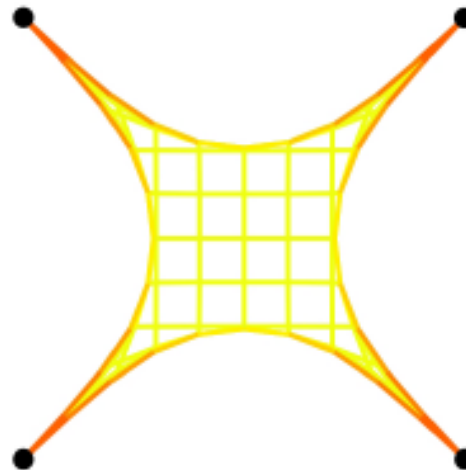
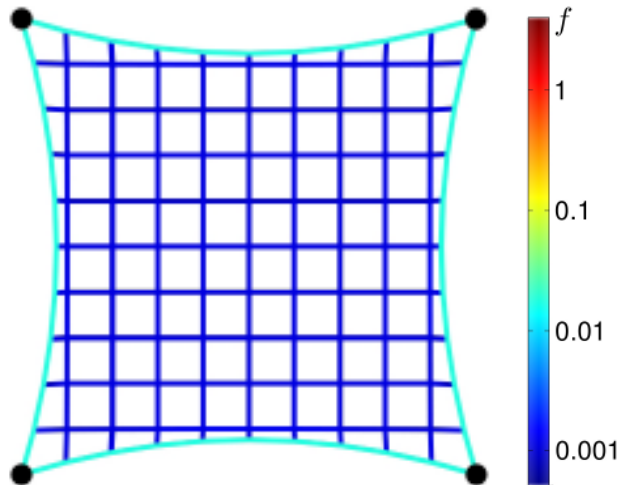
Robustness of arc feature in regard to adhesion geometry and network topology



Spring versus cable networks



Flat contour results from reference shape.



Active cable networks do not have a reference state.

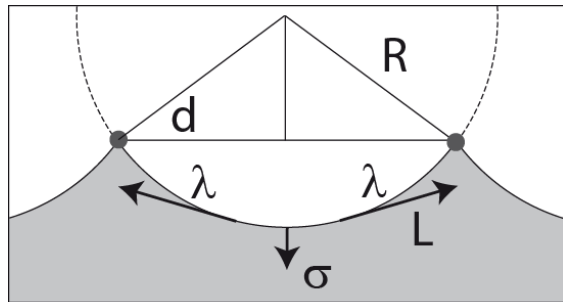
[Guthardt Torres et al. PRE 2012]

Contour model

Tension-elasticity model

Line tension results from elastic deformation:

