

Microbial methane metabolism

Cornelia Welte

Associate Professor

KITP Forum

March 04, 2021

Let me introduce myself

2008 MSc in Biology (Microbiology/Biochemistry)

2011 PhD in Microbiology (Microbial Biochemistry)

2011-14 Postdoc at Bonn U (GER) and RU (NL)

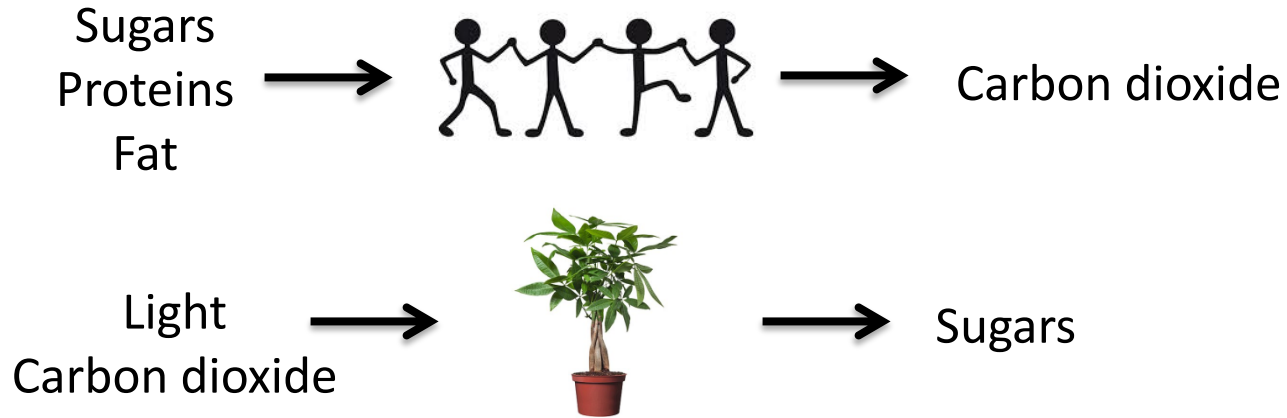
2015-20 Assistant Professor RU

2021- Associate Professor RU

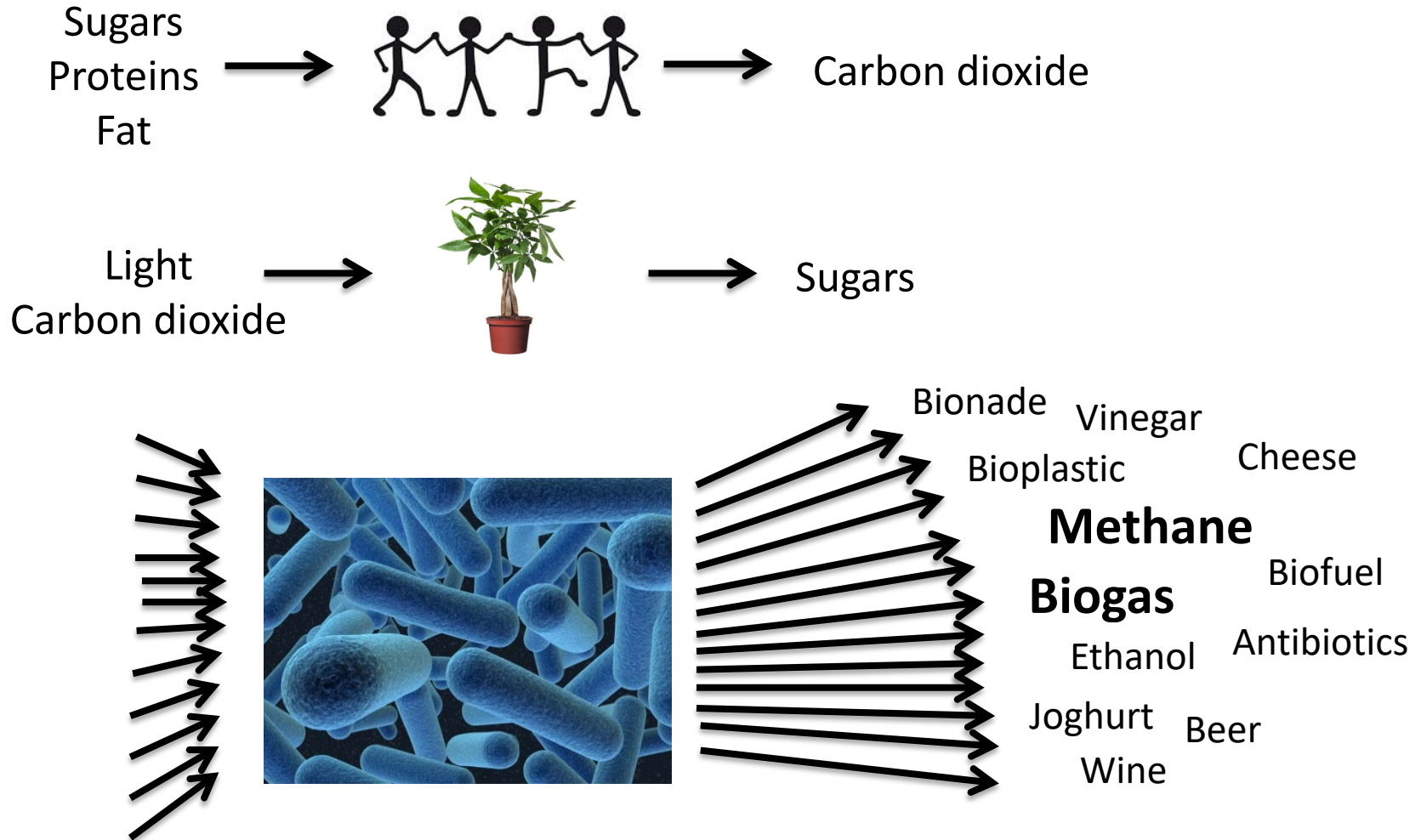
Head of the Microbial Interactions research group

I get excited about anaerobic methane cycling microbes and symbiosis between pest insects and microbes

