

Nutrient Homeostatic Networks

Outline

1. Background and mechanistic information
2. Systems level - bistability & buffering

Nutrient and Ion Homeostasis

ENVIRONMENT

nutrients, ions
dynamic, unpredictable

Nutrients, ions:

- essential for cellular processes
- toxic in excess

INTRACELLULAR

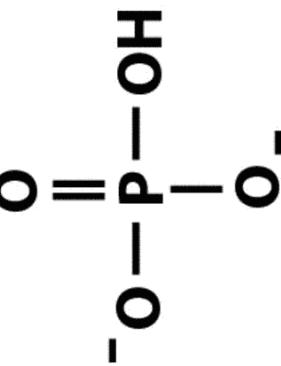
nutrients, ions
~constant



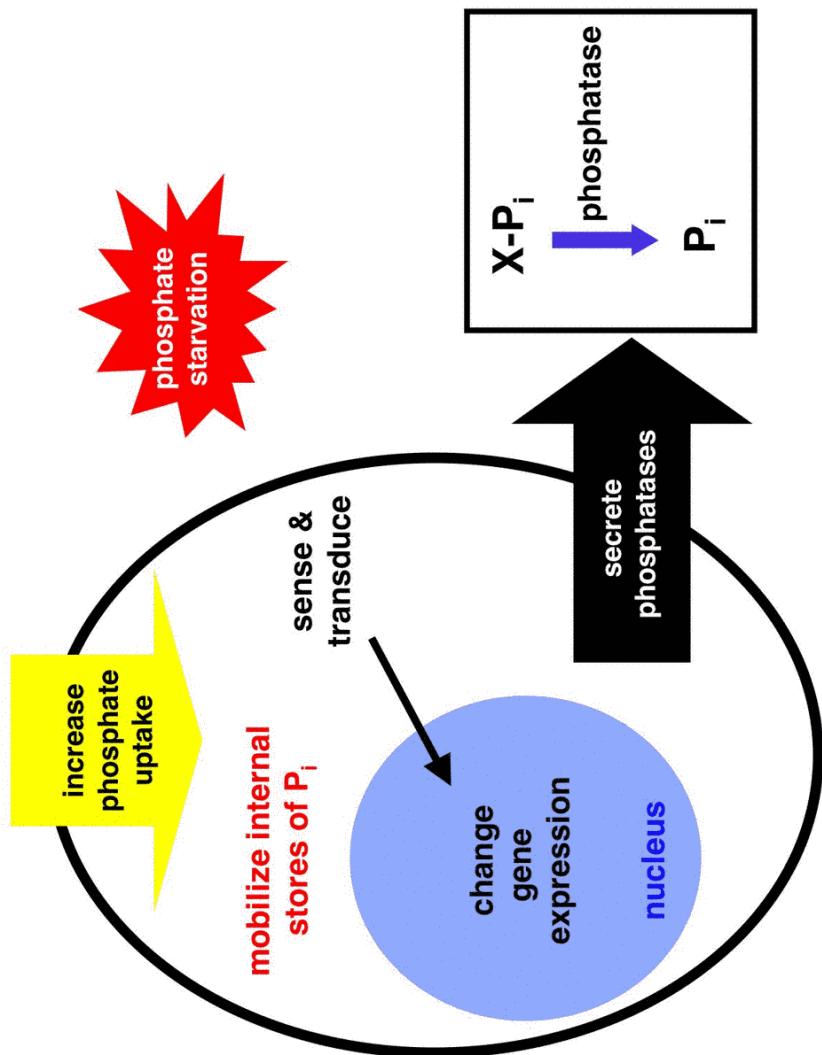
budding yeast cell

Inorganic Phosphate

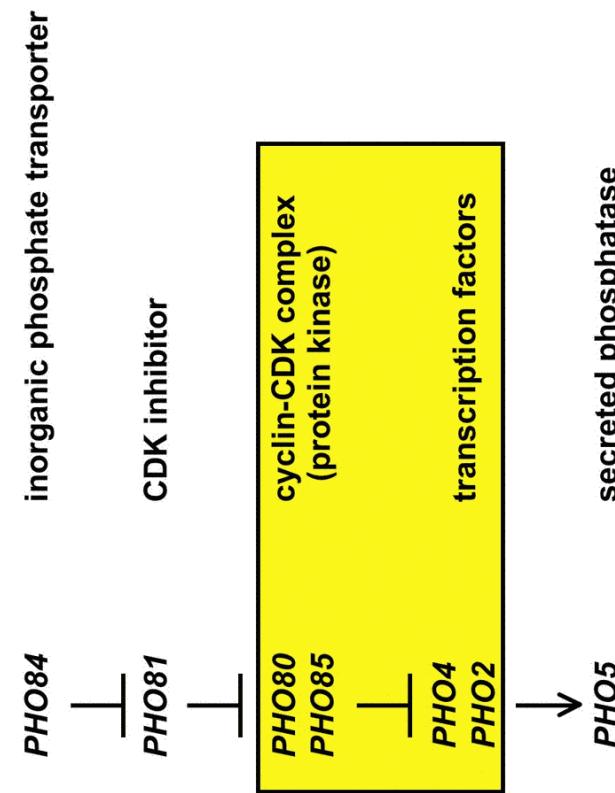
P_i



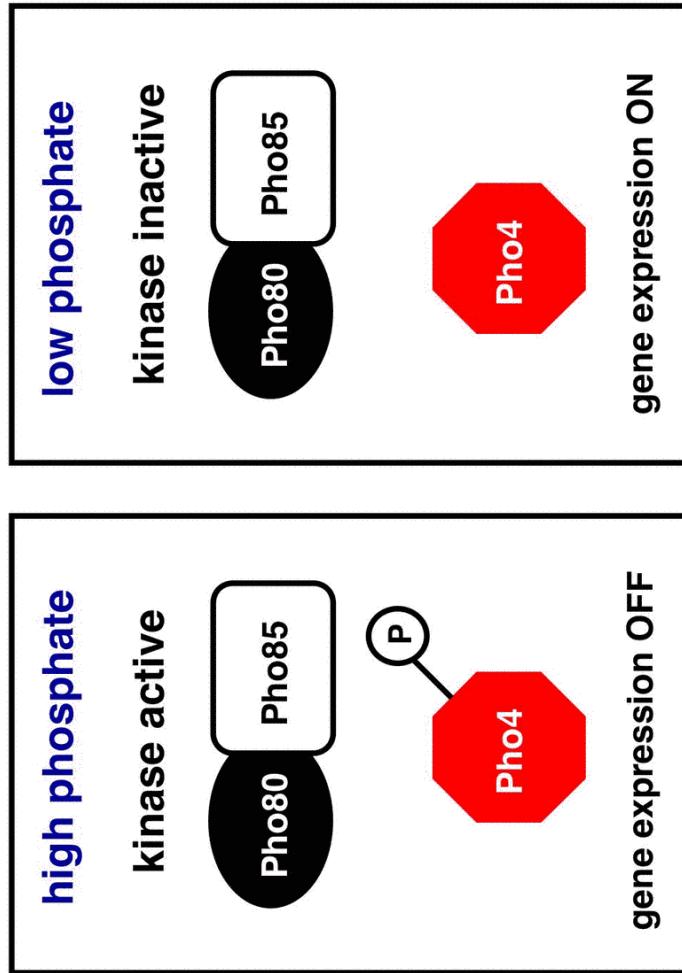
- Important for synthesis of ATP and other nucleotides, phospholipids
- Want to maintain constant levels inside cell
 - Cannot be made - must be obtained from environment