$$\lambda = EA \frac{L - L_0}{L_0}, \quad L_0 = \alpha d$$



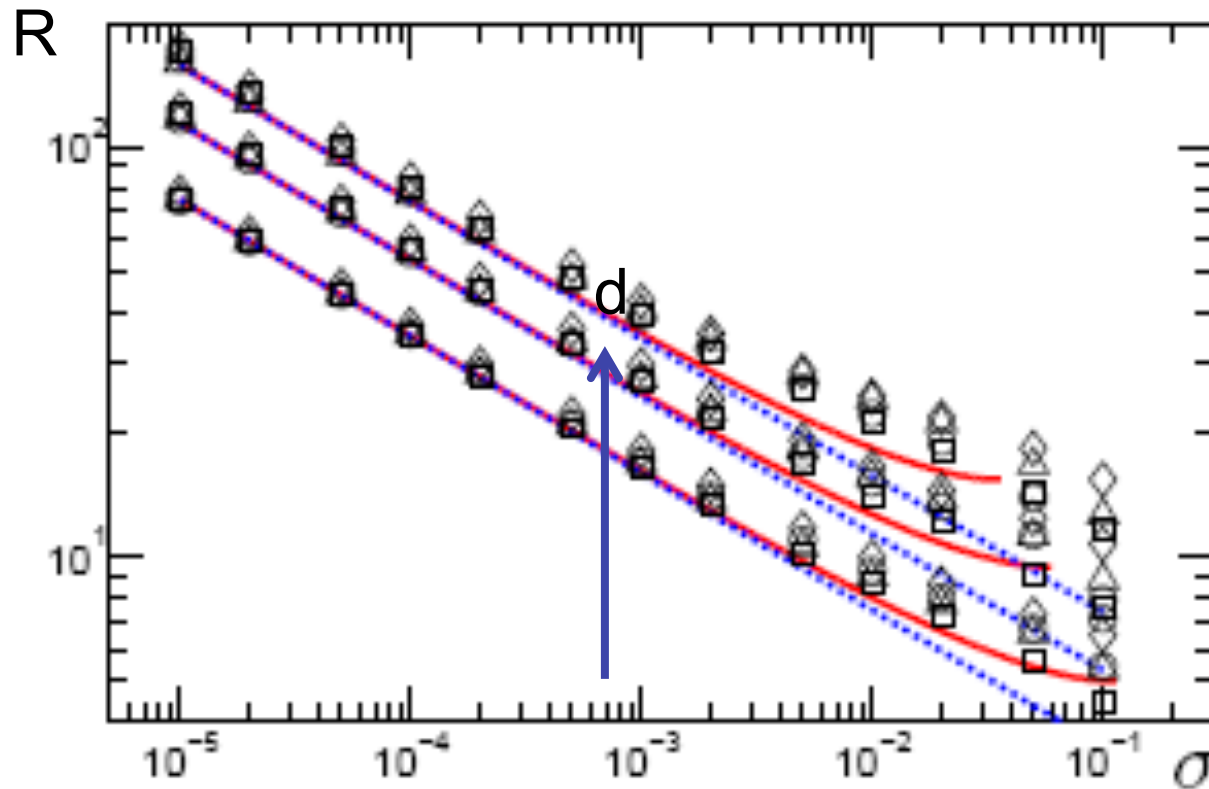
geometry:
$$R = \frac{d}{2 \sin\left(\frac{L}{2R}\right)}$$

$$\Rightarrow R = \frac{EA}{\sigma} \left(\frac{2R}{\alpha d} \arcsin\left(\frac{d}{2R}\right) - 1 \right)$$

self-consistent equation for $R(d, \sigma)$

[Bischofs et al. BPJ 2008, PRL 2009]

Modified Laplace law



Theoretical prediction by TEM-model:

$$R = 24^{-1/3} d^{2/3} \sigma^{-1/3}$$

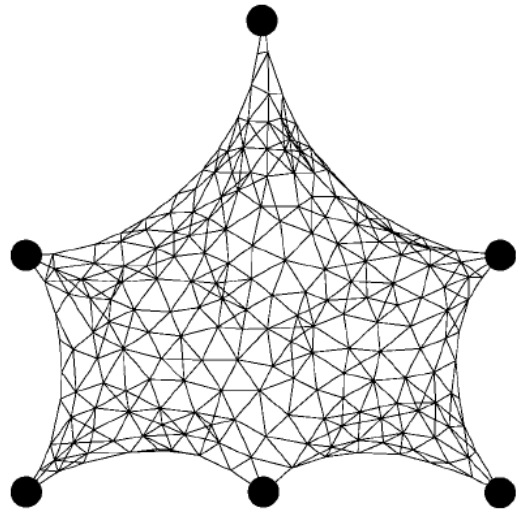
Good agreement between computer simulations, TEM-model and exps.

symbols – network simulations | solid – TEM numerical | dotted – TEM analytical

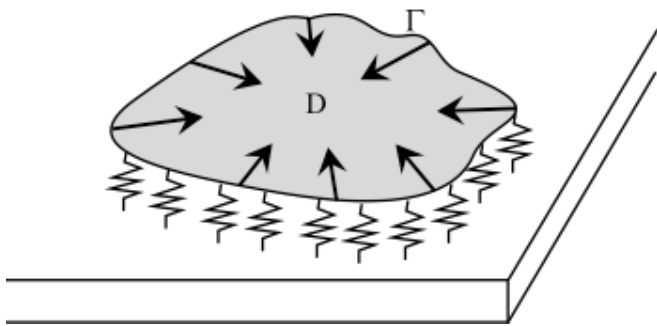
[Guthardt Torres et al. PRE 2012]