Let me introduce myself



Let me introduce myself



ARTIS MICROPIA

Microworld

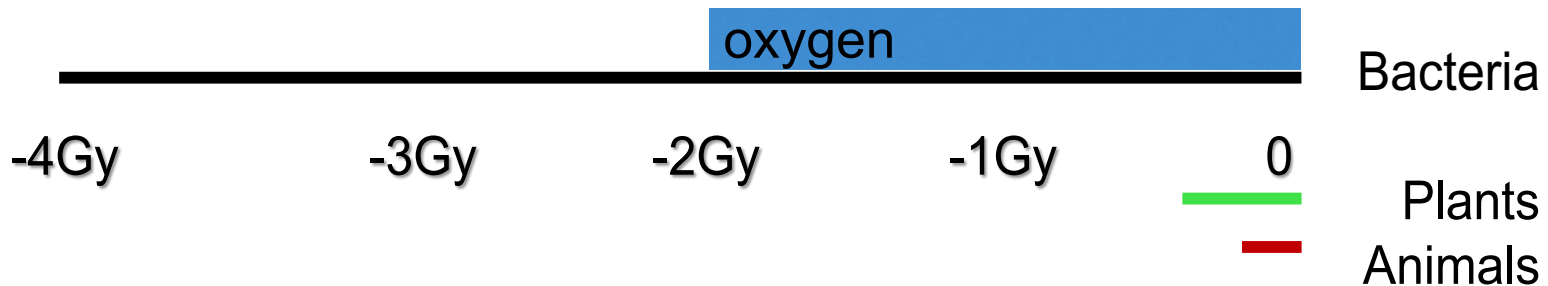
The most powerful life on earth

*When you look from really close, a new world is revealed to you.
More beautiful and spectacular than you could ever have imagined.*

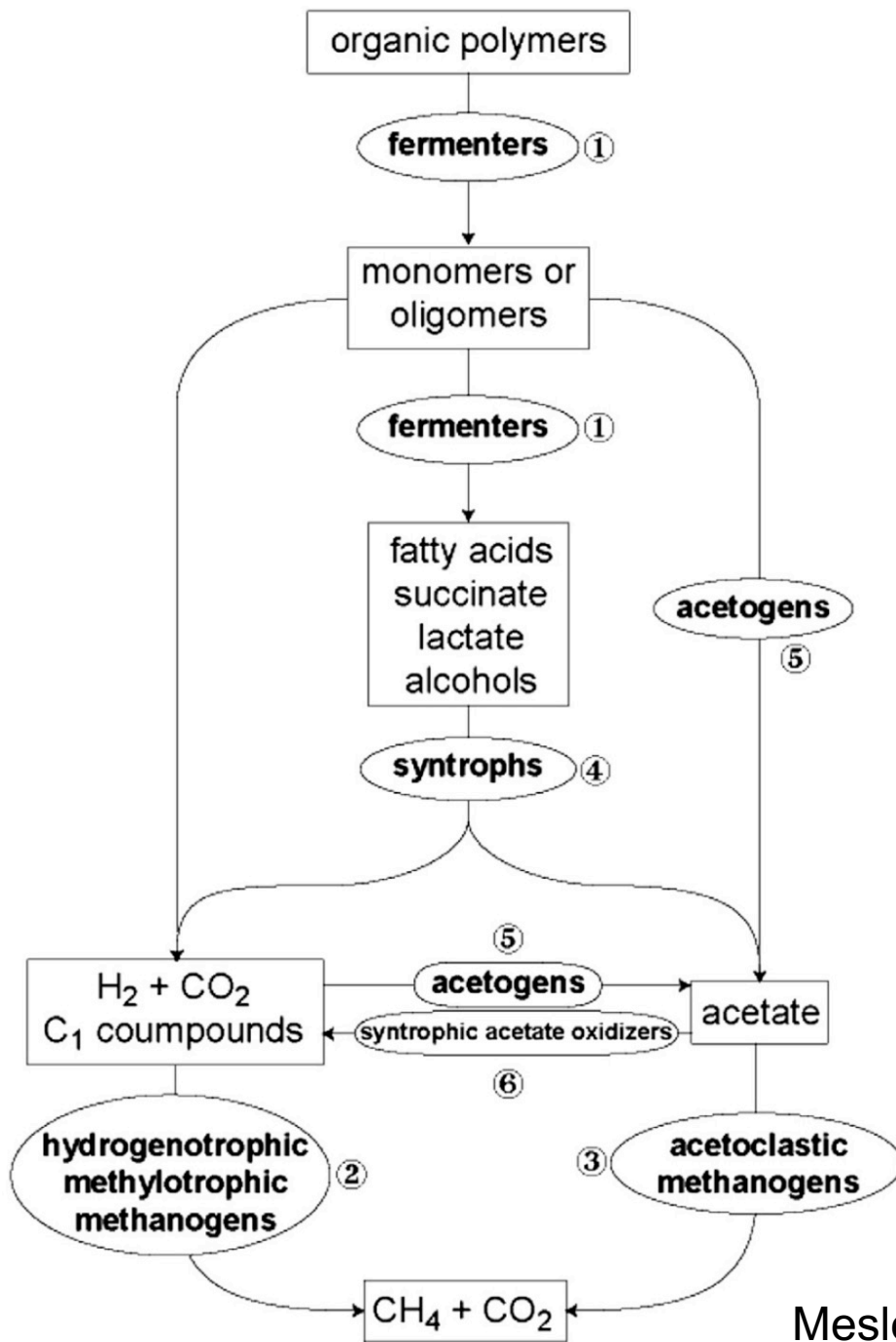
Why care about anaerobic microorganisms?

“The Earth is a microbial planet, on which macro-organisms are recent additions, highly interesting and extremely complex in ways that most microbes are not, but in the final analysis relatively unimportant in a global context.”

