## **Cell and tissue mechanics**

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## **Cells in motion**

#### **Bovine Sperm**



#### **Breast tumor cell**







## Satellite image of the earth at night



Population distribution is governed by the environment (temperature, proximity to water, landscape, ...) and communications.

# Problem of interest: Cancer metastasis - the leading cause of death of most cancer types



#### Cancer metastasis is a physical process where tumor. cells need to generate enough Cancer microenvironment critically regulates tumor cell invasiveness force to invade

**Ref:** Huang et al. Lab Chip, 2017; Wu and Swartz, J. Biomech. Eng, 2013; Kim and Wu, Ann Biomed Eng, 2012.

# Key biophysical and biochemical parameters that drive cell migration



- Diao et al. Lab Chip, 2006,
- Cheng et al, Lab Chip, 2007,
- Haessler et al. Biomedical Microdevice, 2009.
- Haessler et al. PNAS, 2011.
- Kim et al. PlosOne 2013
- Geum et al. Euro. Phys. Journal, 2016.
- Huang et al. Lab Chip, 2017.
- Kim et al. Integrative Biology, 2020.

### **Mechanical stress**



- Rong et al. Biophysical Journal, 2011
- Hall et al. Biophysical Journal, 2012
- Hall et al. Experimental cell research, 2013
- Hall et al. PNAS, 2016.
- Huang et al. Integrative Biology, 2017.
- Suh et al. Integrative Biology, 2019

## **OUTLINE:**

## Single cell mechanics within a 3D biomatrix

- Single cell migration
- Mechanical driver
- Chemical gradient driver

## **Tumor spheroid invasion**

- Tumor spheroid formation
- Tumor spheroid invasion (chemical and mechanical driver)

## Single cell migration in 3D



Integrin

Cells are supported by a 3D fiber network Differ from <u>2D</u> cell migration

Cell-ECM tensional balance is regulated by cell traction force.



Wu and Swartz, J. Biomech. Eng. 2014.

### Cell migration in two dimensional space (2D) versus 3D



## Many cell types requires the 3D environment to exhibit physiologically realistic phenotypes

Figure adapted from: Wu and Swartz, Journal of biomechanical engineering, 2014.

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## Amoeboid and mesenchymal cell migration

#### **Amoeboid cells**



i	
Amoeboid	Mesenchymal
Round	Elongated
Short-live	Long-live
adhesion	Adhesion
MMP	MMP
independent	dependent
Path	Path
finding	generating

#### **Mesenchymal cells**









[1] Turner et. al. (2011). [2] Paňková, K., et al. (2010).
[3] Sabeh, F., et al. (2009). [4] Pathak et al. (2011) Integr Biol

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## **Cancer cell chemotaxis in 3D microfluidic model**

In collaboration with Prof. Melody Swartz at Swiss Institute of Technology, now University of Chicago



## **Roles of lymph node microenvironment in cancer cell migration**

#### **Clinical wisdom:**

Lymph nodes -first stops of metastatic cancer cells of many cancer types

# Cancer cell types correlated with lymph nodes metastasis

- Breast (Cabioglu et al. 2005)
- Melanoma(Taakeuchi et al., 2004)
- Colorectal (Gunther et al. 2005)
- Head and neck (Wang et al. 2005)
- Prostate (Heresi et al. 2005)
- Non-small lung (Takanami, 2003)
- Gastric (Mashino et al. 2002)

Ref: Shields et al. Cancer Research, 2007



#### Gene profiling (breast cancer cells):

Chemokine receptors are implicated in metastatic breast cancer cells

Hypotheses:

Breast tumor cells are chemotactic in SDF-1 $\alpha$  (ligand to CXCR4) and CCL19 (ligand to CCR7) and gradients.

# Creating chemokine gradients using a hydrogel-based microfluidic platform



Diao et al. Lab on a Chip, 2006, Cheng et al, Lab on a Chip, 2007, Haessler et al. Biomedical Microdevice, 2009. Haessler et al. PNAS, 2011. Kim et al. PlosOne 2013 Geum et al. Euro. Phys. Journal, 2016.

# 3D *in vitro* cell culture using type I collagen (derived from rat tails)

Type I collagen



#### 20µm

Reflective confocal image of collagen Fibers (Cross and Stroock, Biomaterials, 2010).



MDA-MB-231 cells embedded in collagen<sup>15</sup>

#### **Tumor cell versus immune cell chemotaxis**





#### Dendritic cells CCL19 gradient

Haessler et al. Biomedical Microdevice, 2009

Breast tumor cells (MDA-MB-231) SDF-1 $\alpha$  gradient

Tumor cells are highly heterogenic in terms of morphology and motility

# Quantifying tumor cell chemotaxis in cytokine gradients



### Both dendritic and tumor cell chemotaxis is governed by ligand receptor binding kinetics



Solid lines is a fit to:

$$V_{X} = A \frac{\nabla C}{\left(C_{Avg} + K_{D}\right)^{2}}$$

*K*<sub>D</sub> : Ligand receptor association constant A: constant

Fitted  $K_D = 59.2$  nM Reported  $K_D = 55 \pm 15$  nM\*

- Tumor cell chemo-sensitivity is governed by the receptor ligand binding kinetics.
- Fitted K<sub>D</sub> agrees well with the reported value obtained using a FRET method.
  - 1. Hassler et al. PNAS, 2011

Receptor/ligand: CXCR4/SDF-1alpha

- 2. Kim et al. PlosOne, 2013
- 3. Valenzuela-Fernandez et al, I. Biol Chem, 2001 (using FRET)

#### **Tumor cells execute Levy walks**



Levy exponent reaches ~2.0 when CCL 19 concentration is close to its kinetic constant.