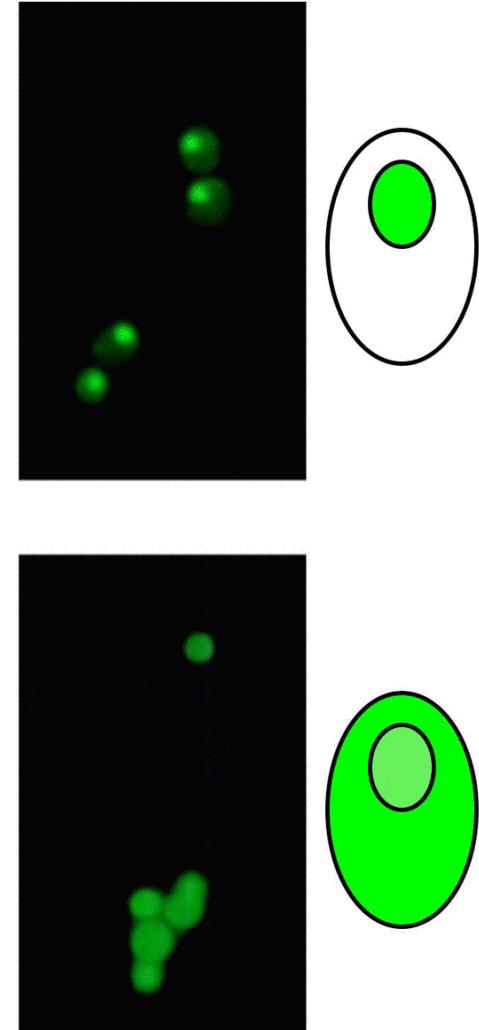
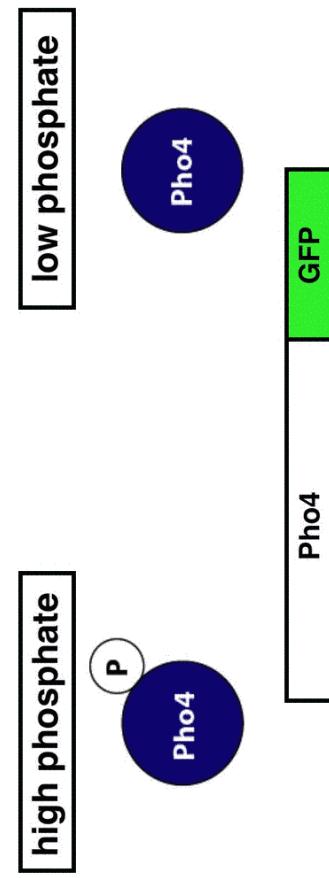
The Pho Pathway

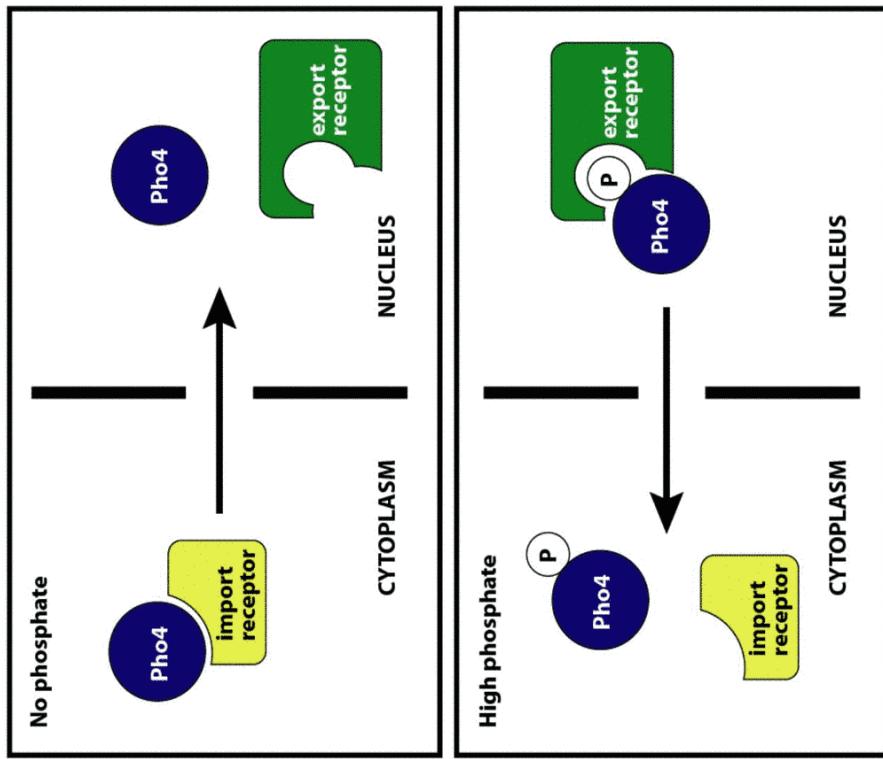
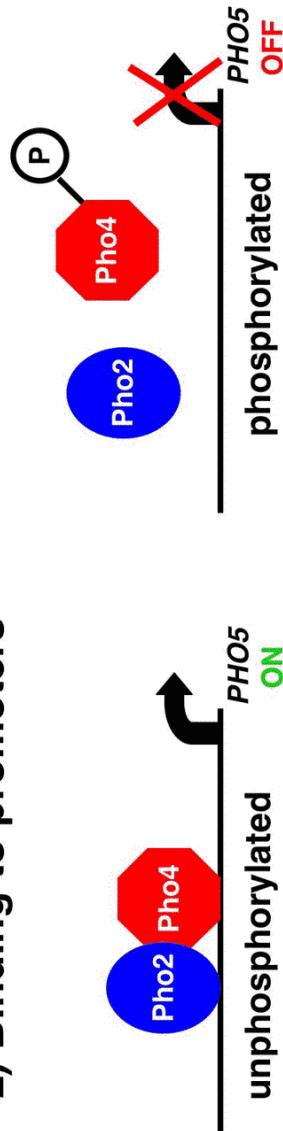


The Phosphate-Responsive Signaling Pathway

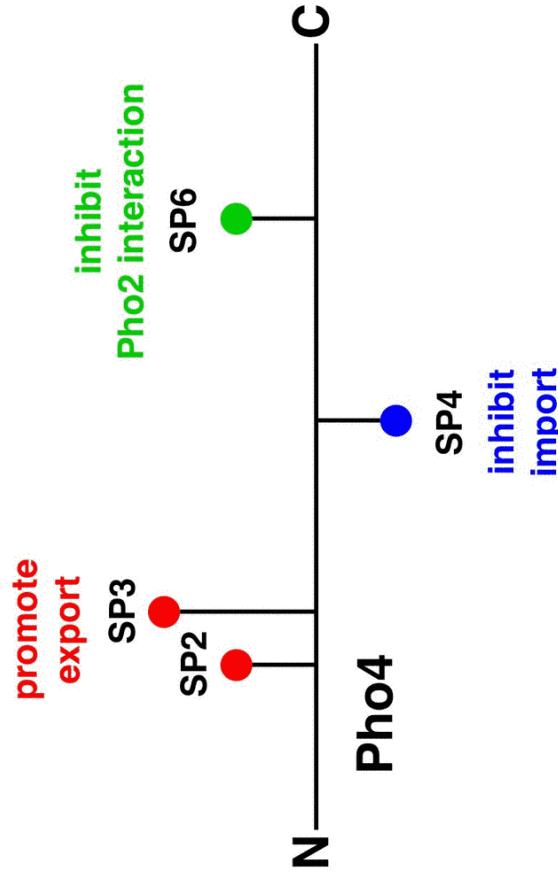


Pho4 Localization is Regulated by Phosphorylation

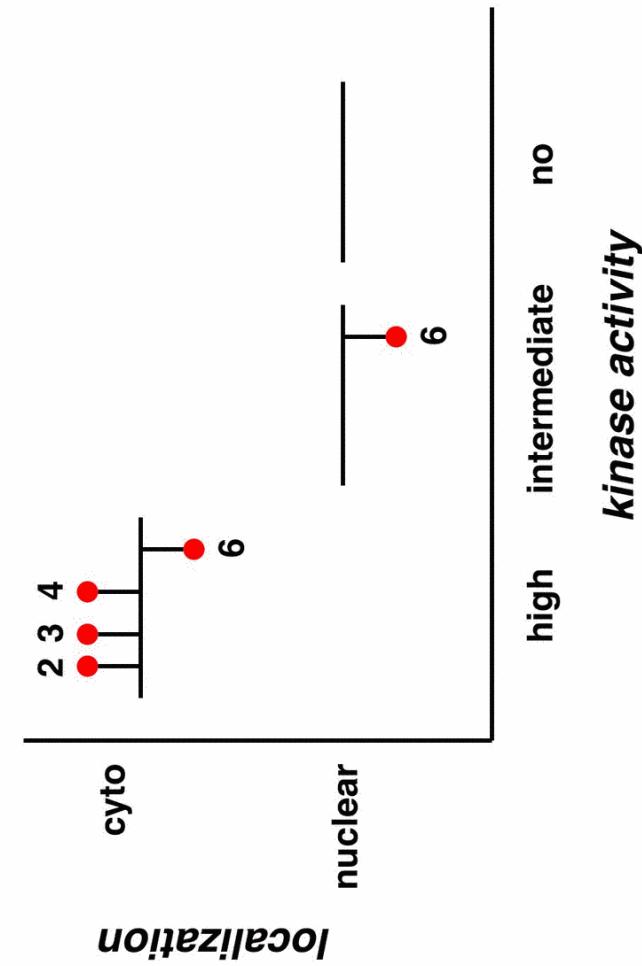


Pho4 localization**Phosphorylation Regulates Pho4 In Two Ways****1) Location within the cell (subcellular localization)****2) Binding to promoters**

