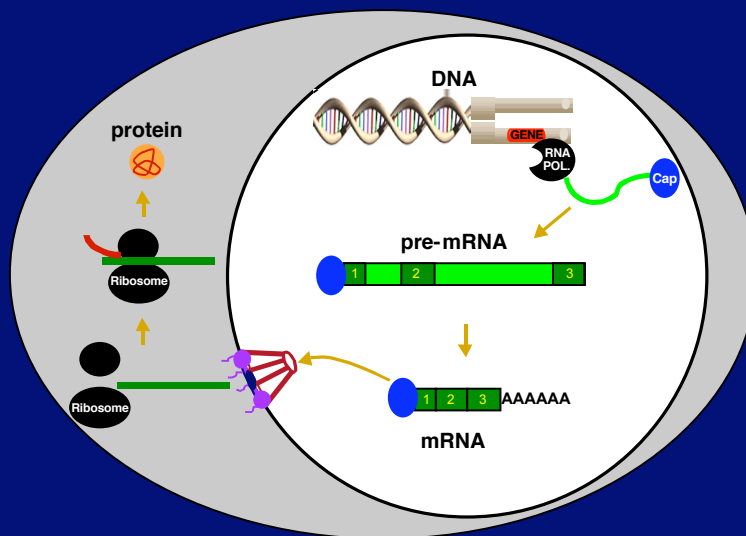


Computational Molecular Biology of Genome Expression and Regulation

Michael Q. Zhang
Cold Spring Harbor Laboratory

- An ESE (enhancer-sequence-element) SNP can alter RNA splicing (Brac1:exon18)
- Classification of 5'UTRs by *CART*
- Promoter prediction and analysis
- *CSEdb* and human gene number

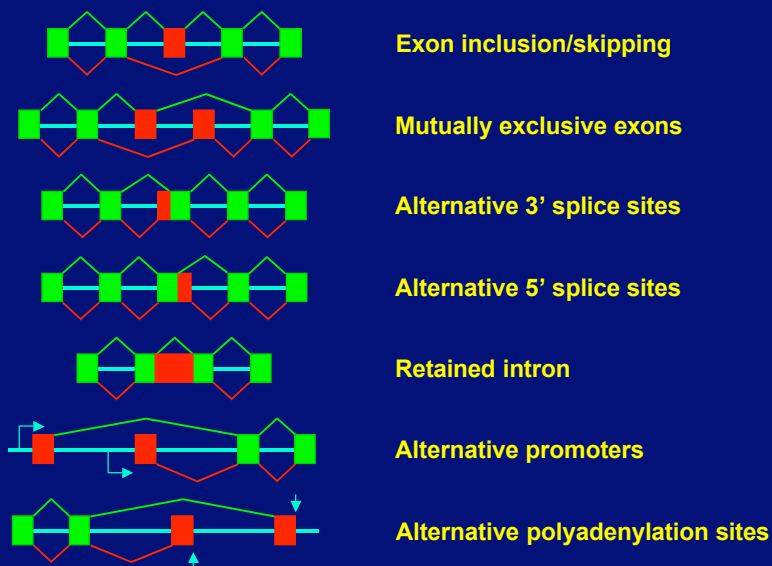
Classical view of eukaryotic gene expression

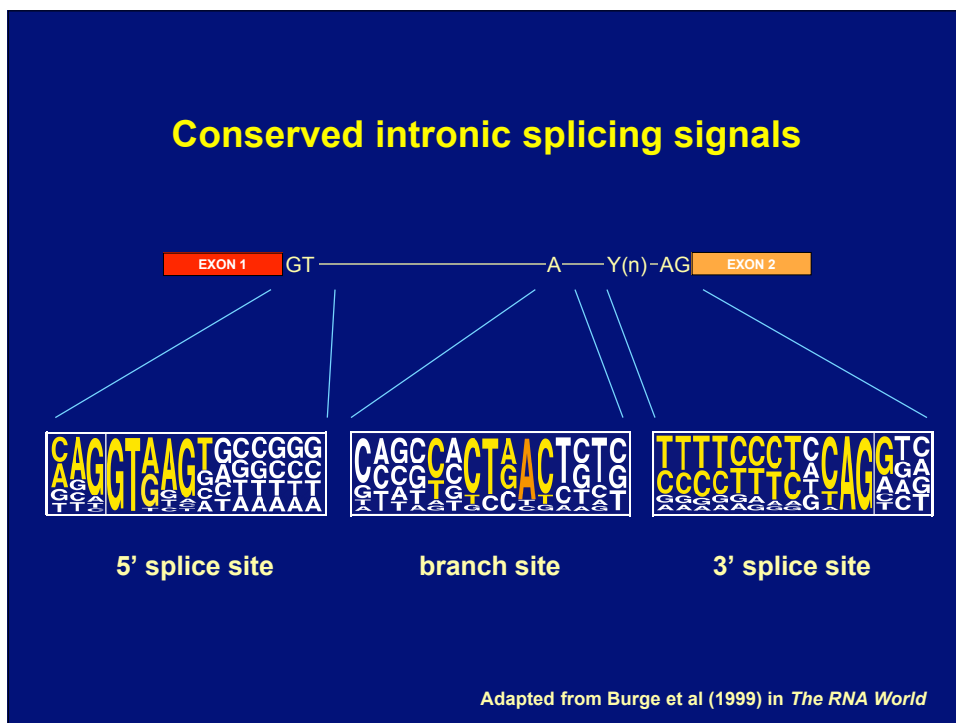
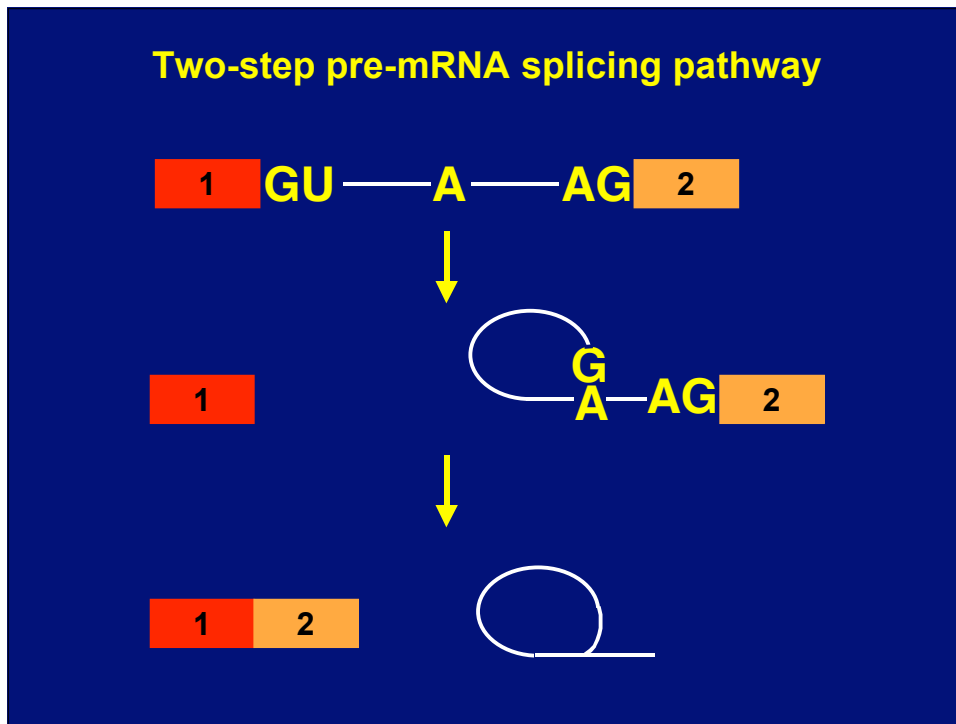