Wheelis *et al.* (1998) PNAS 95:11043-11046



**Anaerobic microorganisms had 4 billion years to evolve
A treasure trove for scientists and society!**



The anaerobic food chain

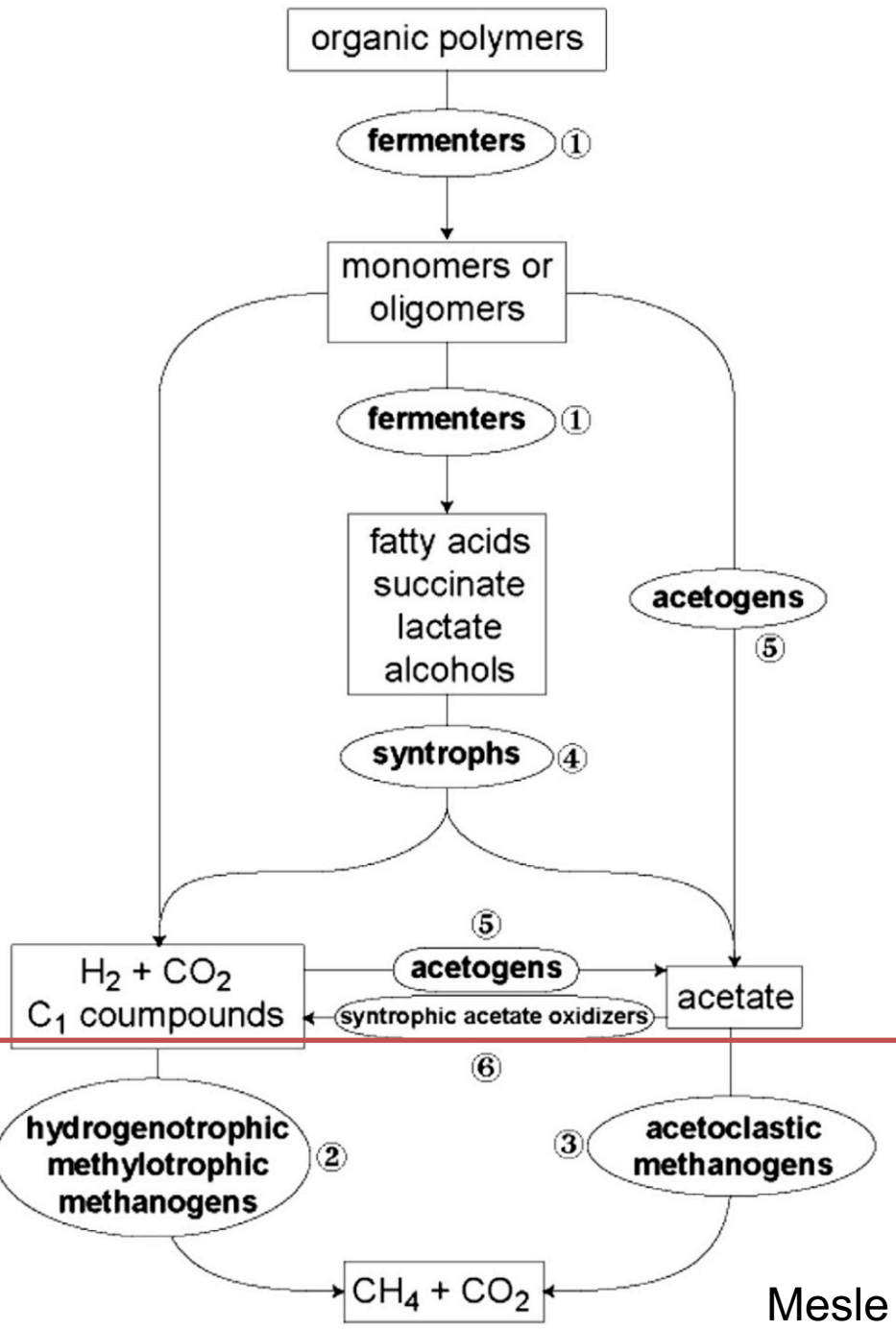
Happening in all kinds of anoxic environments:

- Intestinal systems
- Anaerobic digesters
- Anoxic ocean
- Sediments
- Sub-surface environments

The anaerobic food chain

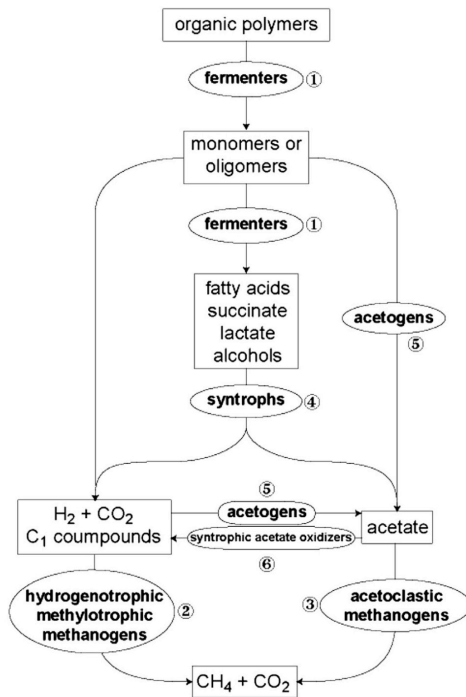
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- Sub-surface environments



Thermodynamics of aerobes vs anaerobes

Anaerobes



Aerobes

Organics + O₂



CO₂

**Aerobes don't have to share
 $\Delta G \lll 0$**

Anaerobes need to cooperate

$\Delta G < 0$

Many live at the thermodynamic limit

Often use "thermodynamic tricks"

Roadmap for today

Part I: thermodynamics of methane generation and what it does (not) tell us

- Required knowledge: ΔG , ATP and pmf, respiration

Part II: the methanogenic metabolic pathway map and an anaerobic thermodynamic trick

Part III (Research Talk):

“How to revert a metabolic pathway and still make a living”

or

“Novel discoveries in anaerobic methane oxidation”