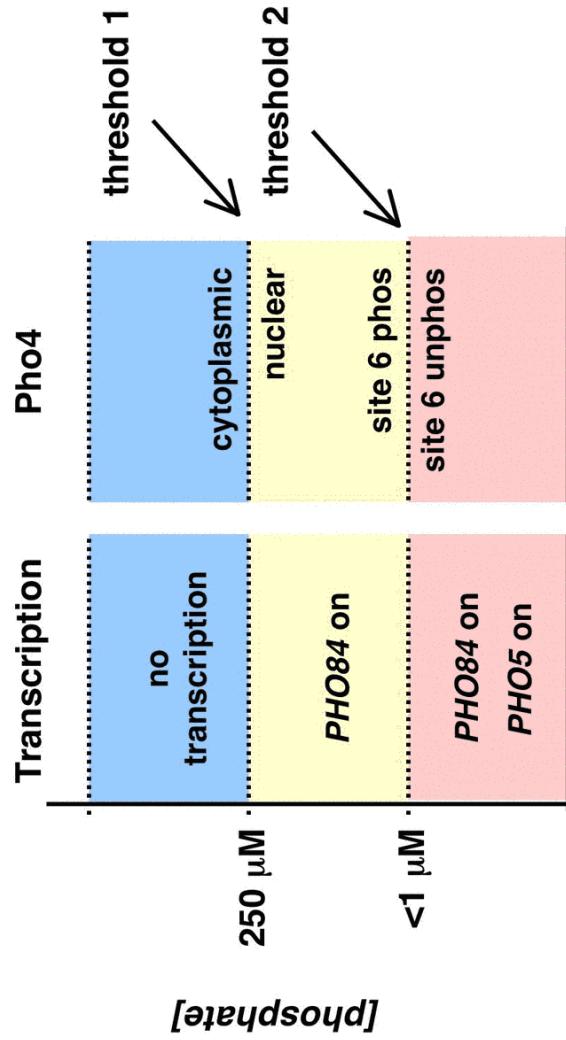
Pho4 is Multiply Phosphorylated



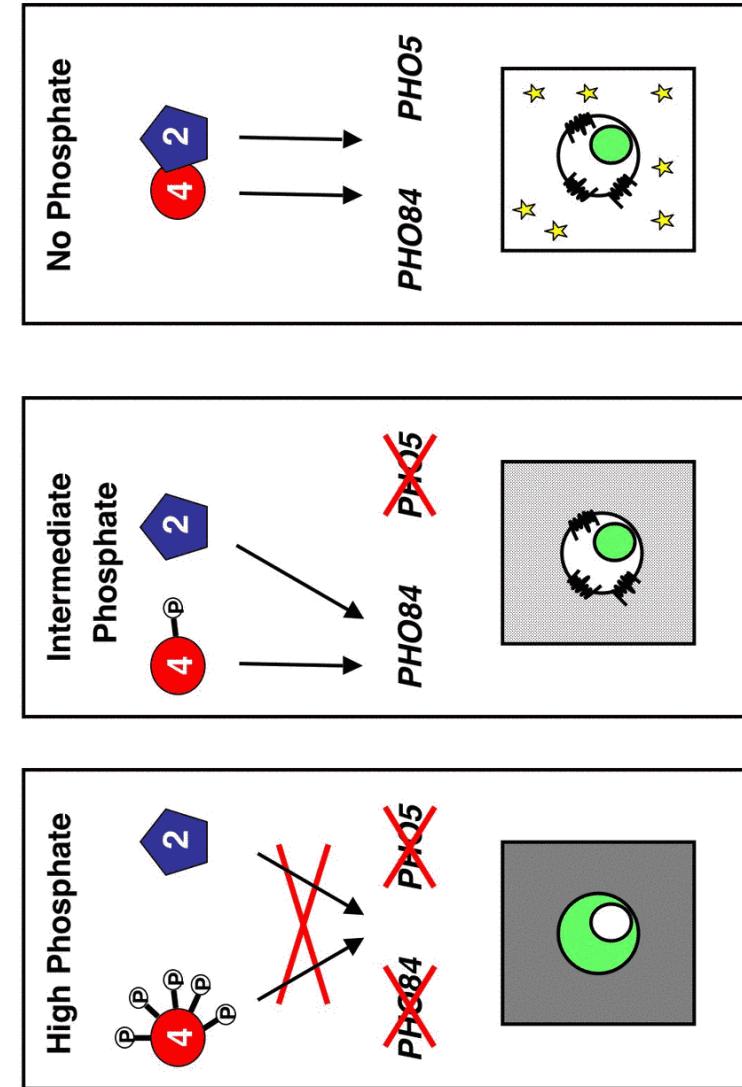
In vitro Studies Predict Accumulation of Pho4 Phosphorylated on Site 6



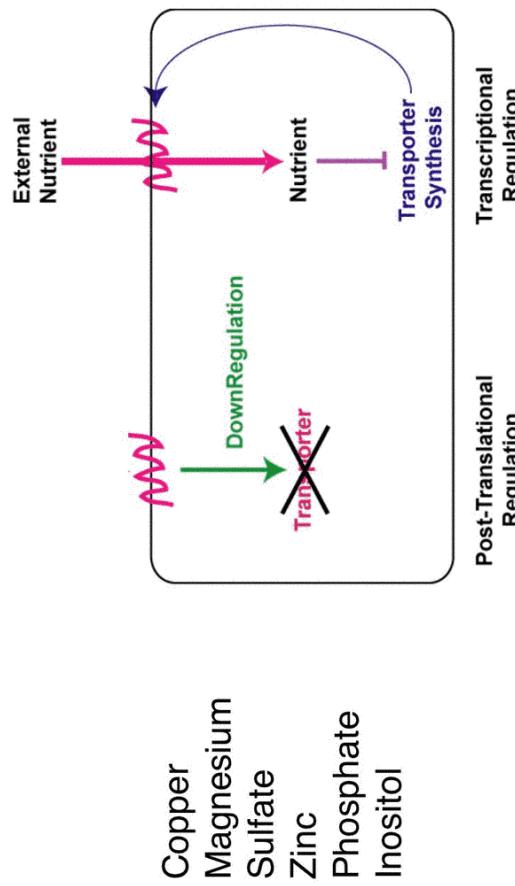
The Kinetic Properties of Pho80-Pho85 Help to Generate Two Thresholds in the *PHO* Pathway



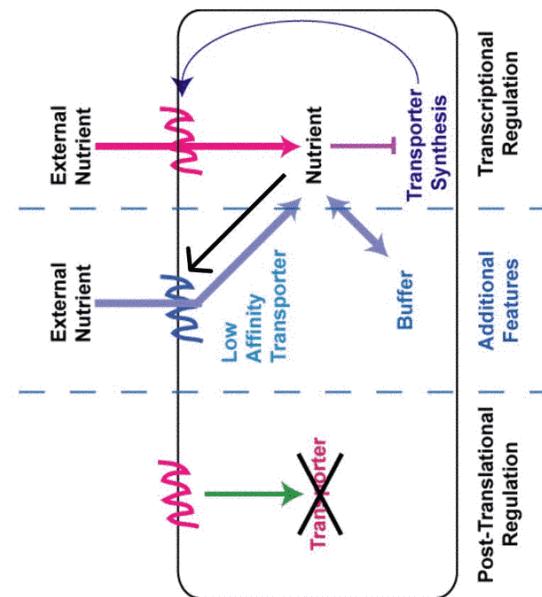
Yeast Cells Tailor Their Response to Environmental Conditions