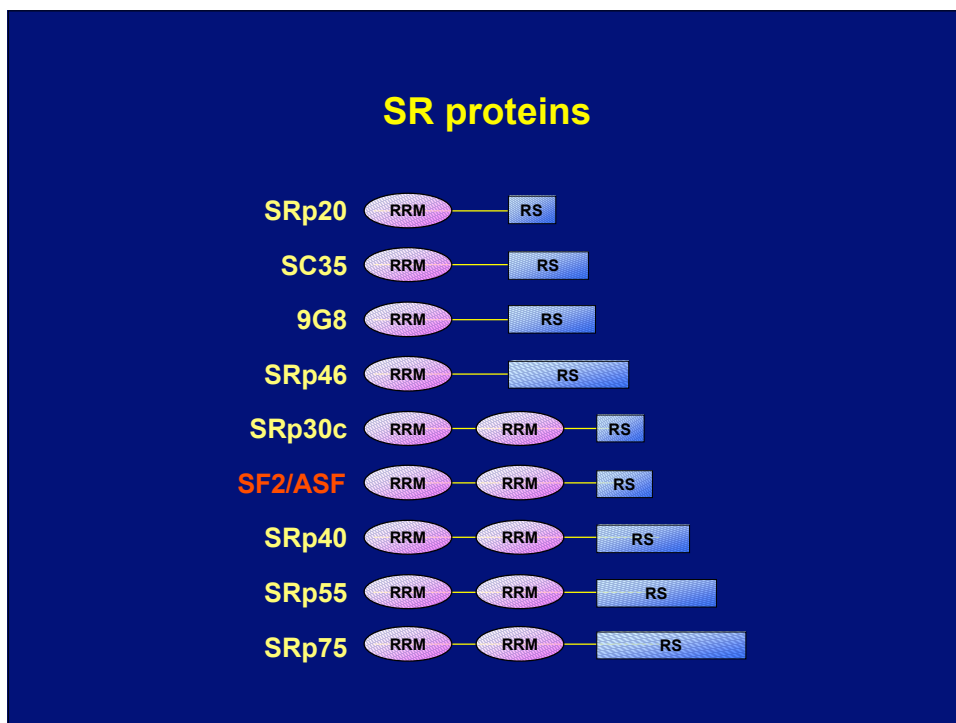
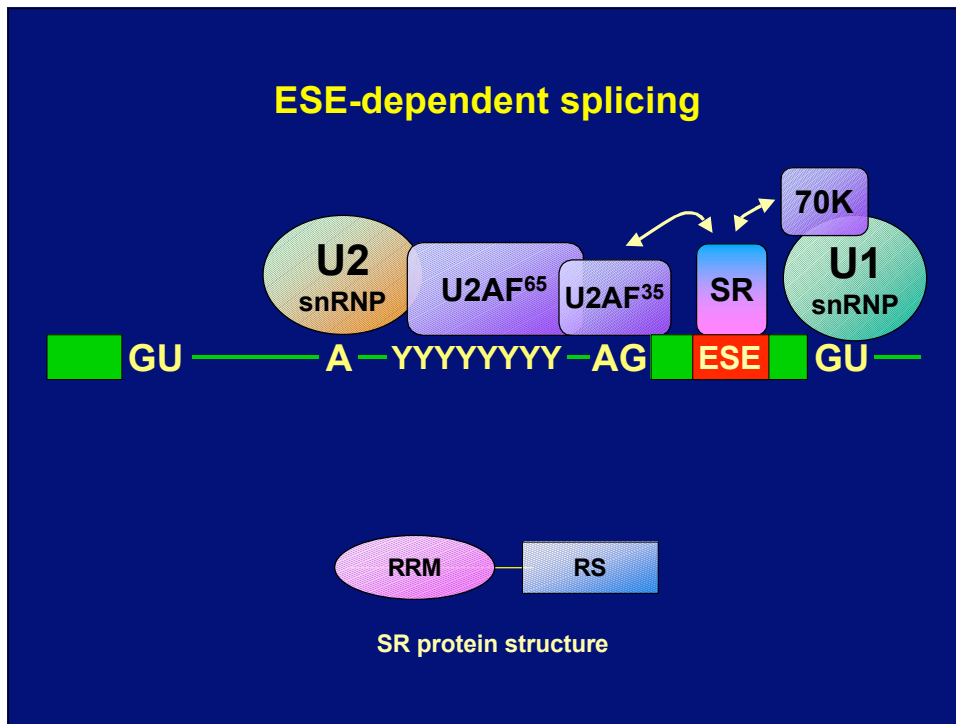
An ESE (enhancer-sequence-element) SNP can alter RNA splicing

(Collaborated with Krainer lab at CSHL:
Nature Genet. 2001)

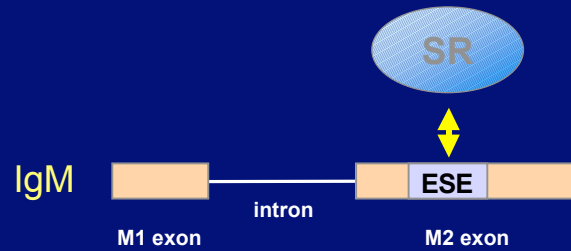
Patterns of alternative pre-mRNA splicing







Functional SELEX approach to study SR protein specificity



IgM splicing in S100 extract requires

- SR proteins
- Exonic Splicing Enhancer

ESE consensus motifs

(G&D1998, MCB1999)