Active elasticity

Coupling cellular contractility to cell area



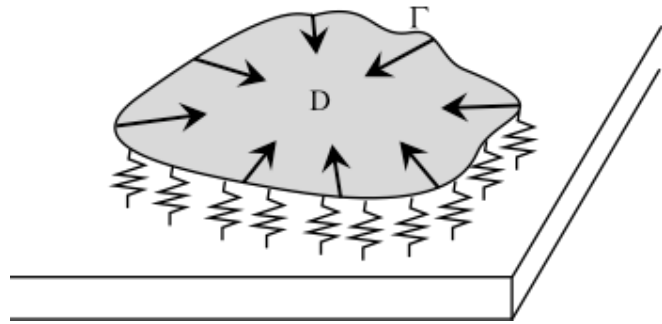
Until now: coupling to point-like adhesions



Now: coupling to adhesion area

A minimal model has to start from continuum mechanics

Simple model: contracting film on elastic foundation



Linear elasticity theory (plane stress):

$$\nabla \cdot \mathbf{F} - T = 0$$

$$\mathbf{F} = \underbrace{\frac{E_C h}{1 + \nu} \left(\epsilon_{ij} + \frac{\nu}{1 - \nu} \epsilon_{ij} \delta_{ij} \right)}_{\text{Elastic stress}} - \underbrace{\frac{1}{2(1 - \nu)} P(x, y) \delta_{ij}}_{\text{Active cell contraction}}$$

Simplest assumption for traction force: $T=ku$

Plus appropriate zero-stress boundary conditions.

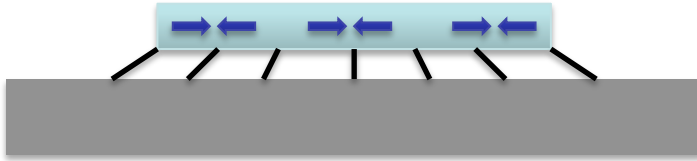
Note contraction analogy to thermal heating/cooling. Model can be implemented with FEM.

[Edwards and Schwarz PRL 2011]

[Banjeree and Marchetti: EPL 2011, PRL 2012]

One-dimensional case (contracting stripe)

force balance: F internal stress, u deformation



$$F(x) - F(x+dx) - ku(x+dx/2) = 0 \Rightarrow \frac{dF}{dx} = ku$$

constitutive relation (E 1d modulus, P active stress):