Nutrient-Sensitive Regulatory Networks



Additional Complexity in Homeostatic Networks



Questions

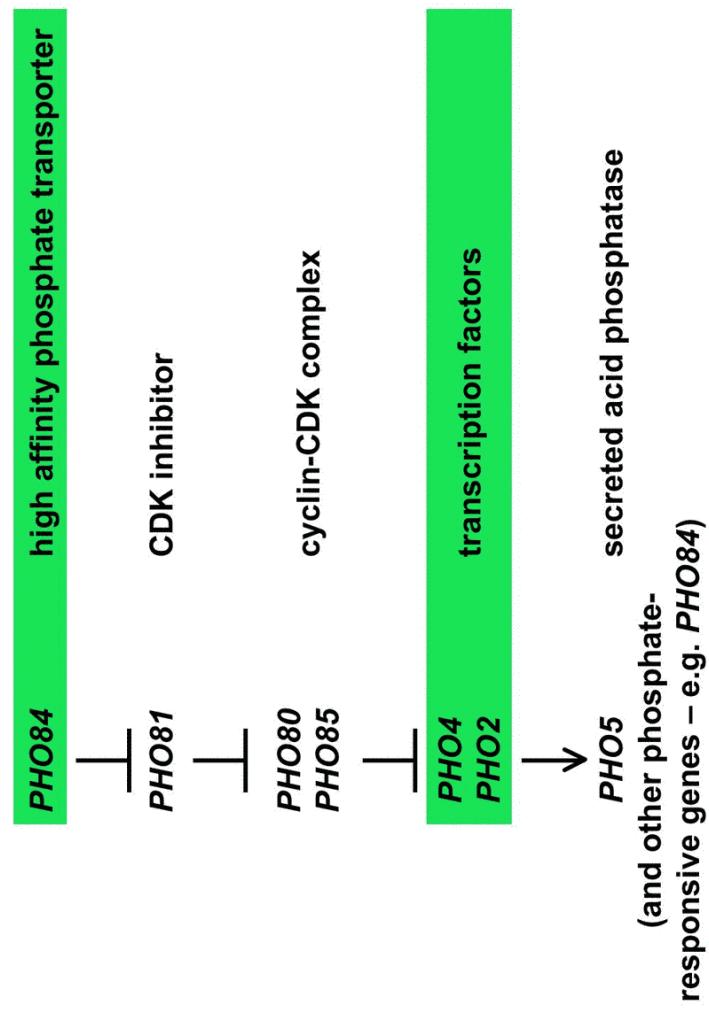
- How does this network structure give rise to homeostasis?**
- Is this network structure sufficient to give rise to homeostasis?**
- Which aspects of the design are critical for this property?**
- Is the same design/architecture used for many nutrient homeostasis systems?**

Strategy

- Use single cell reporters to characterize dynamics, response of system**
- Develop computational model based on network structure, experimental measurements**
- 2 properties: bistability, buffering**

Bistability

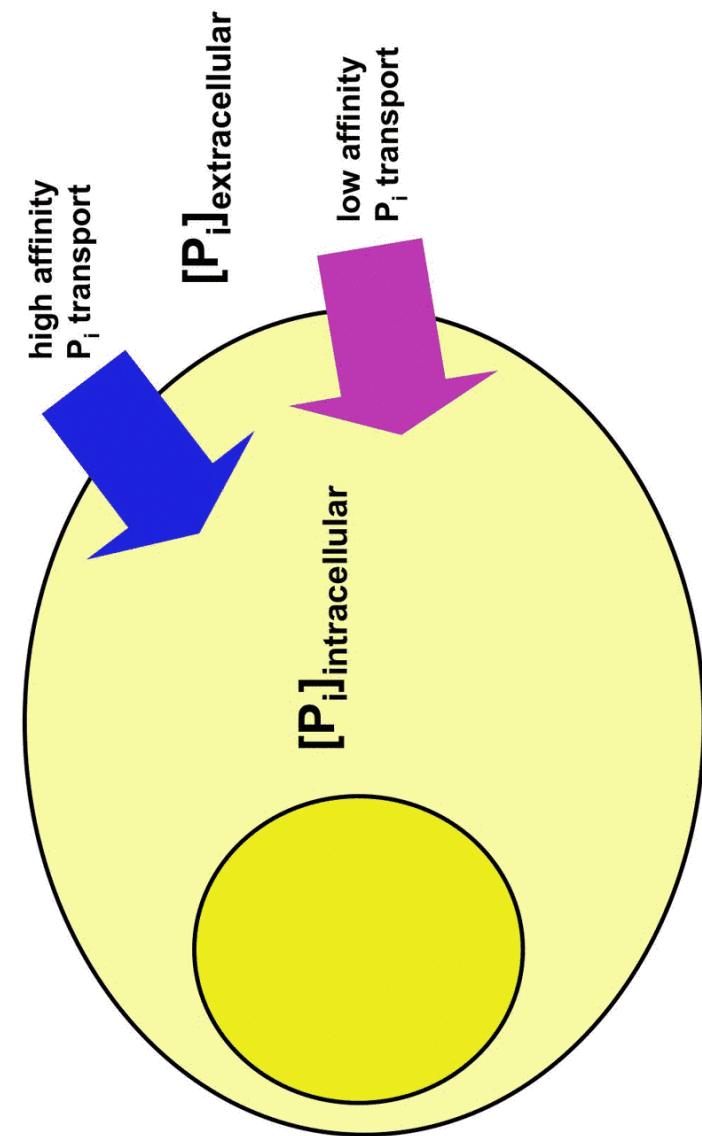
The Pho Pathway



Oshima et al.

Why does deletion of high affinity transporter turn the Pho pathway ON?

Cells Sense Phosphate Internally and Have Two Uptake Systems



Bistability

either

low affinity transport
ON

or

high affinity transport
ON

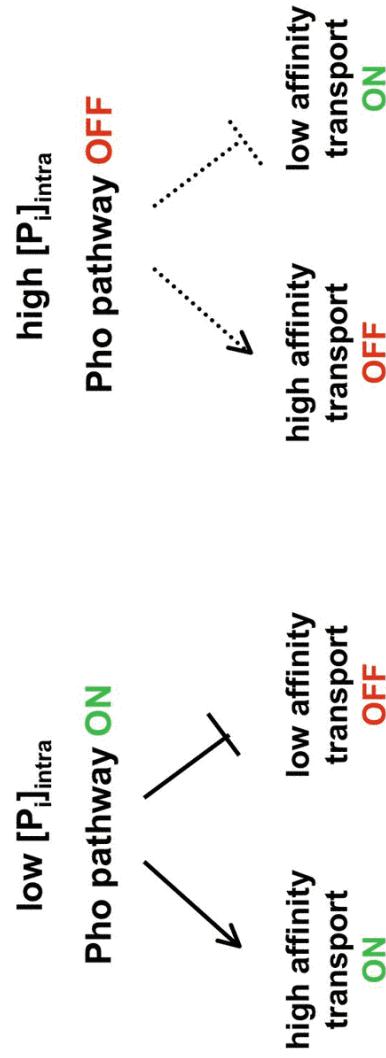
Arsenate Sensitivity

- arsenate enters cells through phosphate transporters
- proxy for phosphate uptake

	WT	<i>ars^S</i>	<i>ars^R</i>	P_i uptake defect	no P_i uptake defect
<i>pho84Δ</i>					
<i>pho84Δ pho81Δ</i>			<i>ars^S</i>		
<i>pho84Δ pho4Δ</i>			<i>ars^R</i>		

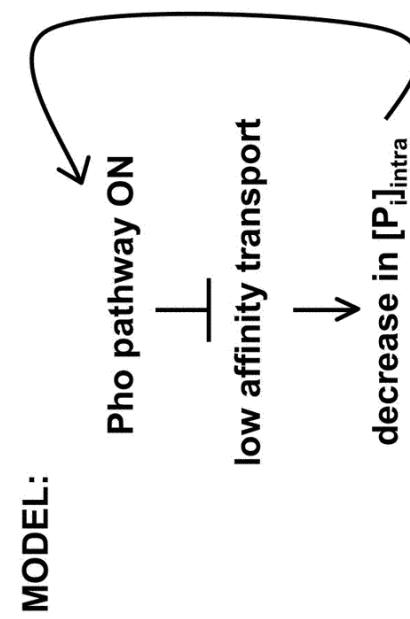
Inhibition of low affinity transport by Pho pathway requires Pho4

Model for Source of Bistability



Deletion of low affinity transporters \longrightarrow high affinity transport **ON**
(Harashima 2003)

Why does deletion of high affinity transporter turn the Pho pathway ON?