Kim et al. Integrative Biology, 2020.

## What did we learn?

Tumor cell migration is governed by ligand receptor binding kinetics via either chemotaxis and/or chemokinesis.

## **Question:**

Which is more effective for reaching a distant target, chemokinesis or chemotaxis?

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### Single cell mechanics

# In collaboration with Vivek Shenoy at U Penn and Herbert Hui at Cornell





Xingzhen Feng



Farid Alisafaei

# A 3D traction force microscopy for single cell force generation in collagen gel



Breast tumor cell (MDA-MB-231) migrating within collagen matrix embedded with fluorescent beads

# Cell-matrix cross talk revealed by the deformation of surrounding matrix

Rong et al. Biophysical Journal, 2011 Hall et al. Biophysical Journal, 2012 Hall et al. Experimental cell research, 2013 Hall et al. PNAS, 2016.

## Mapping 3D fiber network deformation field



#### Bead displacement field around a breast tumor cell embedded in type I collagen

Ref: Hall, PNAS, 2016; Wang H, et al. 2014, Biophys. J. 107(11):2592-2603.

#### **Engineering collagen microstructure and mechanical properties through fibril cross linking and gel density**



#### Methods

- Gel density
- Ribose glycation
- Polymerization temperature

# A set of collagen gel representative of tumor microenvironment



Collagen matrices span the physiological range of stiffness for normal and malignant breast tissue

### Material model with fiber alignment (slide from Shenoy)

The overall elastic energy consists the isotropic and fibrous contributions ٠

$$W = W_b + W_f$$
  $W_f = \sum_{a=1}^{3} f(\lambda_a)$ 

Fibers are aligned along principle stretch ( $\lambda_a$ ) orientations  $n_a$ ٠

$$\sigma = \sigma^b + \sigma^f$$
  $\sigma^b = \kappa (J-1)I + \mu \operatorname{dev}(\overline{B})/J$ 

$$\boldsymbol{\sigma}^{f} = \frac{1}{J} \sum_{a=1}^{3} \frac{\partial f(\lambda_{a})}{\partial \lambda_{a}} \lambda_{a}(\boldsymbol{n}_{a} \otimes \boldsymbol{n}_{a})$$



Ban et al., PNAS (2019)



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# Mapping 3D gel deformation field using a 3D force microscopy and a network-inspired material model



#### Only the fiber network material model worked!

Ref: Hall, PNAS, 2016; Wang H, et al. 2014, Biophys. J. 107(11):2592-2603.

# Cancer cells exert sufficient strain to locally stiffen collagen matrices



#### Fibrous nonlinear elasticity is critical for cell- ECM interaction

### A mechanical feedback between cells and ECM



Onset for strain stiffening of the gel

Cell to ECM: Cells exert forces sufficiently to stiffen and align ECM

### A mechanical feedback loop between cells and ECM



ECM to cell:

Stiffer gel promotes larger cell force generation and stiffer cell body

# Fibrous nonlinear elasticity promotes cell force transmission distance



## What did we learn?

Nonlinear anisotropy of the material model is critical for cell function

Biological convergence Biological materials promote cell-cell communication

## **OUTLINE:**

## How does single cell move within a 3D biomatrix ?

- Single cell migration
- Chemical gradient driver
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## How does tumor spheroid invade ?

- Tumor spheroid formation
- Tumor spheroid invasion (chemical and mechanical driver)

### **Tumor spheroid formation and invasion**

In collaboration with Jeffrey Segall



## Why spheroid?



## **Making tumor spheroids**





Well diameter 200  $\mu m$ 

In collaboration with Minglin Ma and Momita Das Green: malignant MDA-MB-231 cells Red: non-tumorigenic epithelial MCF-10A cells Song *et al.* (2016) *Soft Matter* 

## **Tumor spheroids inversion**



Huang et al. Q-bio arXiv, 2020.

### What did we learn?

Cells of different types segregate. Shell – core inversion occurs due to differential growth.

## **Question?**

Can the idea of phase transition be applied here?

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# Tumor architecture critically regulates tumor invasion



Green: Metastatic cancer cells MDA-MB-231 Red: Non-tumorigenic cells MCF10A

# Tumor architecture critically regulates tumor invasion



**Green: Metastatic cancer cells MDA-MB-231 Red: Non-tumorigenic cells MCF10A** 

# Interstitial flow promotes tumor spheroid invasion via down-regulation of E-cadherin

Control



Green: malignant MDA-MB-231 cells





Red: non-tumorigenic epithelial MCF-10A cells

Huang et al. Unpublished.

### Interstitial flows enable co-culture tumor spheroid explosion



Green: malignant cells Red: non-tumorigenic cells

- Interstitial flows increase the spheroid sizes
- Greater impact on the non-tumorigenic cells.

## **Tumor spheroid chemotaxis in EGF gradients**



No EGF gradients

With EGF gradients

Suh et al, Unpublished, 2020.

## What did we learn?

Tumor architecture regulates tumor invasion. Cell-cell adhesion regulates tumor invasion.

### **Question:**

Can we predict tumor invasion knowing single cell characteristics and dynamics?

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