$$F = E \left(\frac{du}{dx} + P \right)$$

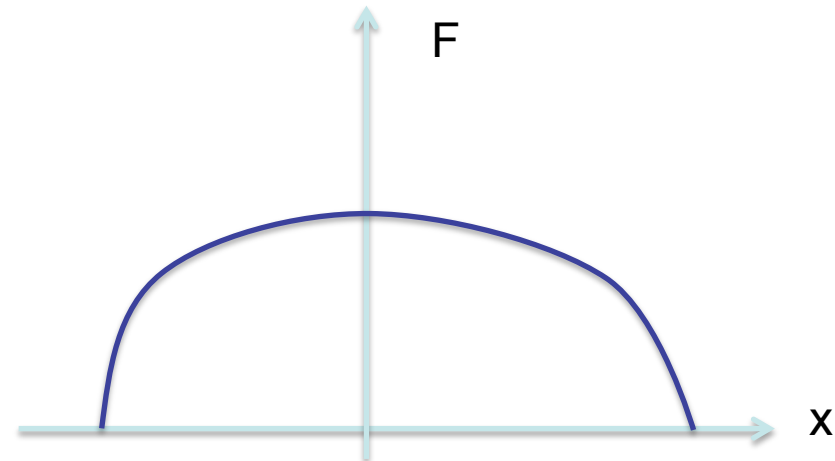
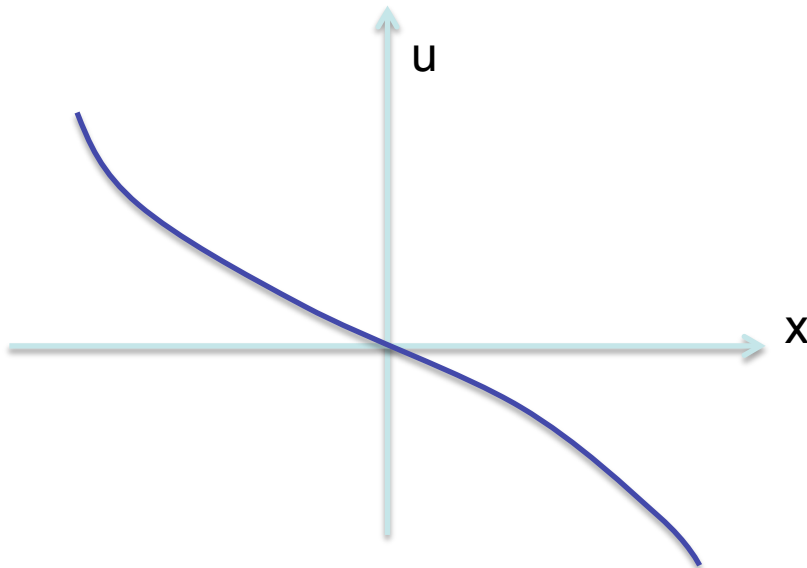
$$P = \text{const} \Rightarrow \frac{d^2u}{dx^2} - \frac{1}{l^2}u = 0$$

$$l = \sqrt{E/k} \quad \text{localization length}$$

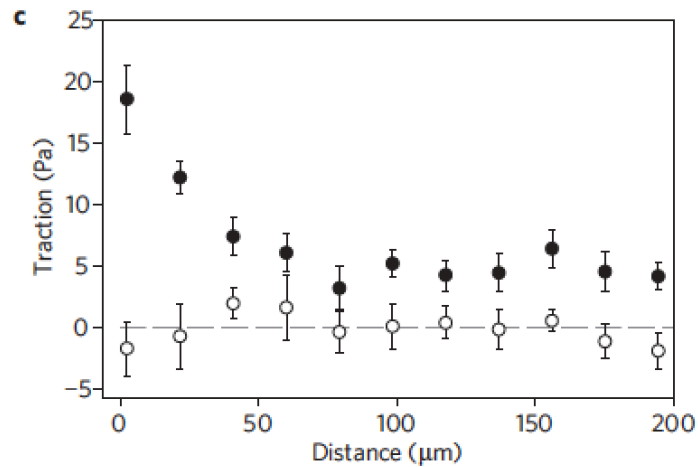
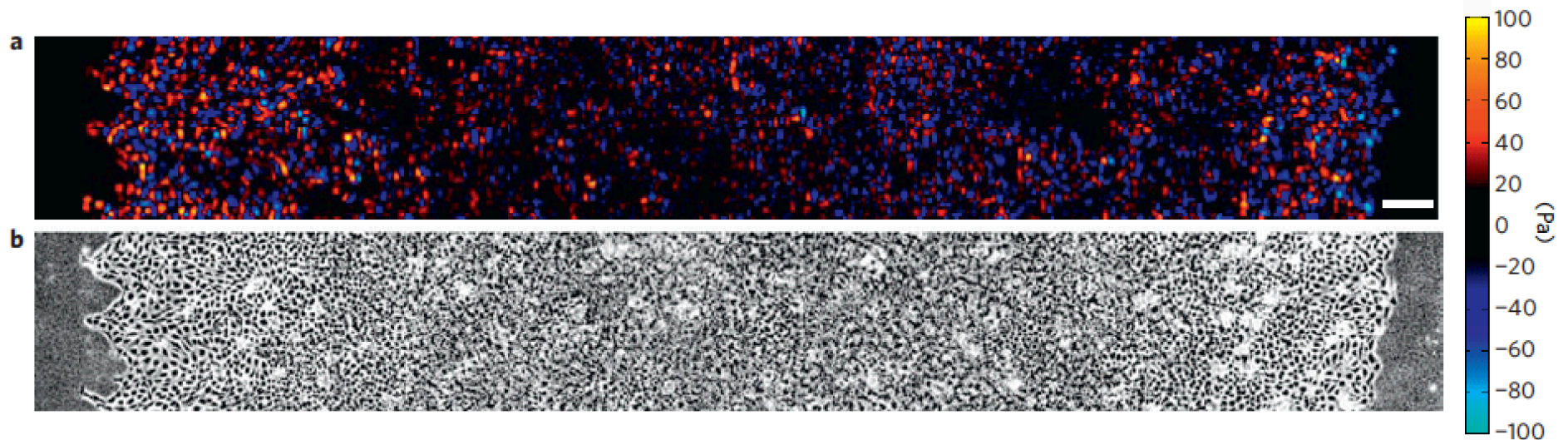
zero stress boundary:

$$F(\pm l_0) = 0 \Rightarrow \frac{u}{l_0} = -P \frac{\sinh(\gamma x / l_0)}{\gamma \cosh(\gamma)}$$

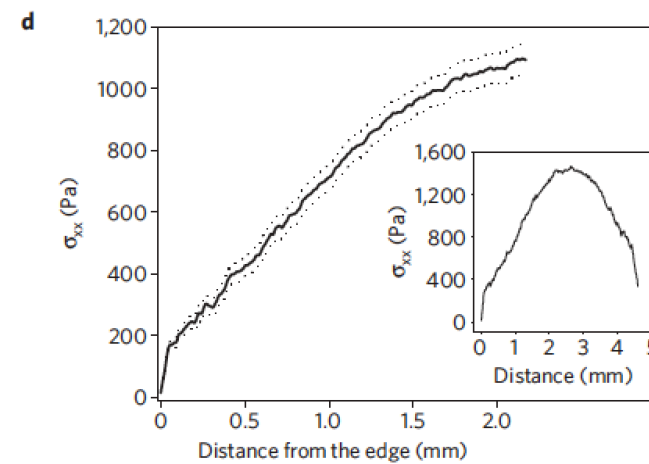
$\gamma = l_0/l$
localization parameter



Traction and stress for cell monolayer



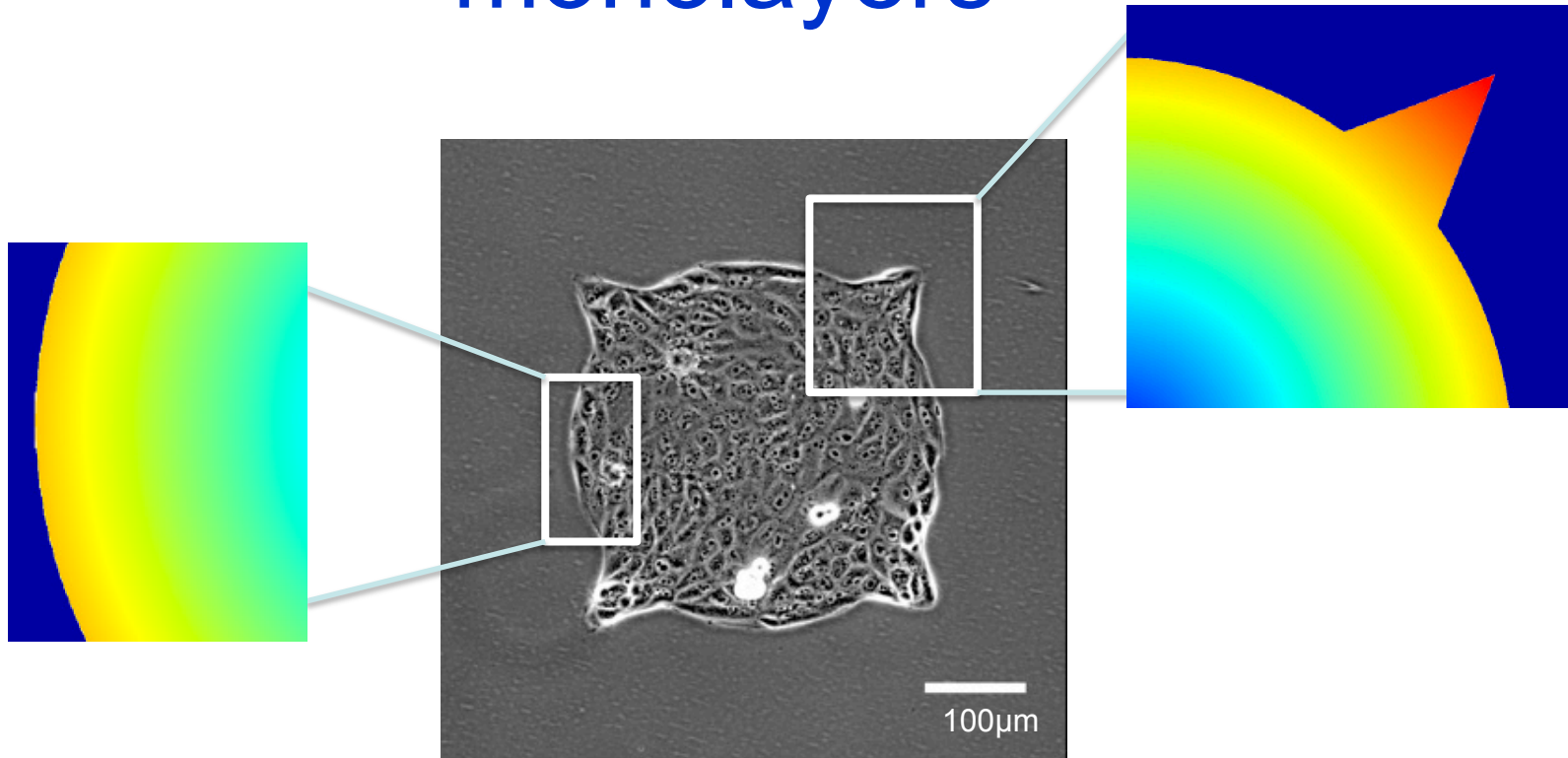
Traction is strongest at the rim



Intralayer stress peaks at the middle

[Treat et al. Nature Physics 2009]

Traction forces of patterned cell monolayers



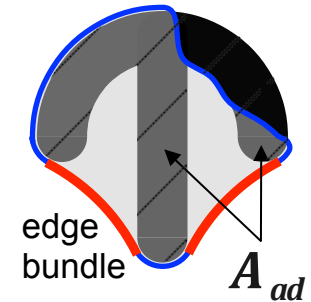
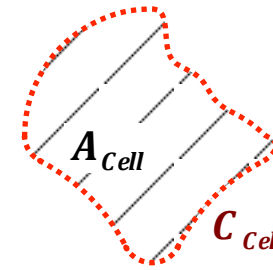