If inhibition of low affinity transport by Pho pathway requires Pho4...

PREDICT: $pho84\Delta$
 $pho84\Delta pho4\Delta$

Pho pathway ON



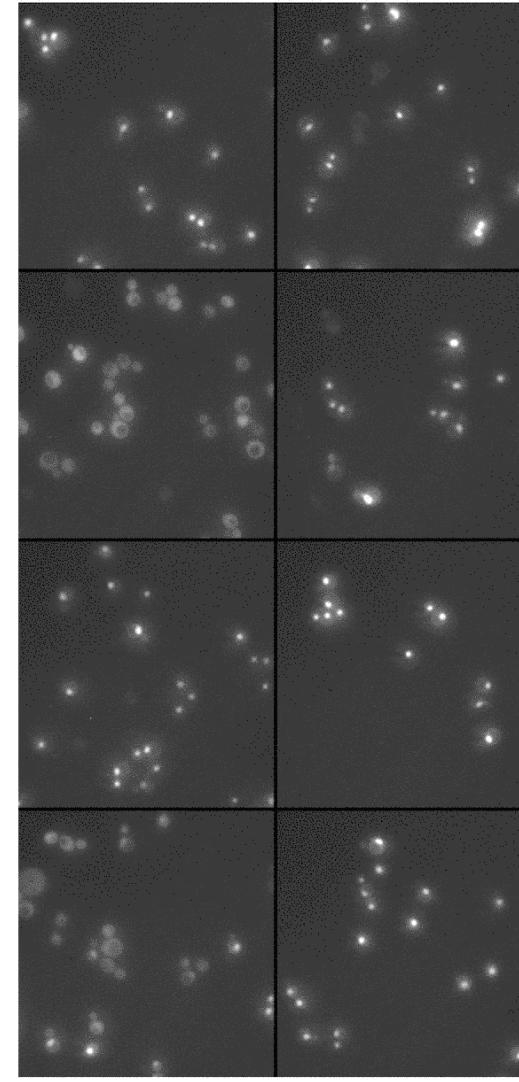
Pho pathway OFF



Pho4 cytoplasmic
 $pho84\Delta pho4\Delta$

Monitor Pho4 Δ DBD-GFP – cannot dimerize or activate transcription but localization regulated appropriately

PHO84 PHO4 *pho84 Δ PHO4* *pho84 Δ pho4 Δ* *pho80 Δ pho4 Δ*

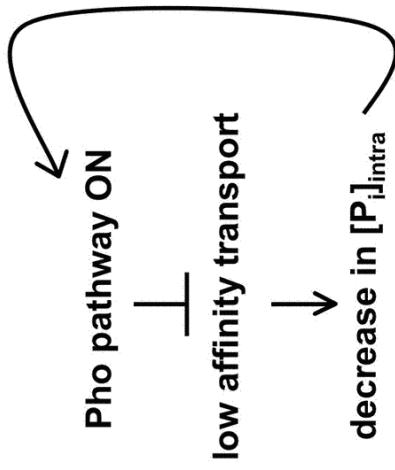


pRS-Pho4 Δ DBD-GFP

If low affinity transport is not repressed, *pho84 Δ* strain is not starving for phosphate and does not turn on Pho pathway

Implies Pho84 does not make a significant contribution to phosphate uptake in high phosphate

Why does *pho84 Δ* turn on the Pho pathway?



MODEL: *pho84 Δ* was transiently starved for phosphate

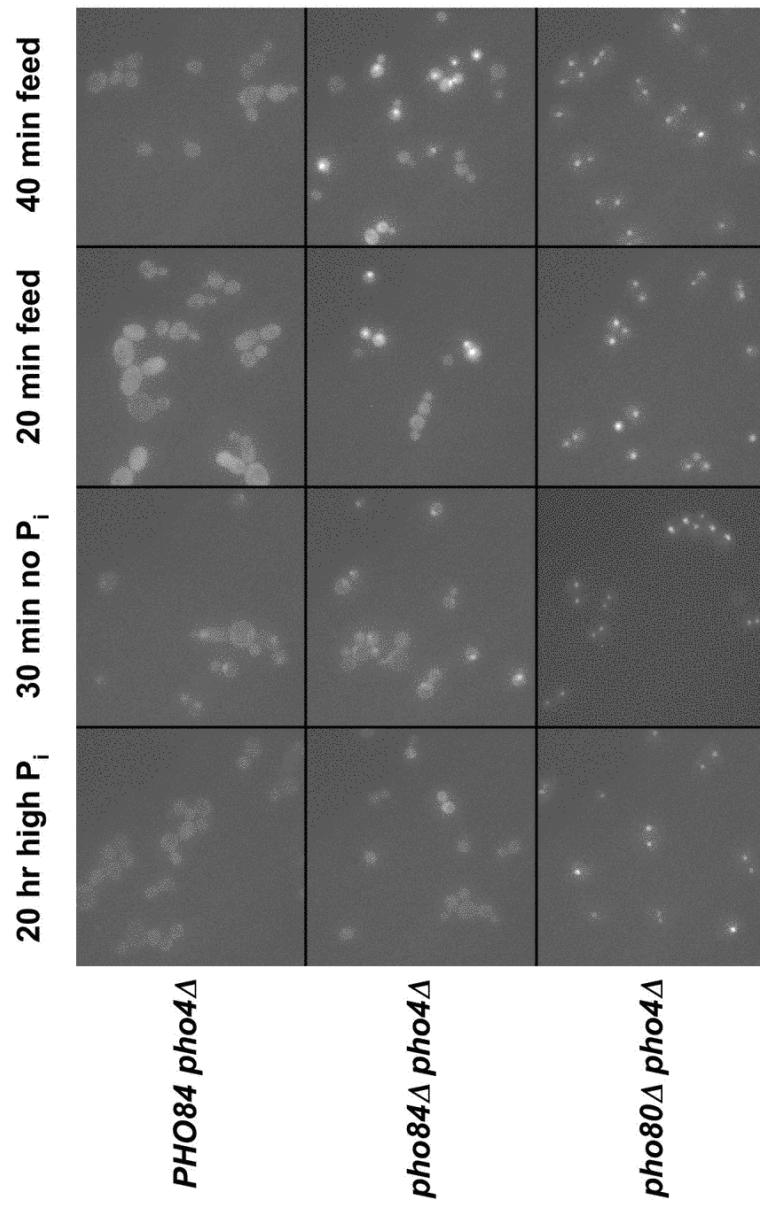
PREDICTIONS:

- (1) *pho84 Δ* should be able to exist with Pho pathway OFF
- (2) Pho84 is important for buffering against transient decreases in extracellular phosphate
- (3) Phosphate uptake defect in *pho84 Δ* strain should be dependent on growth conditions