CACACGA
GGCGGT
GTGTG

SF2/ASF

Max : 6.589
Thr: 1.956

GGCCCTG
GATTAGCA
ATAGAA

SC35

Max : 6.221
Thr: 2.383

TCACAGG
CTATCG
AGGCAC

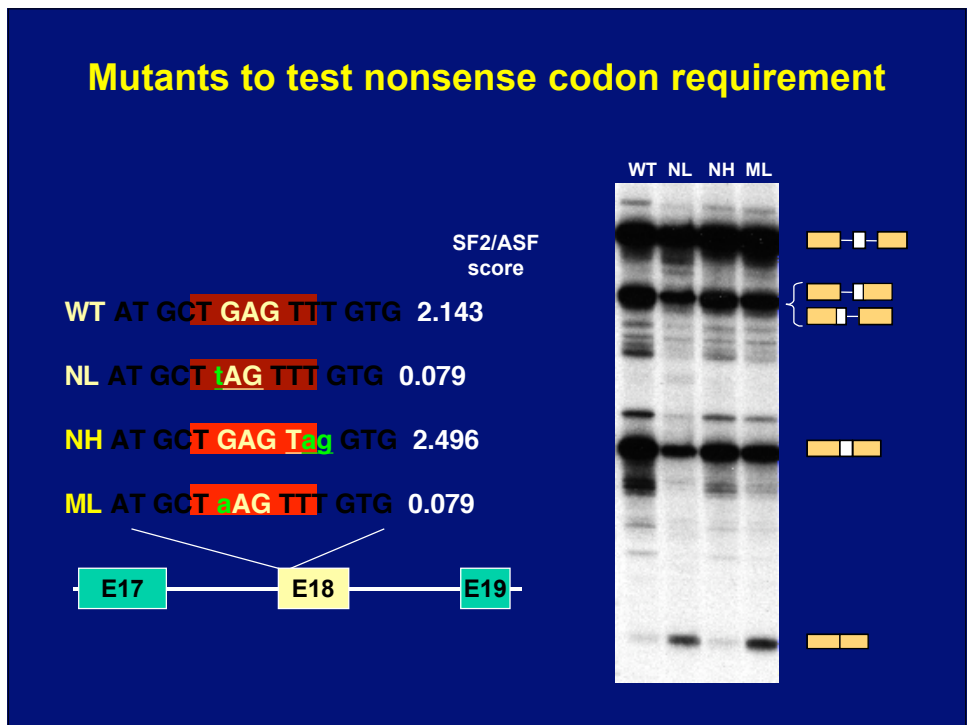
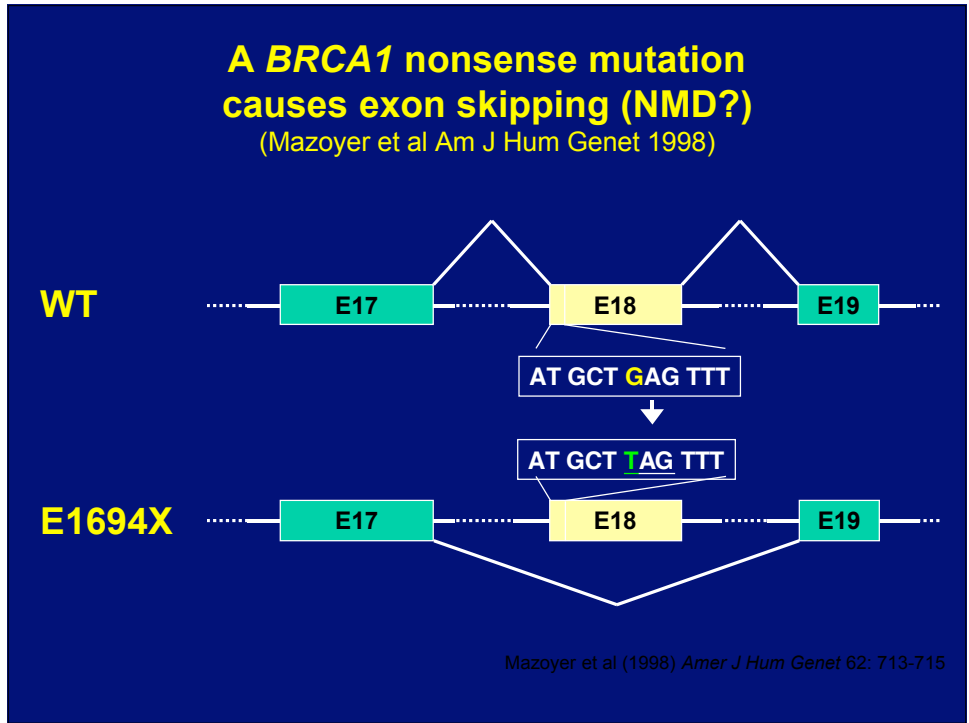
SRp40