Roadmap for today

Part I: thermodynamics of methane generation and what it does (not) tell us

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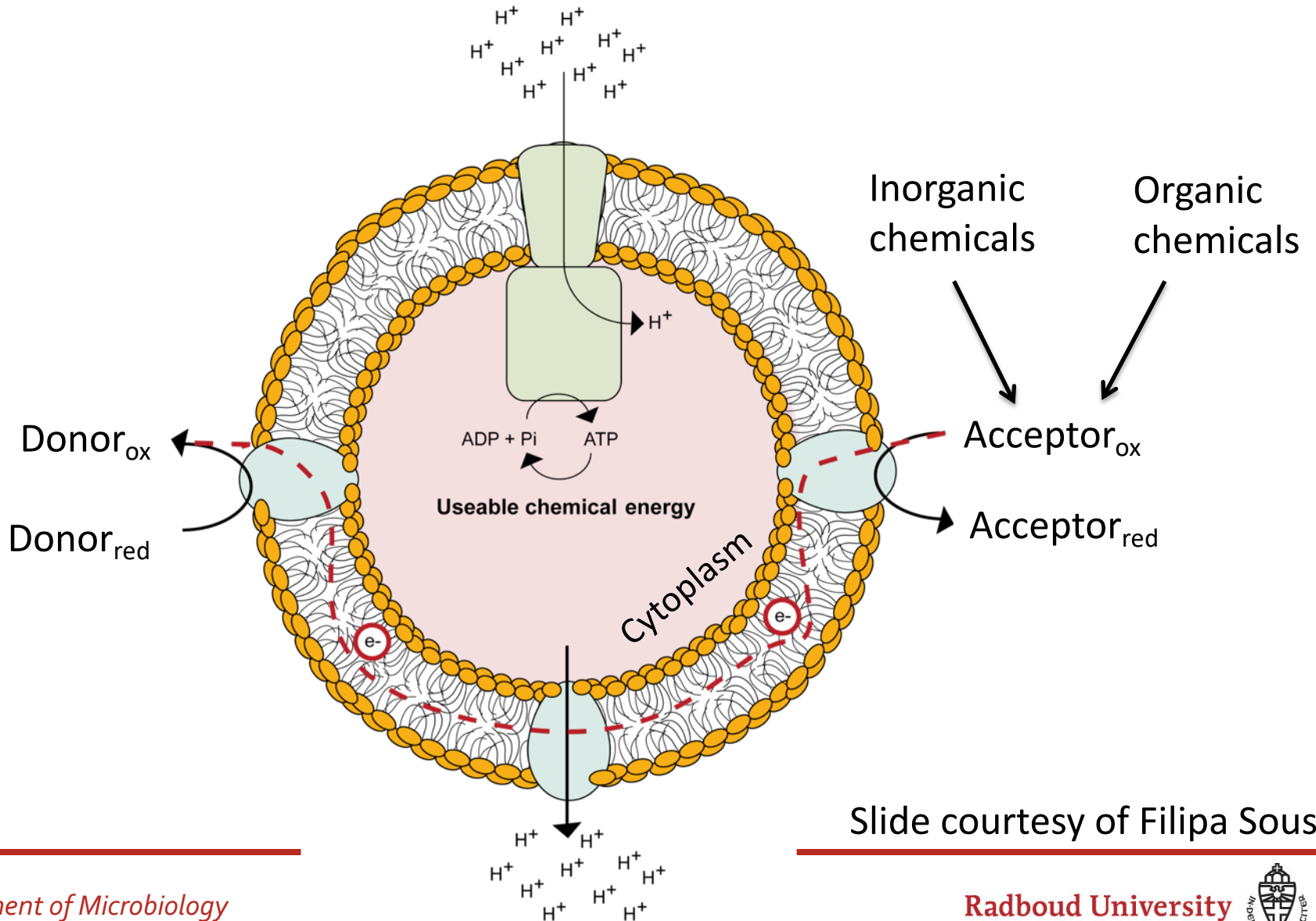
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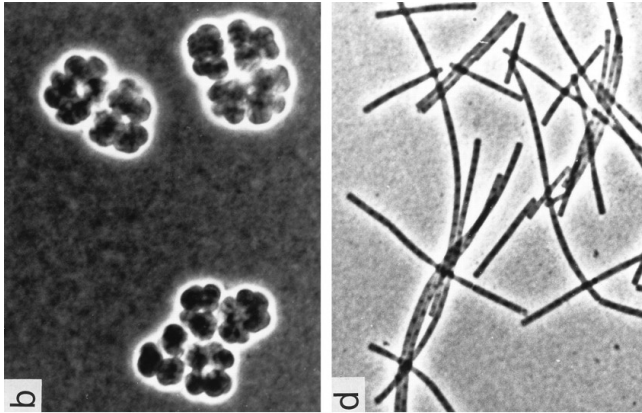
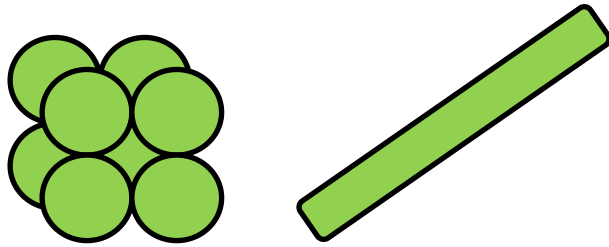
“Novel discoveries in anaerobic methane oxidation”

Principles of respiration



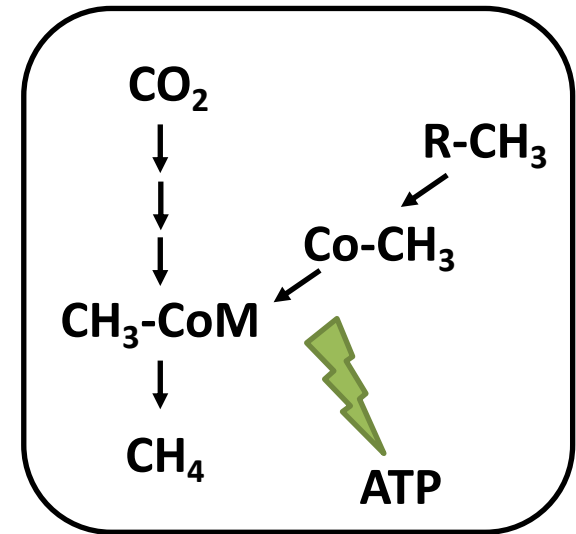
Slide courtesy of Filipa Sousa

Methanogens: ancient (simple?) metabolism



Grosskopf et al 1998

Acetate
Methanol
Methylated amines
Methyl sulfide
 $H_2 + CO_2$
↓
 CH_4

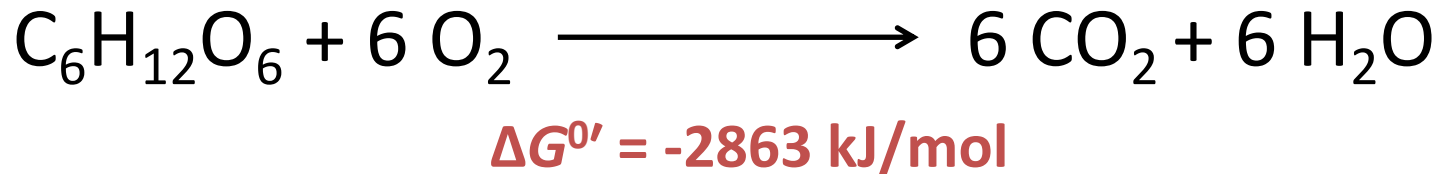


Methanogenesis is the energy conserving pathway of methane generating archaea (methanogens)