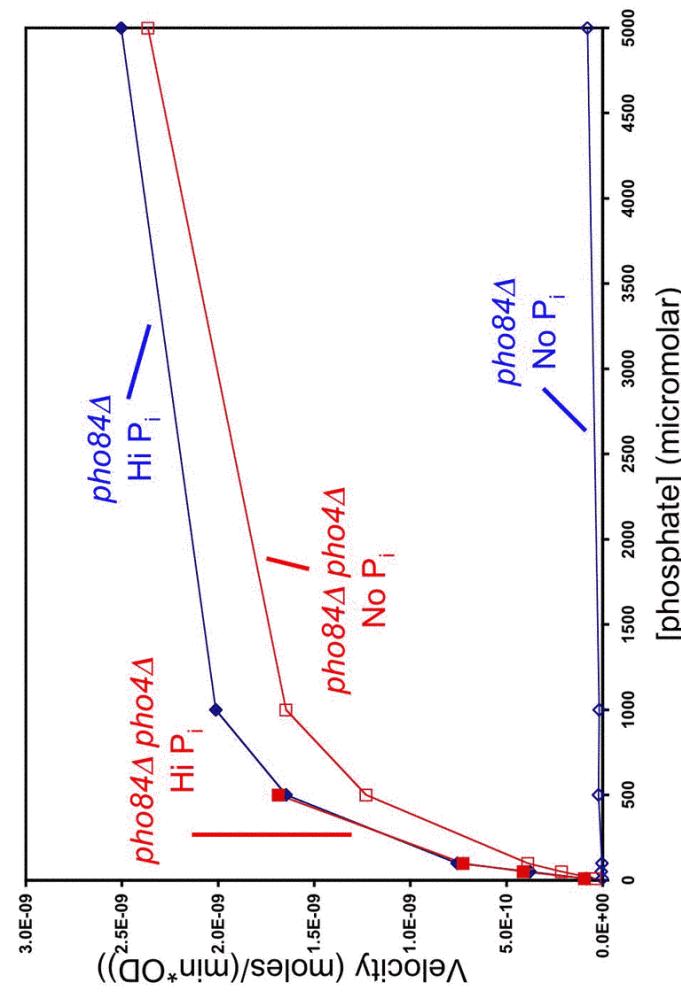
EXPERIMENTS:

Grow *pho84 Δ* cells containing Pho4-GFP for prolonged period of time in high phosphate – predict Pho4 will be cytoplasmic
Transiently deprive *pho84 Δ* cells of phosphate, add back phosphate – predict that cells will still have Pho4 in nucleus

Measure phosphate uptake of *pho84 Δ* cells grown in hi or no phosphate – predict that phosphate uptake rate will be Pho4-dependent and growth-condition dependent



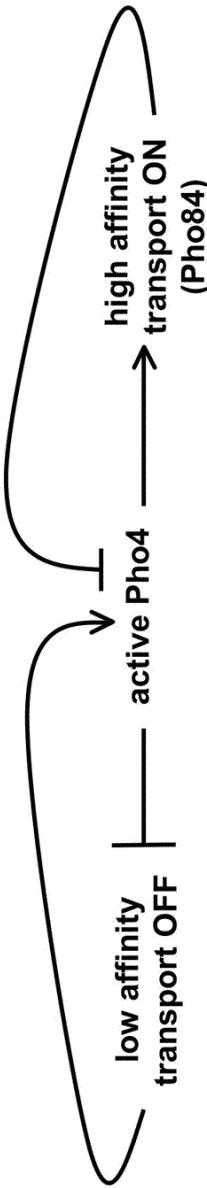
Phosphate Uptake in *pho84Δ* Strain



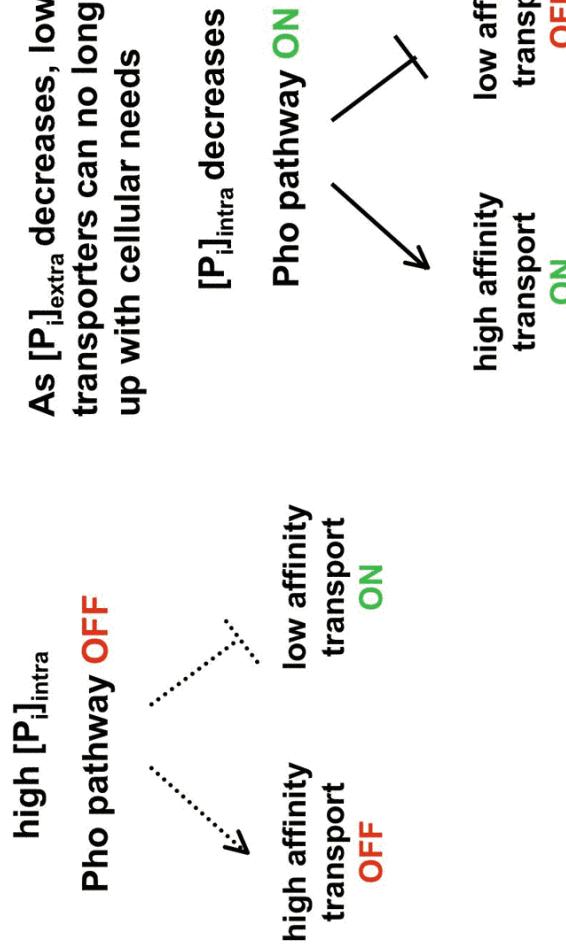
Conclusions...

pho84 Δ strain exhibits hysteresis – becomes “stuck” in Pho pathway ON state if transiently starved

Implies Pho84 is important for getting out of state where Pho pathway is ON

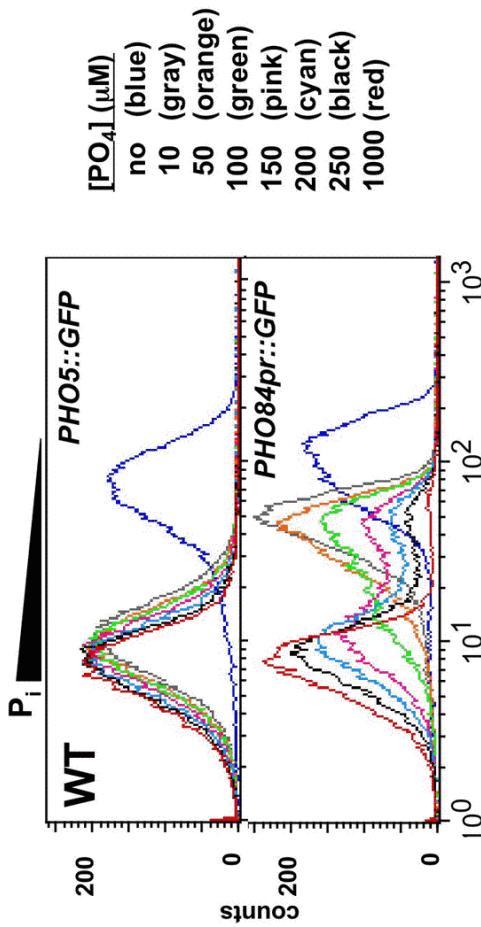


As $[P_i]_{\text{extra}}$ decreases, low affinity transporters can no longer keep up with cellular needs



Predicts that *PHO84* transcription might be bistable

PHO84 Transcription is Bistable

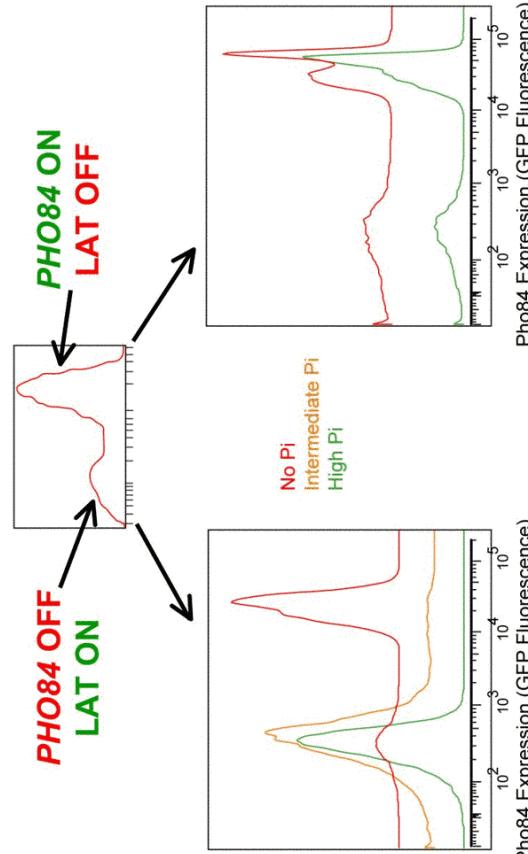


FACS analysis of *PHO5::GFP* and *PHO84pr::GFP* induction
as a function of Pi concentration