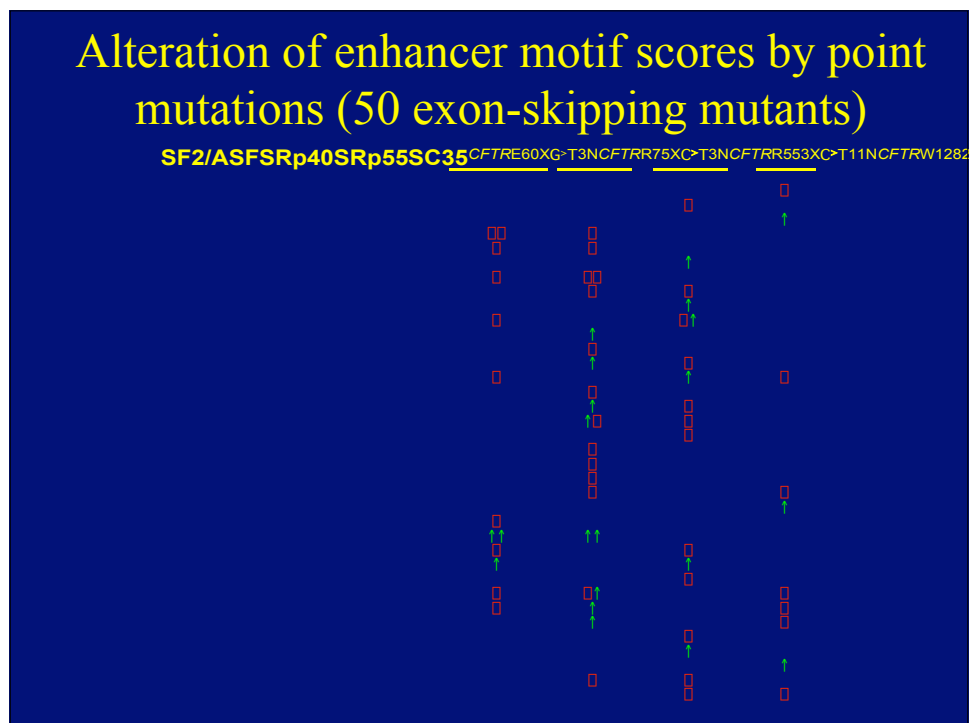
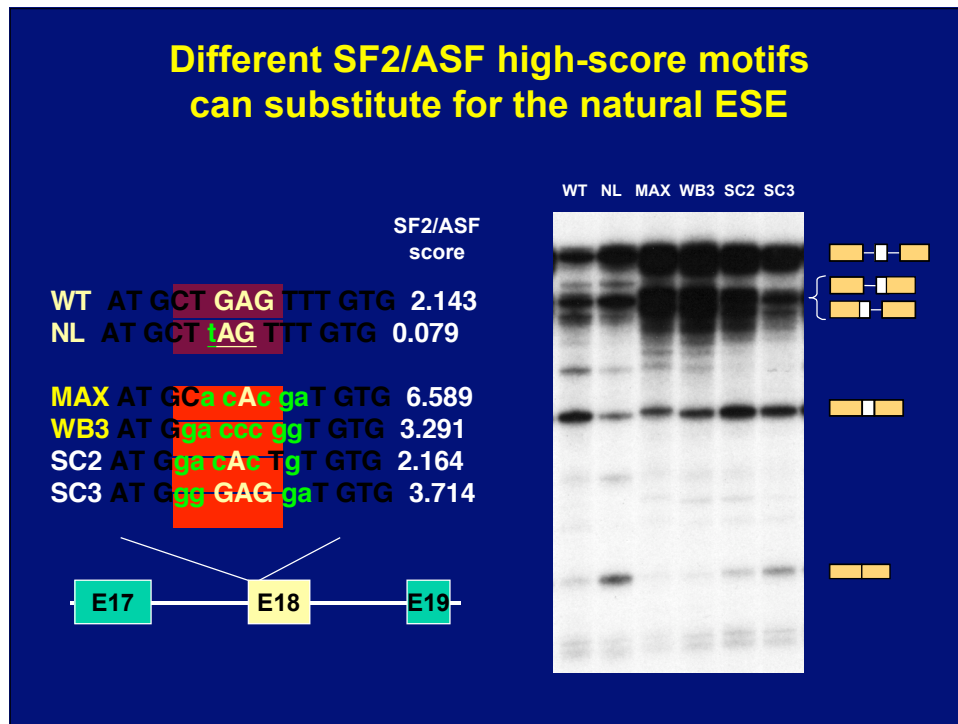
Max : 6.324
Thr: 2.670

TGCGTC
CATAGA

SRp55

Max : 6.135
Thr: 2.676





Computational Molecular Biology of Genome Expression and Regulation

ESEfinder: a web resource to identify ESEs

(<http://exon.cshl.edu/ESE/> Cartegni et al. submitted)

The screenshot displays the ESEfinder web interface. It includes a navigation menu with options like 'background', 'methods & thresholds', 'output', 'graphic representation', and 'caveats'. The main area is divided into sections for 'Choose the matrix and the threshold to be used', 'SR proteins (select one or more)', and 'Thresholds'. Below these are input fields for 'Enter with your input FASTA format' and 'alternatively, upload a file'. The search results are presented in two tables, one for 'SRP40' and one for 'SRP40', each with columns for 'Position', 'Motif', and 'Score'. A bar chart at the bottom right shows the distribution of motifs across different SR proteins: SRP40 (red), SRP40 (green), SRP40 (blue), and SRP40 (yellow).

- **ASDB** (with Stamm/Max-Planck, Nakai/Kyoto, 2001)
- **mATDB** (Wang & Zhang, to be submitted)
- **The RNA-mediated Annealing, Selection and Ligation Assay (RASL): Application in Alternative Splicing** (with Fu&Gribskov/UCSD and Fan/Illumina, NCI funded)

The diagram illustrates the RASL assay process in five steps:

- Step 1: Oligo Annealing**: A pre-mRNA with a 5' cap (Cap) and a poly(A) tail (p(A)_n) is shown. Two zip-code oligos, zip1 and zip2, are annealing to the pre-mRNA.
- Step 2: Poly A⁺ Selection**: The pre-mRNA is selected based on its poly(A) tail.
- Step 3: RNA-dependent Oligo Ligation**: The zip-code oligos are ligated to the pre-mRNA in a sequence-dependent manner.
- Step 4: Amplification using Universal Primers**: The ligated pre-mRNA is amplified using universal primers.
- Step 5: Hybridization to Zip-Code Arrays**: The amplified pre-mRNA is hybridized to a zip-code array for detection.

Classification of 5'UTRs by *CART*

(Collaborated with Sunoga lab at Tokyo U.
Genome Res. 2000)

Forms of translation regulation by 5'UTR



Some examples

Gene Name	Length	uAUGs	Mechanism
c-mos(ovarian mRNA)	80	0	secondary structure
c-mos(testicular mRNA)	300	4	uAUG
RAR beta2	461	5	uORF
PDGF2/c-sis	1022	3	IRES
TGF beta 1	840	0	secondary structure
ATM (cancer)	146-884	1 to 8	uAUG
AR	1116/1127	1	uAUG
c-myc	408	0	IRES
FGF-2	484	0	IRES
IGFII (p3)	1170	0	secondary structure
IL-15	313	10	uAUGs
TGF beta 3	1104	11	uAUGs
TGF beta 3 (Breast cancer cells)	297	0	highly expressed
Spi-1	151	0	secondary structure

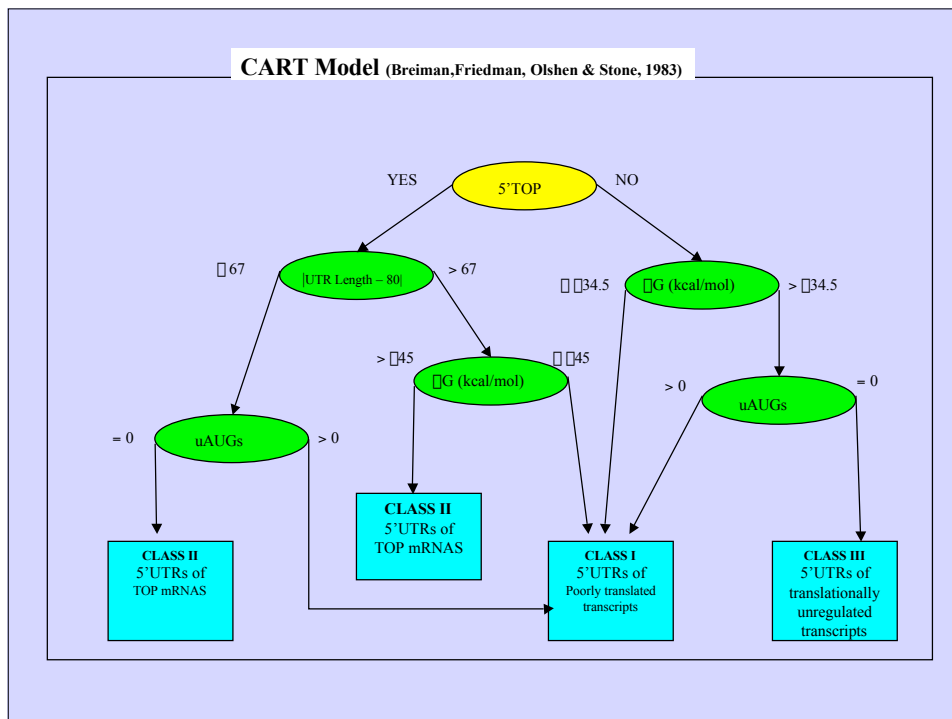
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5'UTR database