Geometry determines stress distribution
as predicted by active elasticity

[Rausch et al., Biointerphases 2014]

Cellular Potts Model

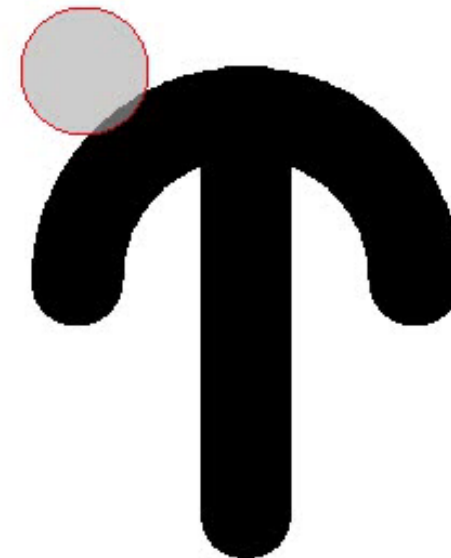
Cellular Potts Model

0	0	0	0	0
0	0	1	0	0
0	1	1	0	0
0	1	1	1	0
0	0	1	1	0



$$E = \underbrace{\sigma A_{cell}}_{\text{surface tension}} + \underbrace{J C_{cell}}_{\text{simple line tension}} + \underbrace{\sum_{\text{bundel } i} \frac{EA}{2L_{0,i}} (L_i - L_{0,i})^2}_{\text{elastic line tension}} - \underbrace{\frac{W}{A_{ref} + A_{Ad}} A_{Ad}}_{\text{adhesive energy}}$$