Slow Interconversion Between Two States

Grow cells 3 hrs in intermediate P_i

PHO84 ON
LAT OFF

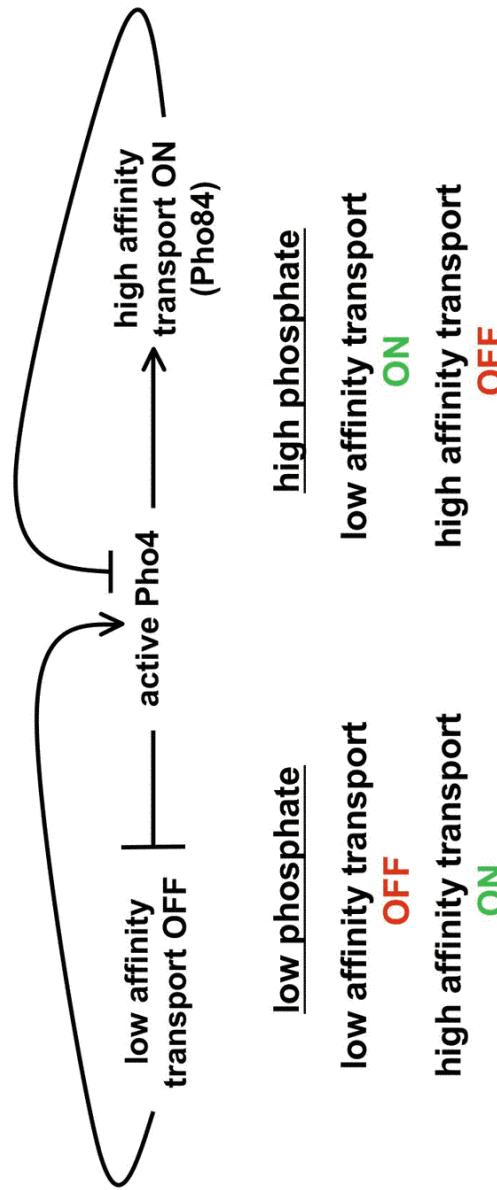


Grown 20 hrs post-sort

Properties of Two Populations of Cells in Intermediate Phosphate

PHO84 ON	Pho4 localized to nucleus
	<i>High affinity transport?</i>
PHO84 OFF	Pho4 localized to cytoplasm
	<i>Low affinity transport?</i>

Summary



Feedback from repression of low affinity transport by the Pho pathway probably contributes to bistability of *PHO84* induction
- test by interfering with feedback and examining bistability

Is this bistability advantageous?

Two populations may have different growth rates or fitness when grown in hi or no phosphate

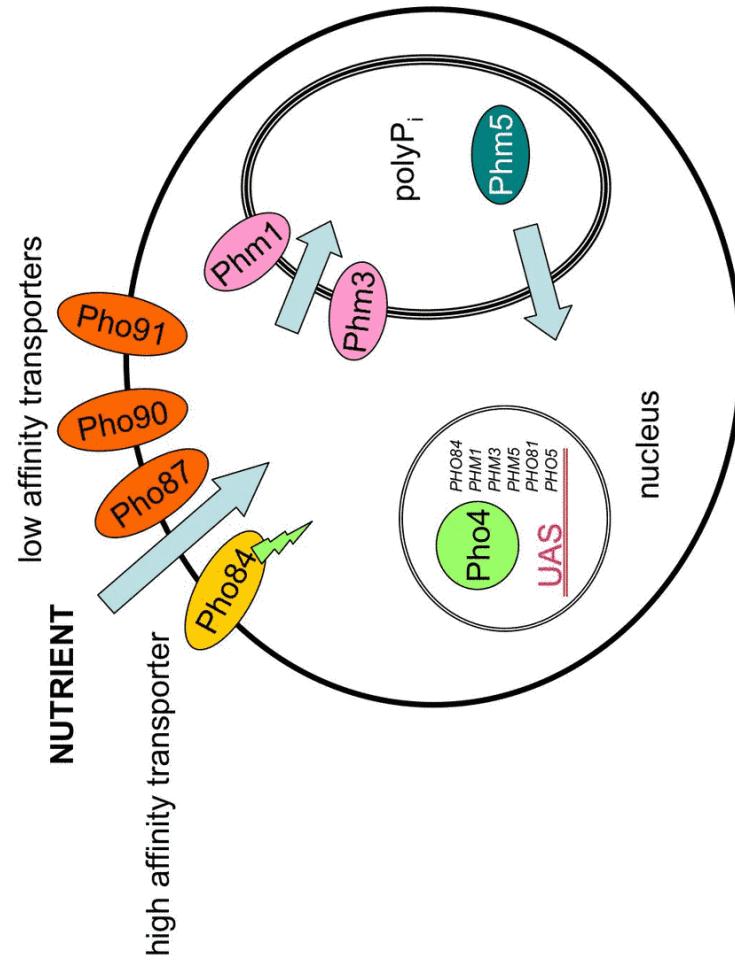
Test by sorting cells, measuring growth rate in different [phosphate], arsenate resistance

What generates split in population?

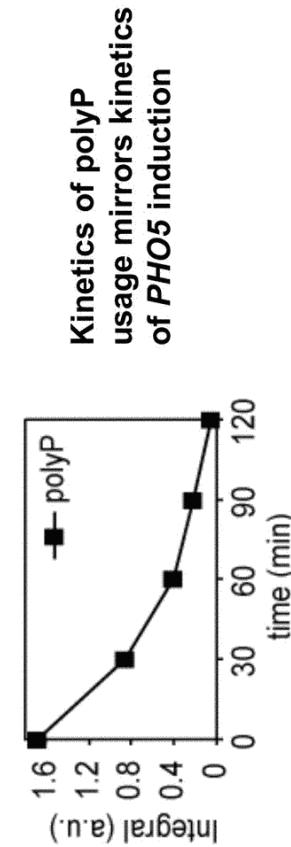
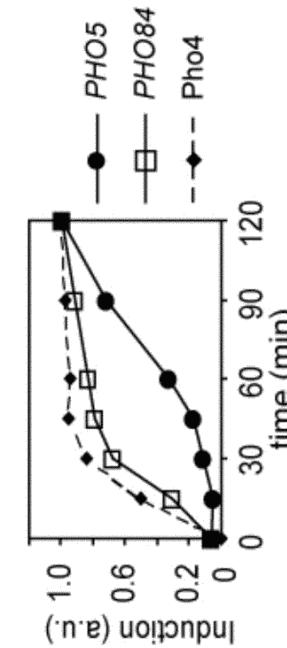
Role of stochasticity?

Buffering

P_i metabolism and signalling



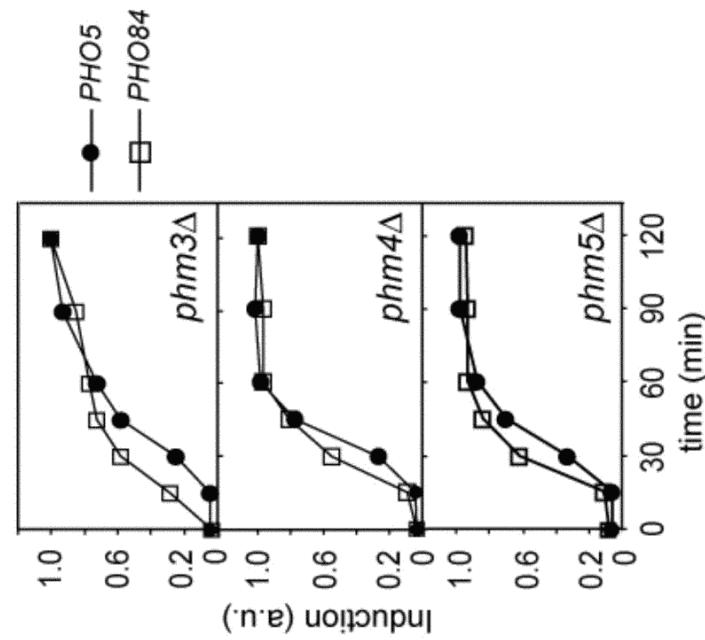
PHO84 and *PHO5* Exhibit Very Different Kinetics of Induction in No Phosphate