- a set of 954 human 5'UTR sequences was obtained from *5' end-enriched cDNA library* (Suzuki et al. 2000) with their mRNA start sites mapped
- a second set of 1613 full-length 5'UTR sequences retrieved from UTRdb (Pesole et al. 2000) database
- all the redundant and ambiguous sequences were eliminated and finally a non-redundant set of 2312 5'UTR sequences was prepared for the analysis

CART classification of human 5'UTR sequences

- **Class I (226):** 5'UTRs of growth factors, their receptors, transcription factors, proto-oncogenes, cytokine receptors and tumor suppressor genes. Most of these are understood to be **translationally repressed** mRNAs.
- **Class II (70):** This class consists of TOP mRNAs. (5'terminal oligopyrimidine tract-5'TOP), The **translation is regulated in growth dependant manner**.
- **Class III (76):** 5'UTRs of highly expressed genes, tubulins, globins, globulins, myosins, caseins, glycolytic enzymes, beta-actin, gamma-actin and histones. These transcripts are believed to be **efficiently translated or (at least) not repressed at the translational level**.



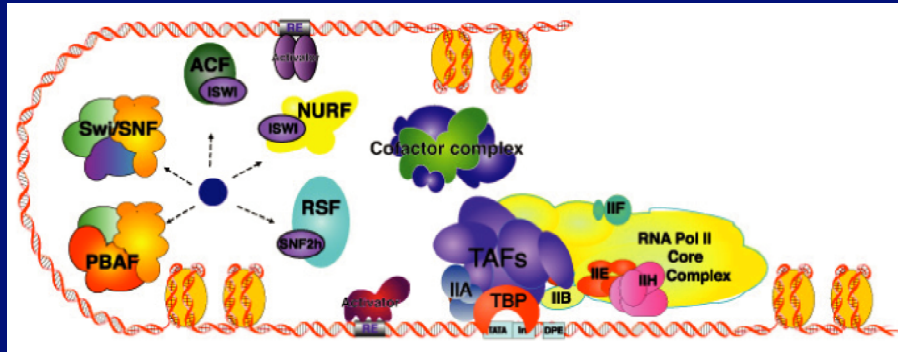
Cross Validation Classification:

Actual Class	Predicted Class			Actual Total
	I	II	III	
I	210 (93.0)	1 (0.4)	15 (6.6)	226
II	0 (0)	70 (100)	0 (0)	70
III	10 (13.2)	2 (2.6)	64 (84.2)	76
Predicted Total	73	220	79	372

RESULTS

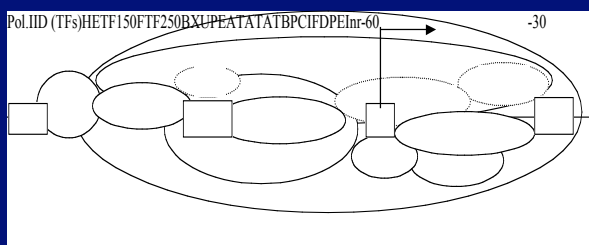
- ΔG was the most discriminative variable for the three classes. ΔG was followed by presence of TOP, 5'UTR length, number of stable free energies, presence of stable secondary structure within the first 100 bp from the cap site, CDS length, A/T ratio, G/C ratio, number of uAUGs, GC%, number of uORFs and codon bias, in the order of relative importance for predictive classification.
- More than 90% of Class I 5'UTRs are embedded with stable secondary structures with ΔG less than -50 kcal/mol. Classes II & III are almost free from this translational inhibitory feature. Also 60% of the Class I 5'UTRs have stable secondary structures within the proximity of the transcription start site.
- Presence of uAUGs and uORFs was observed as common feature in Class I 5'UTRs where as Classes II and III are quite free from these features.
- 65% of Class II transcripts are in good start site context followed by Class III with 57% and Class I with 49%.
- There was not any significant difference in GC% between the three classes.
- There was not any significant difference in mean codon bias between the three classes, which indicates that the codon usage and expression level in human genes are not correlated. In contrast, codon bias plays an important role in translational efficiency in some lower eukaryotes, such as yeast (Sharp and Li, 1987).

Promoter analysis *in silico*



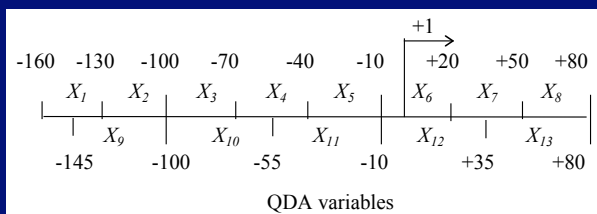
- De novo (database/training set) -> TSS;
- Functional genomics (expression/localization) -> cis-elements;
- Comparative genomics -> TSS & cis-elements

Core_Promoter (Zhang, Genome Res. 1998)



GenBank
M12523:1..1980
ALB gene=serum albumin
Firstexon=1737..1854
AUG=1776
C+G=0.33

Core_Promoter prediction:



TSS	Score
1737	0.637
1736	0.604
1727	0.588
1732	0.534
1731	0.531
1728	0.498
1726	0.428

Computational Molecular Biology of Genome Expression and Regulation

CpG_Promoter (Ioshikhes&Zhang *Nature Genet.* 2000)

CpG island: Length > 200 bp; C + G content > 50%; CpG ratio Obs/Exp > 0.6

- 135 genes
 - 68 have CpG island around promoter
 - 63 recognized
 - SN = 0.47 (0.93)
 - SP = 0.34 (1 Pos./26 kb; 1/36 kb is in fact)
- Promoter Scan gives
SN = 0.44
SP = 0.06 (1 Pos. / 4.7 kb)