- Cell is represented by an ensemble of spins on a lattice
- Metropolis dynamics
- Combines elements of contour model and active elasticity continuum model



Traction forces from CPM

- Contractile forces are balanced by adhesive substrate

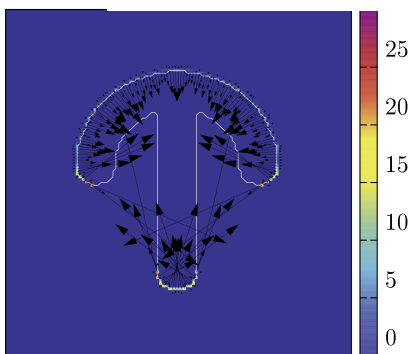
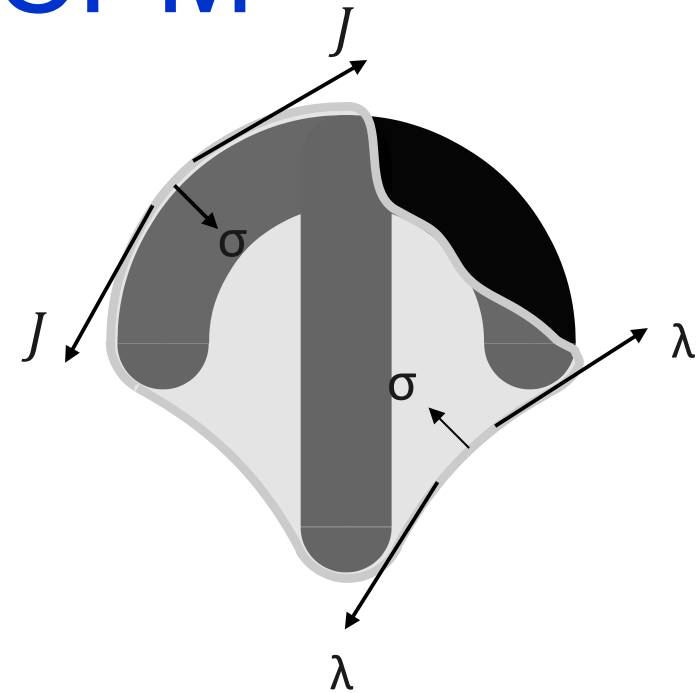
- At adhesive boundaries

$$\vec{f} = [\sigma + J \kappa] \vec{n}$$

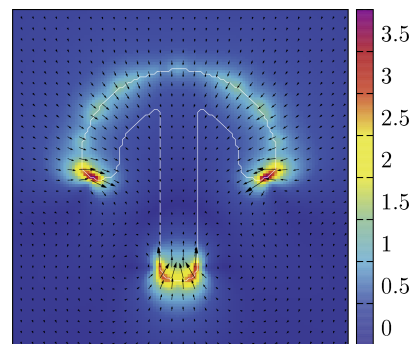
\vec{n} normal
 κ curvature

- At endpoints of edge bundles

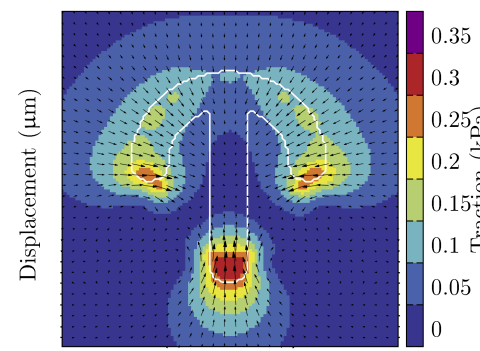
$$\vec{F}_{arc} = \lambda e \vec{t}, \quad \vec{t} \text{ tangent}$$