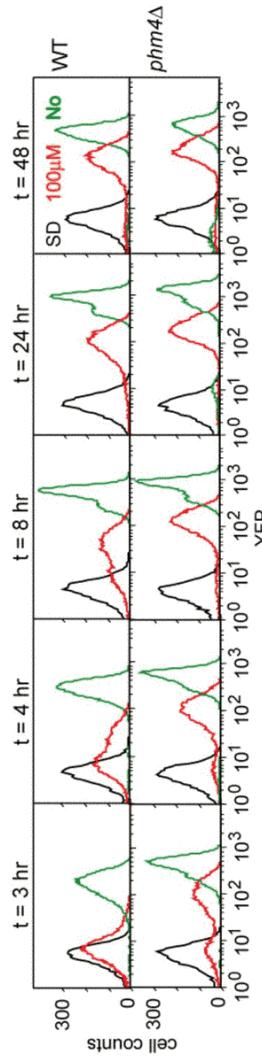
PREDICTION:

If polyP is functioning as a buffer that is mobilized to slow *PHO5* induction, cells lacking polyP should induce *PHO5* rapidly

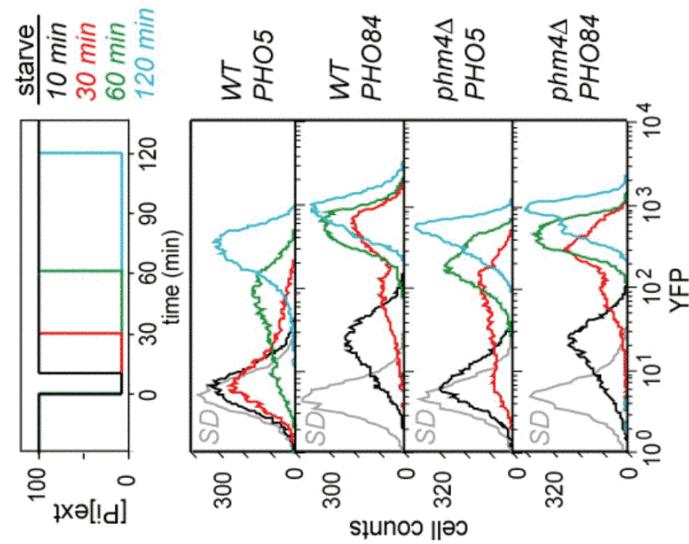
No Kinetic Delay in *PHO5* Induction in Cells With Altered Polyphosphate Metabolism



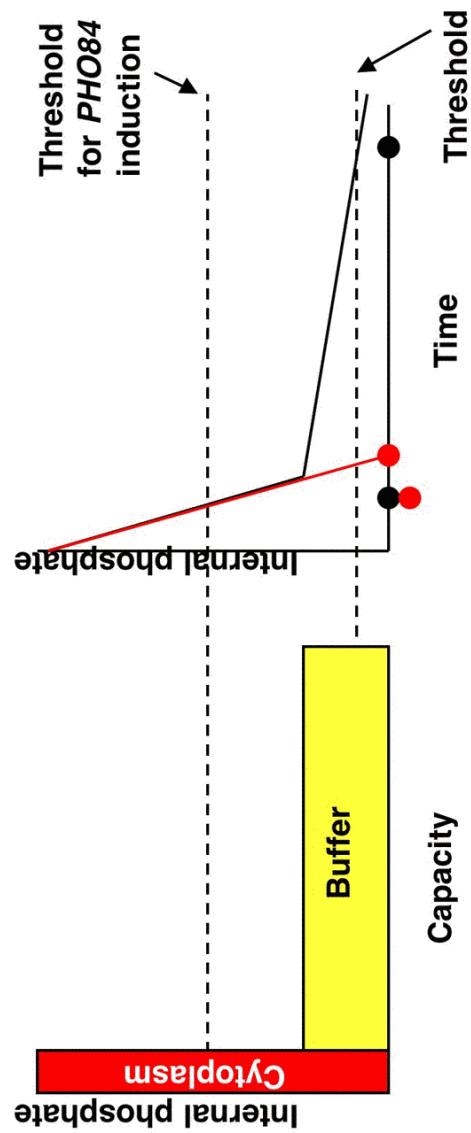
Loss of Polyphosphate Does Not Affect Threshold of Phosphate Required for *PHO5* Induction



Polyphosphate Mobilization Buffers Cells From Transient Fluctuations in Phosphate



Buffering and Sensitivity



Jonathan Raser
Dennis Wykoff
Brian Margolin

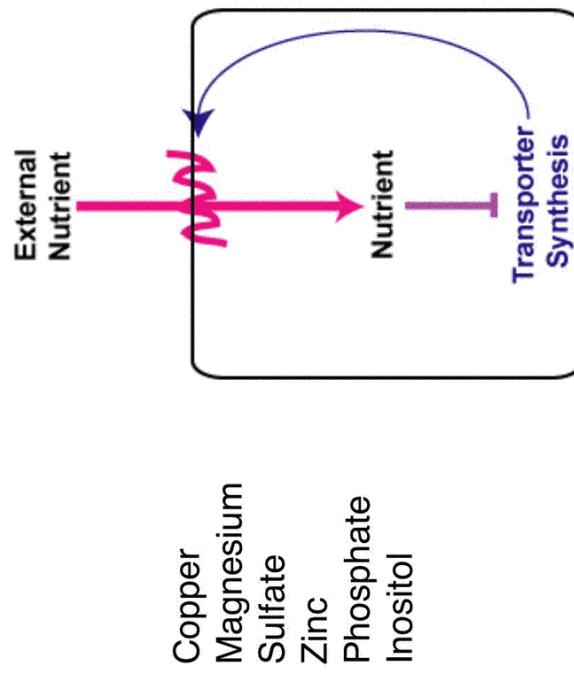
Melissa Thomas

Mike Springer

Experimental design

- For the pRS-Pho4- Δ DBD-GFP experiments, the strains were grown O/N in SD, starved for 45', then either fed with 11 mM Pi for 15' (labeled 'high Pi') or continued to starve (labeled 'no Pi'). This starve/feed was done to produce uniformly nuclear signal in the pho84 delete, and mimics our traditional practice of diluting from an overnight saturated culture.
- For the YCp50-Pho4-GFP experiments, the strains were grown in log phase in SD complete for ~20 hours (1st picture), then starved for 30' in no Pi (2nd picture), then fed with 7 mM Pi, equivalent to SD, for 20 minutes (3rd picture) and 40 minutes (4th picture). The only source of Pho4 in these strains is the plasmid.
- The TIF files are labeled logically; if you open them with Photoshop, you have to adjust the contrast/brightness (they will look blank until you do so). The JPEG's for import into Powerpoint are called 'Pho4feed' and 'Pho4DBDGFP'.

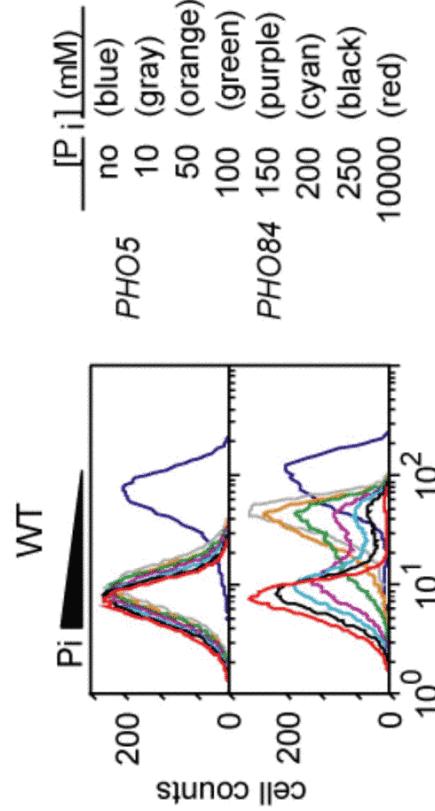
Nutrient-Sensitive Regulatory Networks



pho and zinc homeostasis projects

- experimental measurements
 - transcription/transporter abundance
 - steady-state and dynamic behaviors
 - perturbation analysis (mutation and conditions)
 - physiological assays
- computational modeling and control theory analysis
 - fit and predict experimental data
 - model ideal behavior and control

PHO84 Transcription is Bistable



FACS analysis of *PHO5::GFP* and *PHO84pr::GFP* induction
as a function of Pi concentration