GenBank	CpG_Promoter prediction:		Core_Promoter prediction:	
	CpG islands associated	Promoter-	TSS	Score
D87675	8813..9319		8921	0.100
>301kb	9328..9547	+	8923	0.094
App gene encodes	9761..10203	+	8920	0.089
Amyloid precursor protein	117256..117511	-	8919	0.084
Firstexon=9001..9204	176132..176342	-	8922	0.078
AUG=9148	257735..257942	-	8918	0.058
	261475..261750	-	8783	0.056

First exon prediction (FirstEF) (Daluvuri,Grosse&Zhang, *Nature Genet.* 2001)

Performance statistics of FirstExonFinder based on cross validation

Exon Type	Sn	Sp	CC
CpG-related	0.92	0.97	0.94
Not CpG-related	0.74	0.6	0.65
All Exons	0.86	0.83	0.83

Promoter Prediction accuracy of FirstEF and PromoterInspector for Ch22

Program	TP	FP	Sn	Sp
FirstEF	46	40	79.30%	53.50%
PromoterInspector	28	37	48.30%	43.10%

Prediction Accuracy for Ch21&22 (Number of Real Promoters: 58)

Chromo-some	Number of Exponentially mapped first exons	Number of correctly predicted first exons	Completely non-coding exons	Predicted non-coding exons
21	42	37 (88%)	14	10 (71%)
22	79	69 (87%)	28	23 (82%)
Total	121	106 (88%)	42	33 (79%)

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FirstEF program

FirstEF: first-exon and promoter prediction program for human DNA

[README](#)

[Research License instruction and agreement form](#)

Note: the maximum acceptable sequence file length is 100 KB.

Please paste your sequence in FASTA format (First title line begin with a ">")

Or, give the name of a file containing the sequences in FASTA format

Browse...

cutoff value for the first-exon a-posteriori probability: 0.50

Cell Cycle Regulation (Spellman *et al.* MBC, 1998)

(Brown & Botstein labs – Futcher & Zhang labs)

Alpha cdc15 cdc28 Elu

M/G1

G1

S

G2

M

SCPD - The Promoter Database of *Saccharomyces cerevisiae*

Zhu & Zhang, 1999

- Genes: Explore the promoter regions of ~6000 genes and ORFs in yeast genome
 - Provide information on genes with mapped

Name	Gens	Group	Motif	Sites	(% Genes)	Sites	(% Genes)	out of 256 controls	Choet al	Cl	258	G1	MCB	AC	GC	G1	SCB	CRCC
SW15			TAACCTT TTAGAAAAAGTTAAACAATAC															
CLB1			CGCCCAAAAGGGAAAAATCAACAAATCA															
CLB2-1			CGACCGAATCAGGAAAAAGTCAACAACGA															
CLB2-2			TTTCCCTAAACGGGCTCAAATATGTAACA															
BUD4			TGACCCGATTTGGAAAAAGTTAAACAACA															
RIM3			TTTCCCTAATAGGTTAAACGTTAAATAAG															
CDC20			TGCGGAAAGAGCAAAAAGTAAATAAGTTG															
YOR315W			TGCGCCAAATAGGATAAAGTAAATAACATA															
YHS2			ATACTCAAATAGGAAATATATAACAATAAG															
YRO2			CAACCCGATGAGGAATCATCCCGATCTAAC															
WSC4			TGCGCAAGGTGAATACCGTAAATGATACC															
APC11			TGACCGTAGTGGAACTGTTCCAAACCTTTT															
FAR1			ATACCTAAAGGAAATAGCAATAAATGA															
CDC5			AAACCCAAATAAAGAAATCCAAATA TAGAA															
YML119W			TTTCCGATTAGGAAAGACATAAATAAATA															
UBA2			TTTCTGATTTGGTAAAGACTAAATGAGAA															
DBF2			TTTTCTTTTGGGTTGGTCCCTCGGAAATGG															
CDC16			TTTCTTATTGGGTCACACAAACCGATTA															

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Cell Cycle Regulation (continued)

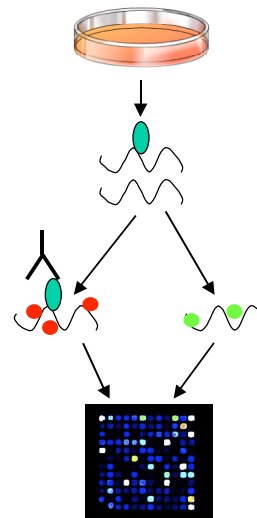
Computationally predicted E2F target genes confirmed by *in vivo* footprint (ChIP) (Kel *et al.* JMB 2001)

Gene	EMBL	Sequence of the potential sites	Position rel. start transcription	Score, q	d(X)
c-fos , <i>Homo sapiens</i>	HSFOS	(+) gttttggccggcagc (+) ggtttggccggccgagc (+) cctttggccggccgagc (-) agtttggccggccgagc	-165 .. -176 -92 .. -103 -90 .. -79 -78 .. -89	0.915 0.836 0.878 0.830	2.92
JunB , <i>Homo sapiens</i>	HS207341	(+) gttttggccggcagc (+) cctttggccggccgagc (+) ggtttggccggccgagc	79 .. 90 91 .. 80 169 .. 158	0.887 0.905 0.820	3.16
TGF-β1 , <i>Homo sapiens</i>	HS164818F	(+) gttttggccggcagc (+) cctttggccggccgagc (+) cttttggccggccgagc (+) gttttggccggcagc (+) ggtttggccggccgagc (+) cctttggccggccgagc	+513 .. +502 -298 .. -287 28 .. 39 40 .. 29 85 .. 96	0.804 0.912 0.928 0.830 0.854	2.03
ARF , <i>Homo sapiens</i>	AF042338	(-) gttttggccggcagc	-265 .. -276	0.859	
Mcm4 (Cdc21), <i>Mus musculus</i>	AB000629	(+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc	-443 .. -432 -431 .. -442 -329 .. -318 -297 .. -286 -127 .. -116 -24 .. -13	0.872 0.935 0.810 0.846 0.809 0.858	
MCM5 (P1-CDC46), <i>Homo sapiens</i>	HS286810	(+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc	-187 .. -176 -175 .. -186 8 .. 19 20 .. 9	0.988 1.005 0.885 0.932	4.91 3.01 4.21
von Hippel-Lindau (VHL), <i>Homo sapiens</i>	AF016238	(+) gttttggccggcagc (+) gttttggccggcagc (+) gttttggccggcagc	+70 .. +59 -258 .. -269 -28 .. -39	0.810 0.838 0.921	2.22
B-myb , <i>Homo sapiens</i>	HSBMV80NA	(+) gttttggccggcagc	-72 .. -83 -53 .. -42	0.831 0.866	5.50
Nucleolin , <i>Homo sapiens</i>	HSNUCLEO	(+) gttttggccggcagc (+) gttttggccggcagc	-297 .. -308 -256 .. -267	0.966 0.814	2.91
Nucleolin , <i>Oryzotilus griseus</i>	CSNUCLEO	(+) gttttggccggcagc	-296 .. -307	0.973	6.67
Nucleolin , <i>Mus musculus</i>	MMNUCLEO	(+) gttttggccggcagc	-306 .. -317	0.973	1.76