Methanogens: ancient metabolism

Table 2 Gibbs free energy values for different methanogenesis substrates

Substrate	Reaction equation	ΔG° (kJ/mol CH ₄)
H ₂ + CO ₂	4 H ₂ + CO ₂ → CH ₄ + 2 H ₂ O	- 131 (a)
HCOO ⁻	4 HCOO ⁻ + 4 H ⁺ → CH ₄ + 3 CO ₂ + 2 H ₂ O	- 145 (a)
CH ₃ CH ₂ OH + CO ₂	2 CH ₃ CH ₂ OH + CO ₂ → 2 CH ₃ COOH + CH ₄	- 121 (e)
H ₂ + CH ₃ OH	CH ₃ OH + H ₂ → CH ₄ + H ₂ O	- 113 (e)
CH ₃ OH + CH ₃ CH ₂ OH	2 CH ₃ OH + CH ₃ CH ₂ OH → 2 CH ₄ + H ₂ O + CH ₃ COOH	- 100 (b)
CH ₃ CHOHCH ₃ + CO ₂	4 CH ₃ CHOHCH ₃ + HCO ₃ ⁻ + H ⁺ → 4 CH ₃ COCH ₃ + CH ₄ + 3 H ₂ O	- 37 (c)
CH ₃ OH	4 CH ₃ OH → CO ₂ + 3 CH ₄ + 2 H ₂ O	- 107 (a)
CH ₃ -COOH	CH ₃ COOH → CO ₂ + CH ₄	- 36 (a)
CH ₃ -SH (CH ₃ -S-R)	4 CH ₃ SH + 3 H ₂ O → 3 CH ₄ + HCO ₃ ⁻ + 4 HS ⁻ + 5 H ⁺	- 49 (b)
Betaine (CH ₃ -N-R)	4 (CH ₃) ₃ N ⁺ CH ₂ COO ⁻ + 2 H ₂ O → 4 (CH ₃) ₂ N ⁺ CH ₂ COO ⁻ + 3 CH ₄ + CO ₂	- 241 (c)
Choline (CH ₃ -N-R)	4 (CH ₃) ₃ N ⁺ CH ₂ CH ₂ OH + 6 H ₂ O → 4 H ₂ NCH ₂ CH ₂ OH + 9 CH ₄ + 3 CO ₂ + 4 H ⁺	- 63 (d)
Trimethylamine (CH ₃ -N-R)	4 (CH ₃) ₃ N + 6 H ₂ O + 4 H ⁺ → 4 NH ₄ ⁺ + 9 CH ₄ + 3 CO ₂	- 31 (d)
2-methoxyphenol (CH ₃ -O-R)	4 2-methoxyphenol + 2 H ₂ O → 4 2-hydroxyphenol + CO ₂ + 3 CH ₄	- 90 (f)



Three statements to think about:

Acetate-dependent methanogenesis has the **lowest Gibb's free energy change** of all methanogenic processes, yet it is the most relevant methanogenic process on our planet.

Low Gibb's free energy change of a microbial metabolic process **does not** equal low growth rate (= speed of cell division) of a microorganism.

The Gibb's free energy change **does not** usually determine the substrate threshold concentration, the minimal required concentrations for a microbe to grow.

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Thermodynamics provides you with a yes/no answer:

If $\Delta G < 0$, a metabolic process can yield ATP

If $\Delta G > 0$, a metabolic process cannot yield ATP

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However, a **minimal ΔG is required**, equaling the amount to transport 1 H^+ across the cytoplasmic membrane (we'll get there in a minute)

Three statements to think about:

Low Gibb's free energy change of a microbial metabolic process **does not** equal low growth rate (= speed of cell division) of a microorganism.

Aceticlastic methanogenesis: $\Delta G = -36 \text{ kJ/mol}$
doubling time $\sim 24 \text{ h}$

Anaerobic methane oxidation with nitrate:
 $\Delta G = -523 \text{ kJ/mol}$
doubling time $\sim 10 \text{ days}$

Three statements to think about:

The Gibb's free energy change **does not** usually determine the substrate threshold concentration, the minimal required concentrations for a microbe to grow.

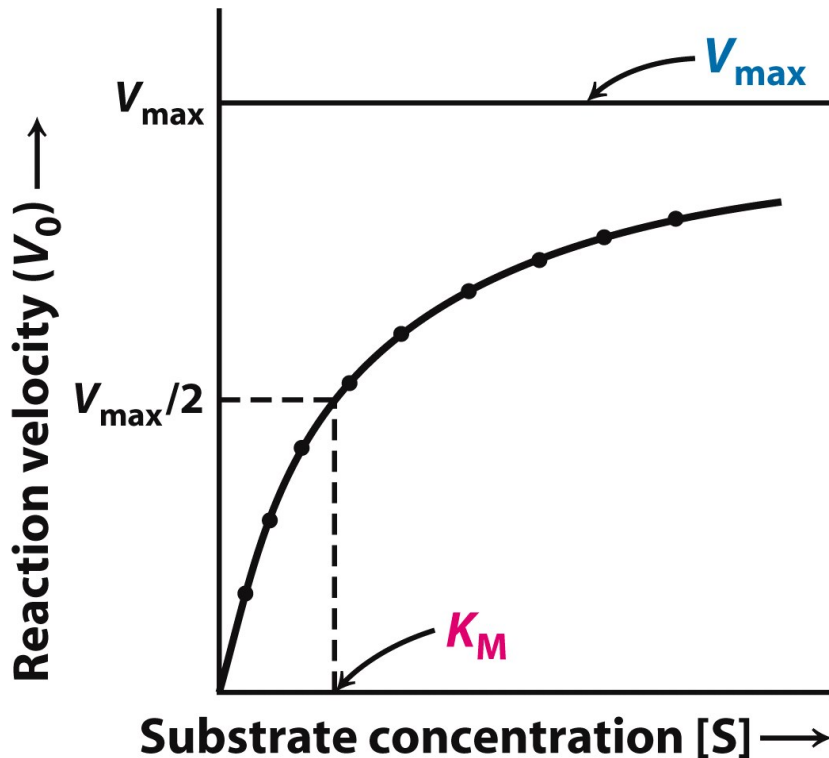
Aceticlastic *Methanosarcina*: grows $> 1\text{mM}$ acetate

Aceticlastic *Methanosaeta*: grows $< 1\text{ mM}$ acetate

What determines the substrate threshold?

Aceticlastic *Methanosarcina*: grows > 1mM acetate

Aceticlastic *Methanosaeta*: grows < 1 mM acetate



Contributing factors:

- Enzyme kinetics
- Enzymatic repertoire
- Substrate availability (gasses, membrane transport)