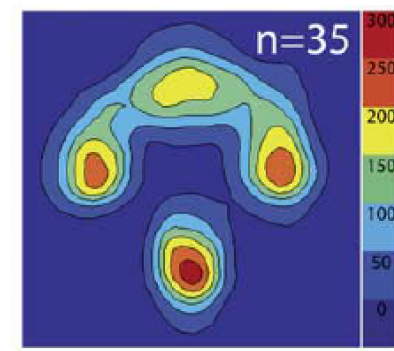
Boundary Forces



Displacements



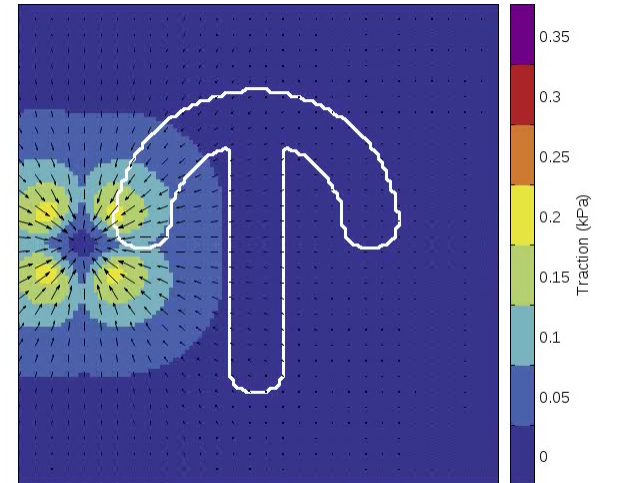
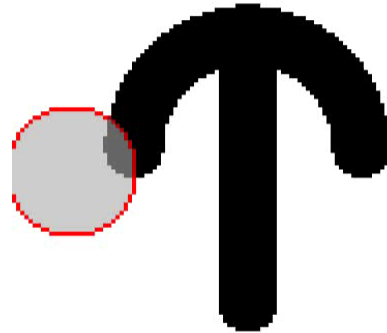
Reconstructed forces



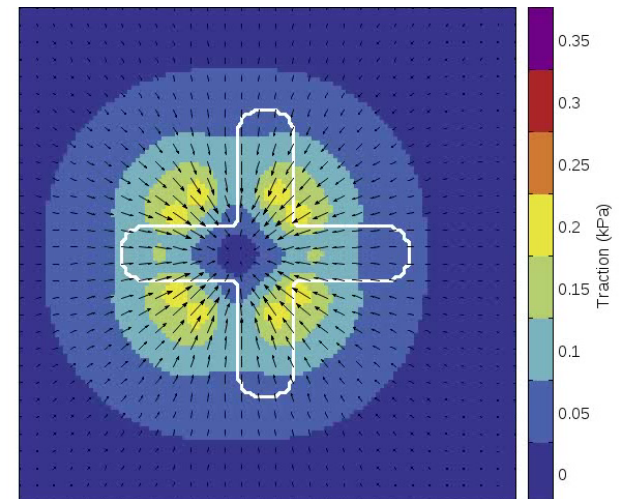
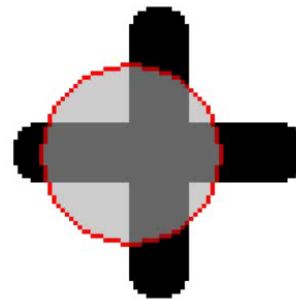
Experiments
(Tseng Lab Chip 2011)

Dynamic traction on patterns

crossbow



cross



Conclusions

Active cable networks	Circular arc morphology, modified Laplace law, represents polymer nature of cytoskeleton and constant pull by motors
Contour model	Represents bulk and contour tensions, easy to use, appropriate for strong contraction and point-like adhesions
Active elasticity	Continuum model, appropriate for coupling to adhesion area, can be implemented with FEM
Cellular Potts model	Dynamical, can be adapted to include elements of all the above models, includes the cell protrusion missing from the other models

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- **Philipp Albert** (cellular Potts model)

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