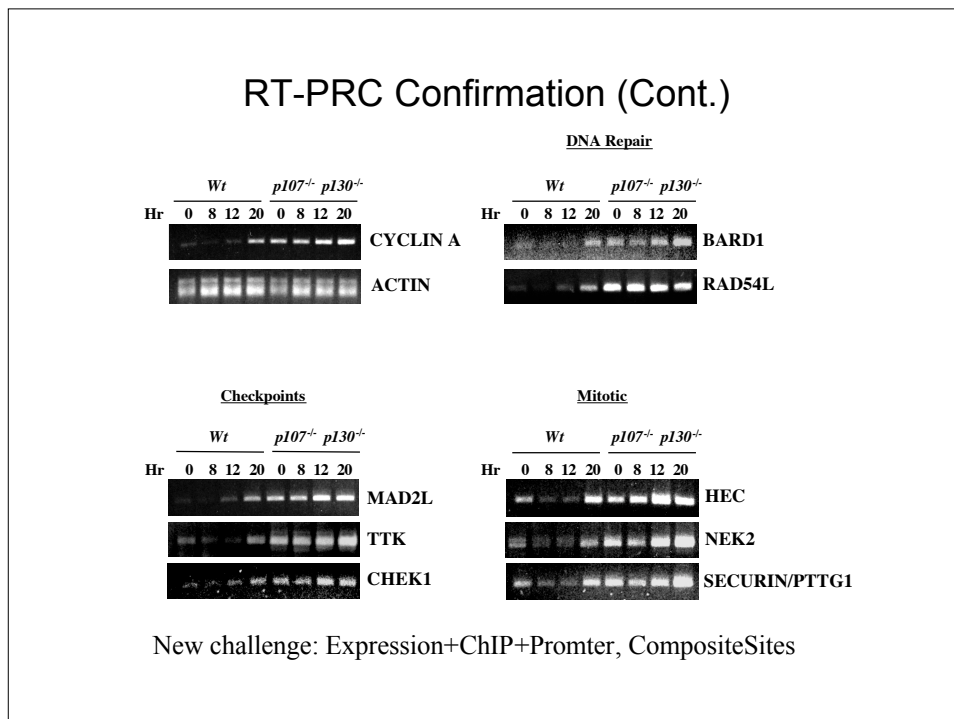
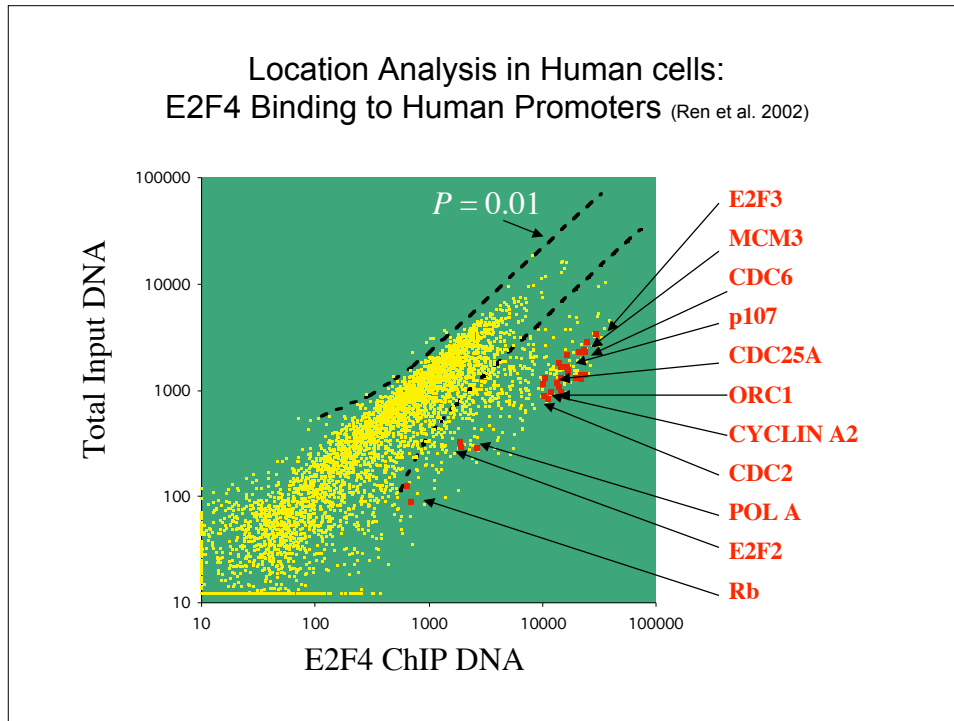
Does genome location analysis work with human cells? — Challenges (In collaboration with Ren Lab UCSD)