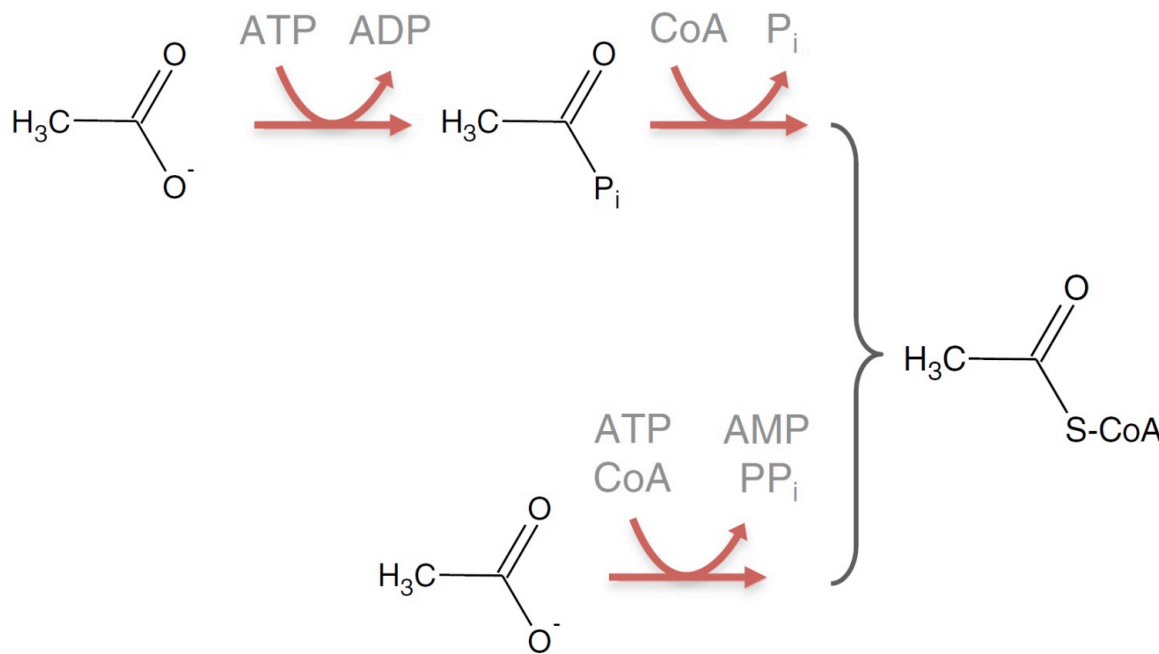
Figure 8.11
Biochemistry, Seventh Edition
© 2012 W. H. Freeman and Company

What determines the substrate threshold?

Methanosarcina: > 1mM acetate, uses 1 ATP for acetate activation

Methanosaeta: < 1 mM acetate, uses 2 ATP for acetate activation

Methanosarcina



**YOU CANNOT HAVE IT
ALL**

Metabolic processes are **either** highly specific (low substrate threshold) **or** highly efficient (more ATP gain)

Methanosaeta

Three statements to think about:

Acetate-dependent methanogenesis has the **lowest Gibb's free energy change** of all methanogenic processes, yet it is the most relevant methanogenic process on our planet.

Thermodynamics provides you with a yes/no answer:

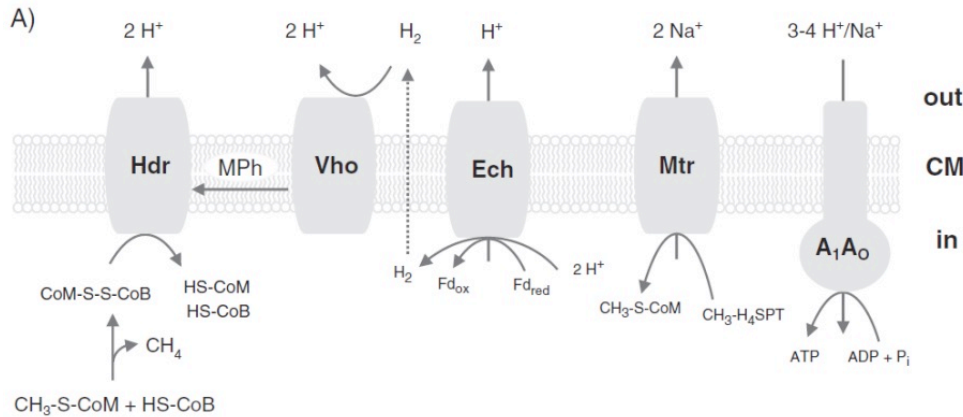
If $\Delta G < 0$, a metabolic process can yield ATP

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The minimal energy quantum

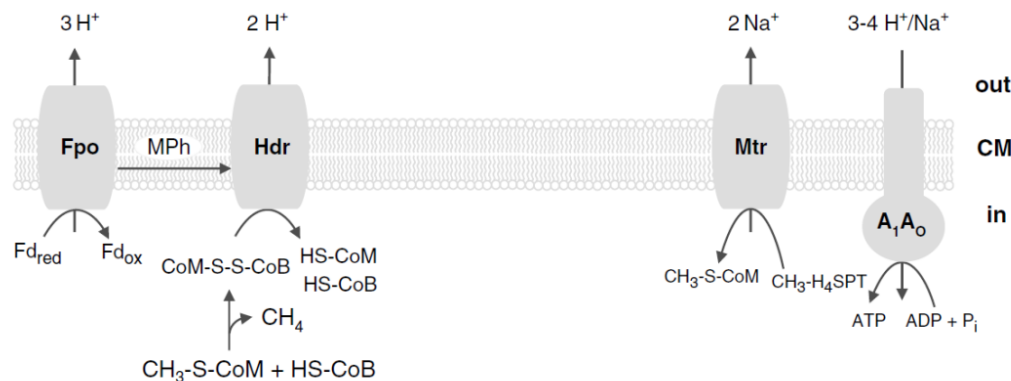
Methanosarcina



Aceticlastic respiratory chains:

7 H⁺ per acetate translocated
 Equals about 2 ATP
 Minus 1 ATP invested
 = 1 ATP net gain

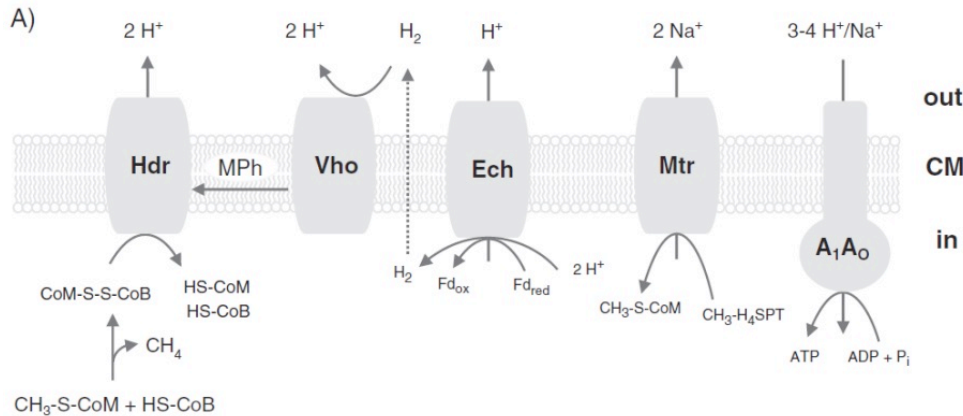
Methanosaeta



7 H⁺ per acetate translocated
 Equals about 2 ATP
 Minus 2 ATP invested
 = no net gain?!

The minimal energy quantum

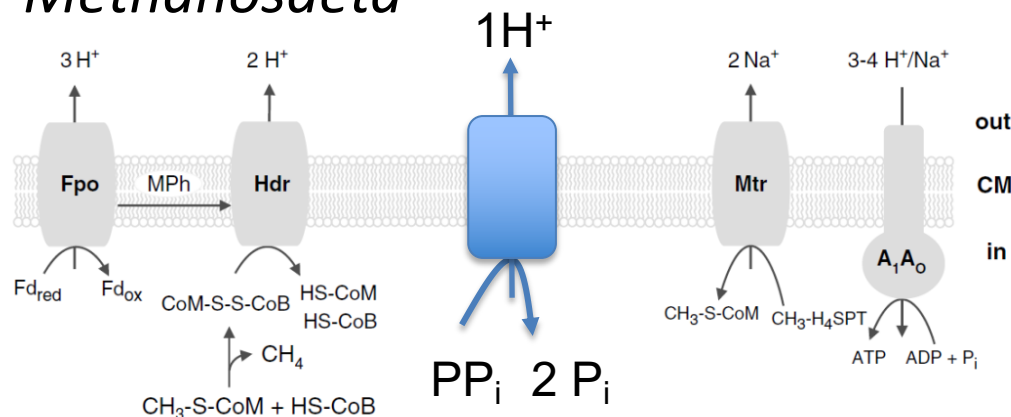
Methanosarcina



Aceticlastic respiratory chains:

7 H⁺ per acetate translocated
 Equals about 2 ATP
 Minus 1 ATP invested
 = 1 ATP net gain

Methanosaeta



7 H⁺ per acetate translocated
 Equals about 2 ATP + 1 H⁺
 Minus 2 ATP invested
 = 1 H⁺ net gain
THE LIMIT OF LIFE

Take home points (part I) & questions

- Thermodynamics is important
- Thermodynamics is not directly quantitative
- Enzyme kinetics determines substrate thresholds
- High substrate affinity and high efficiency are mutually exclusive
- The minimal energy quantum is one translocated ion (H^+/Na^+)

Roadmap for today

Part I: thermodynamics of methane generation and what it does (not) tell us

Part II: the methanogenic metabolic pathway map and an anaerobic thermodynamic trick

➤ **Required knowledge:** ΔE and E^0 , reduction/oxidation

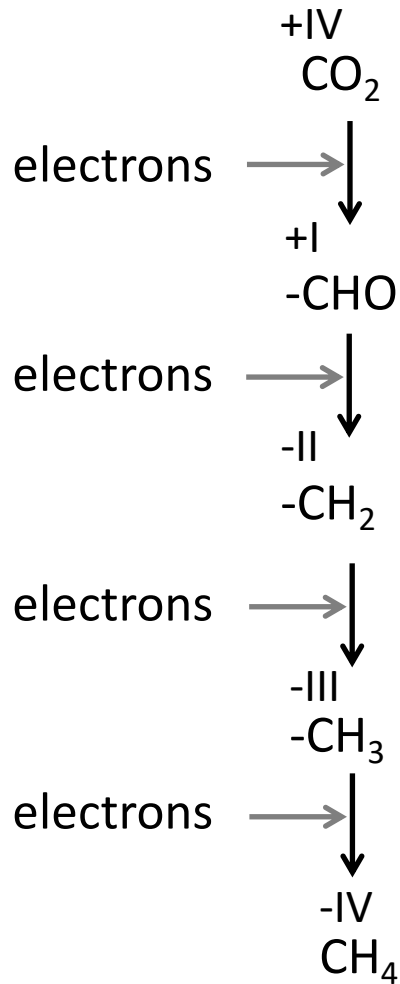
Part III (Research Talk):

“How to revert a metabolic pathway and still make a living”

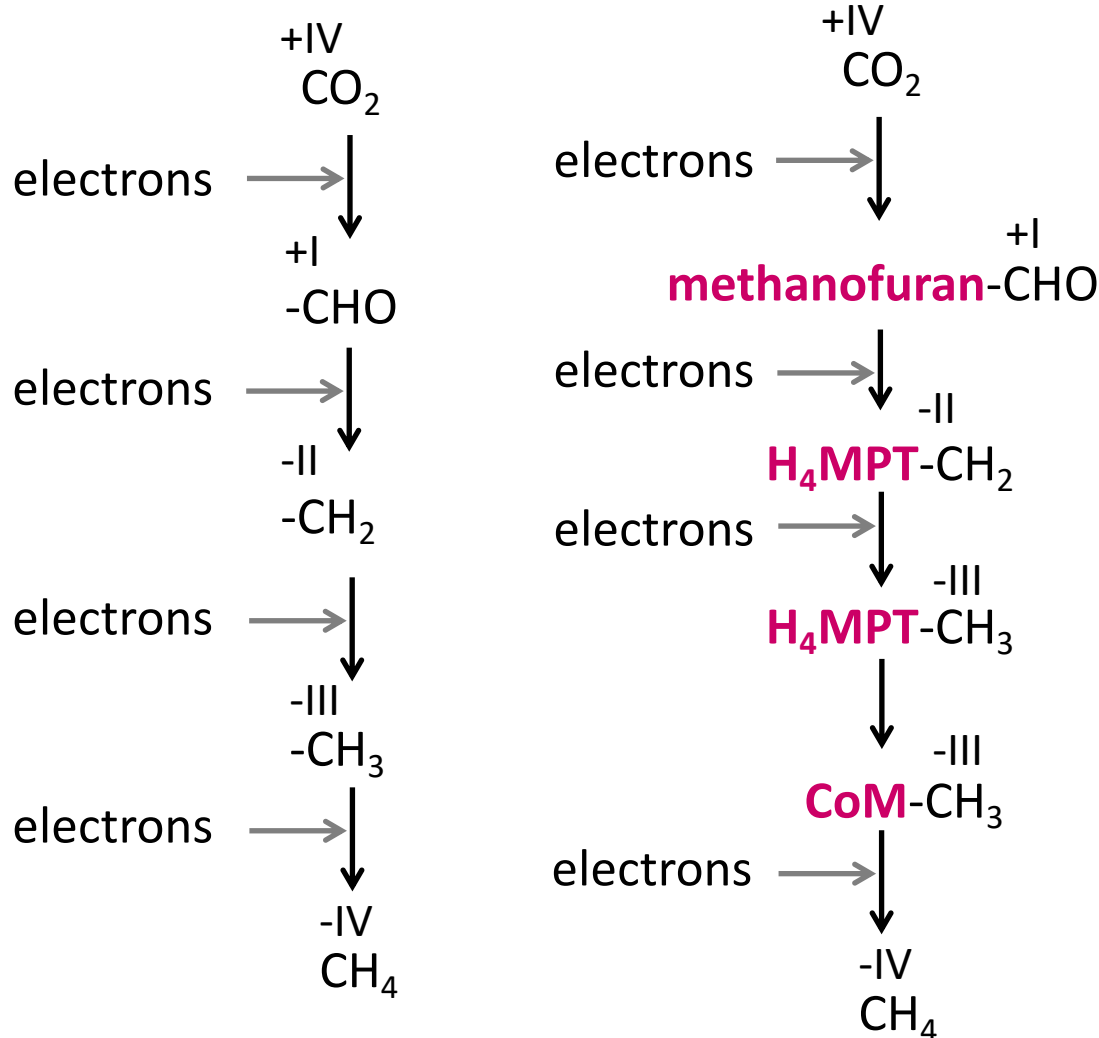
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“**Novel discoveries in anaerobic methane oxidation**”