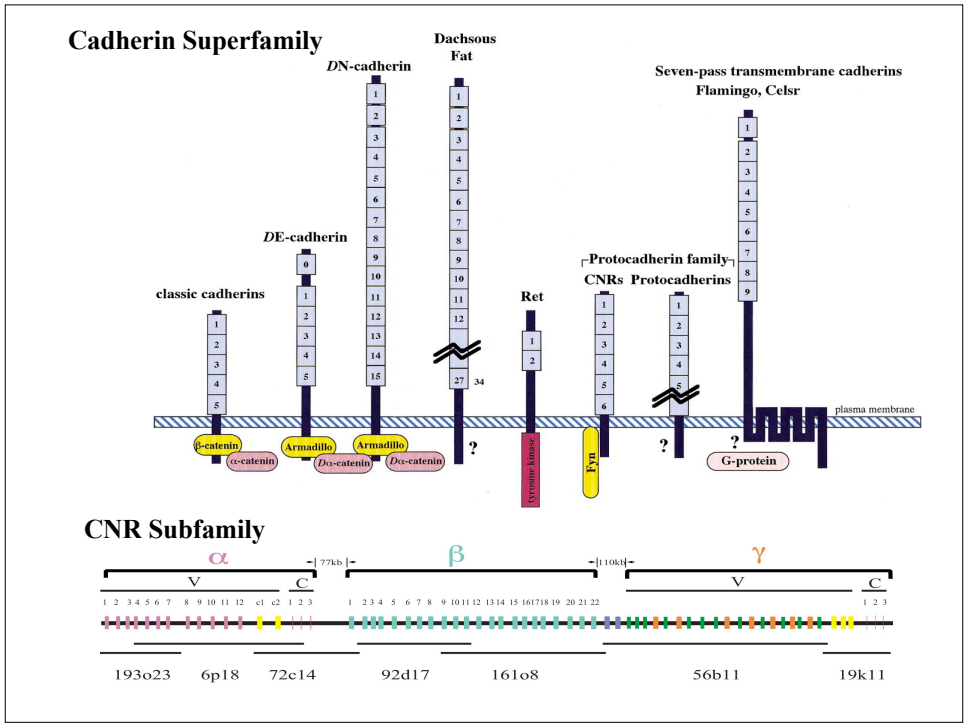
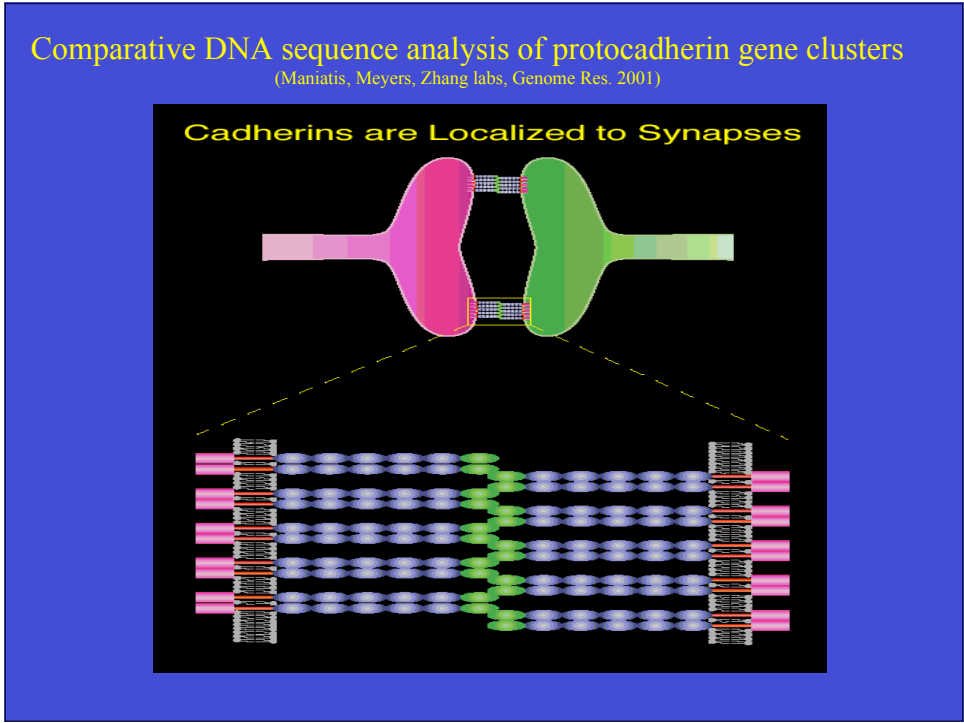
ChIP-chip:

- Genome is 200 times bigger than yeast
- Abundant repetitive sequences
- Annotation of gene structure and function is much less complete
- Many different cell types
- Quality of antibodies



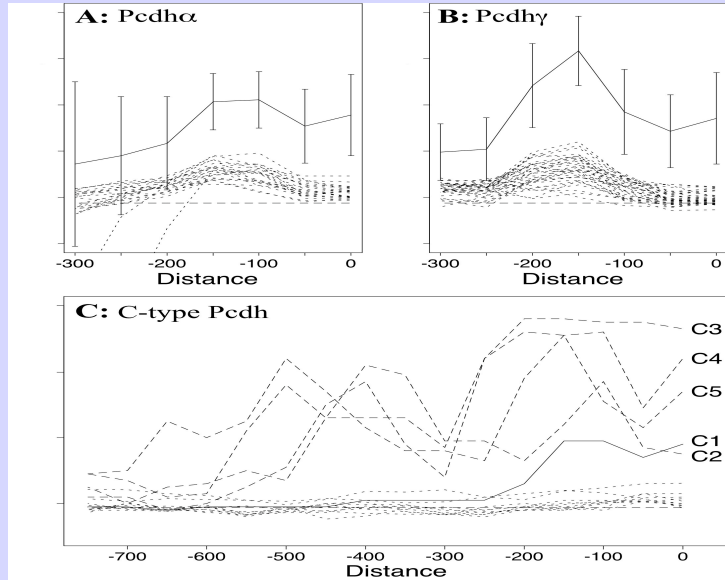
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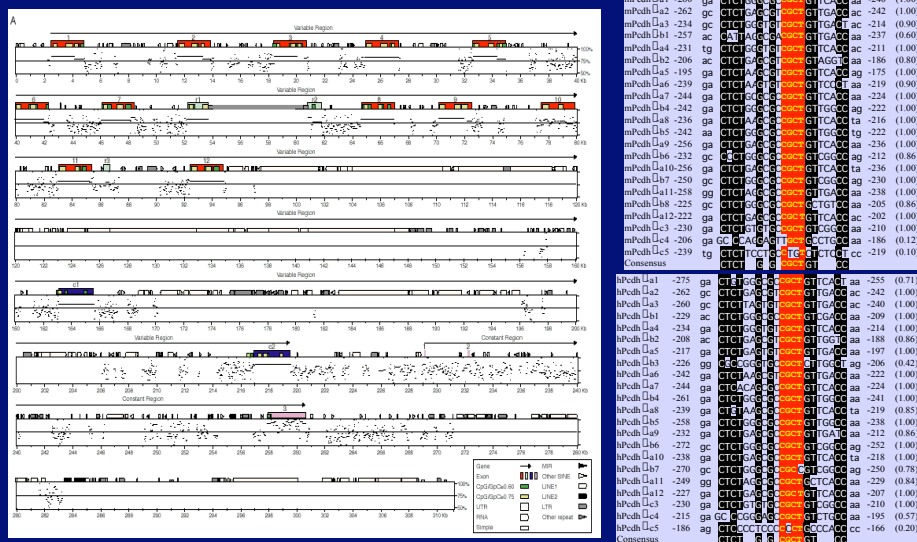


Computational Molecular Biology of Genome Expression and Regulation

Upstream Sequence Similarity



CpG-islands, conserved promoter elements



Pipmaker

Gibbs sampler

Computational Molecular Biology of Genome Expression and Regulation