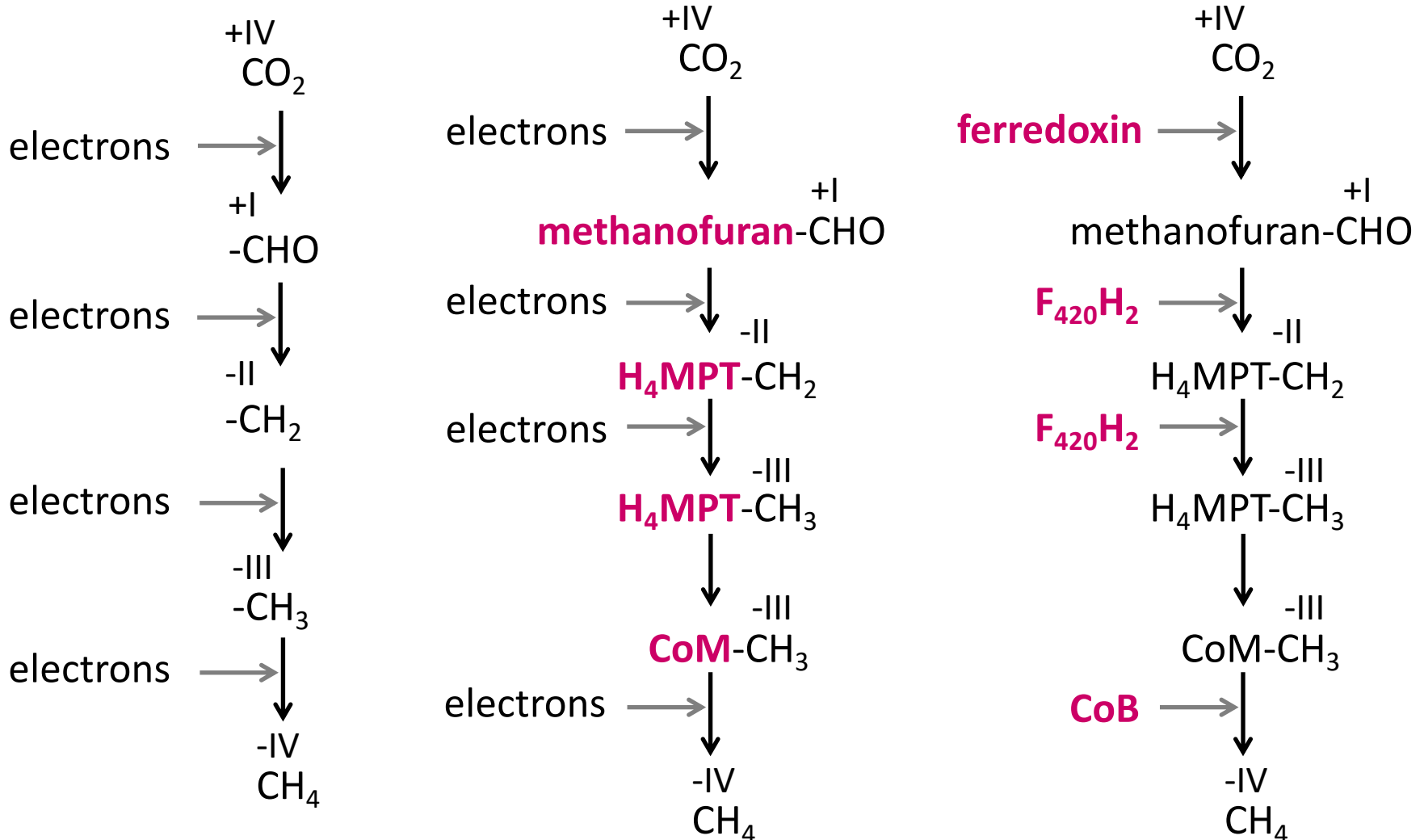
The methanogenic pathway (from H₂+CO₂)



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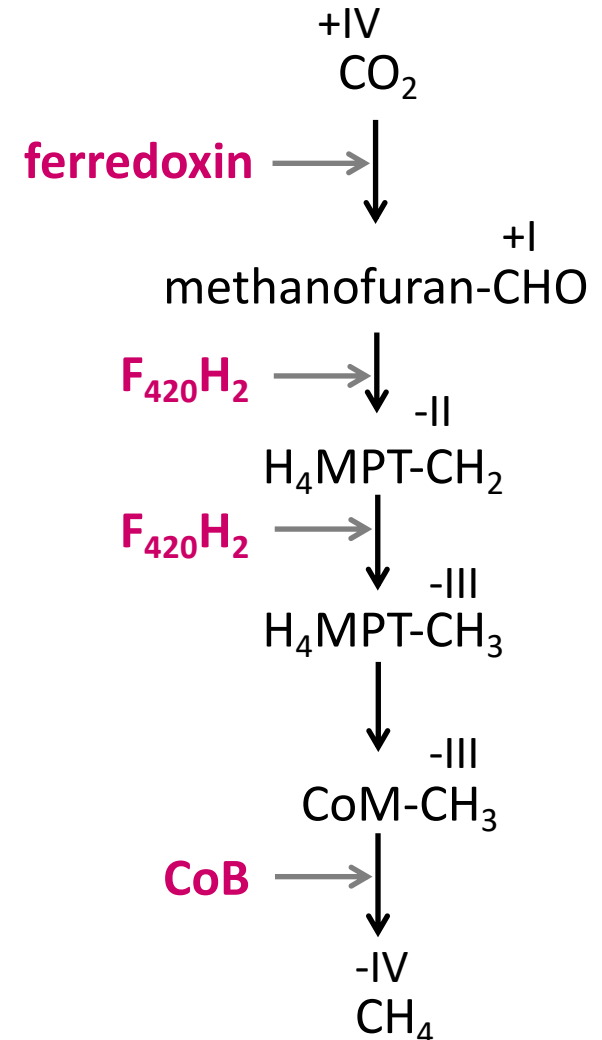
A few key questions:

Why are different cofactors used, and how are they regenerated?

How is energy conserved?

What is limiting the rate of methanogenesis?

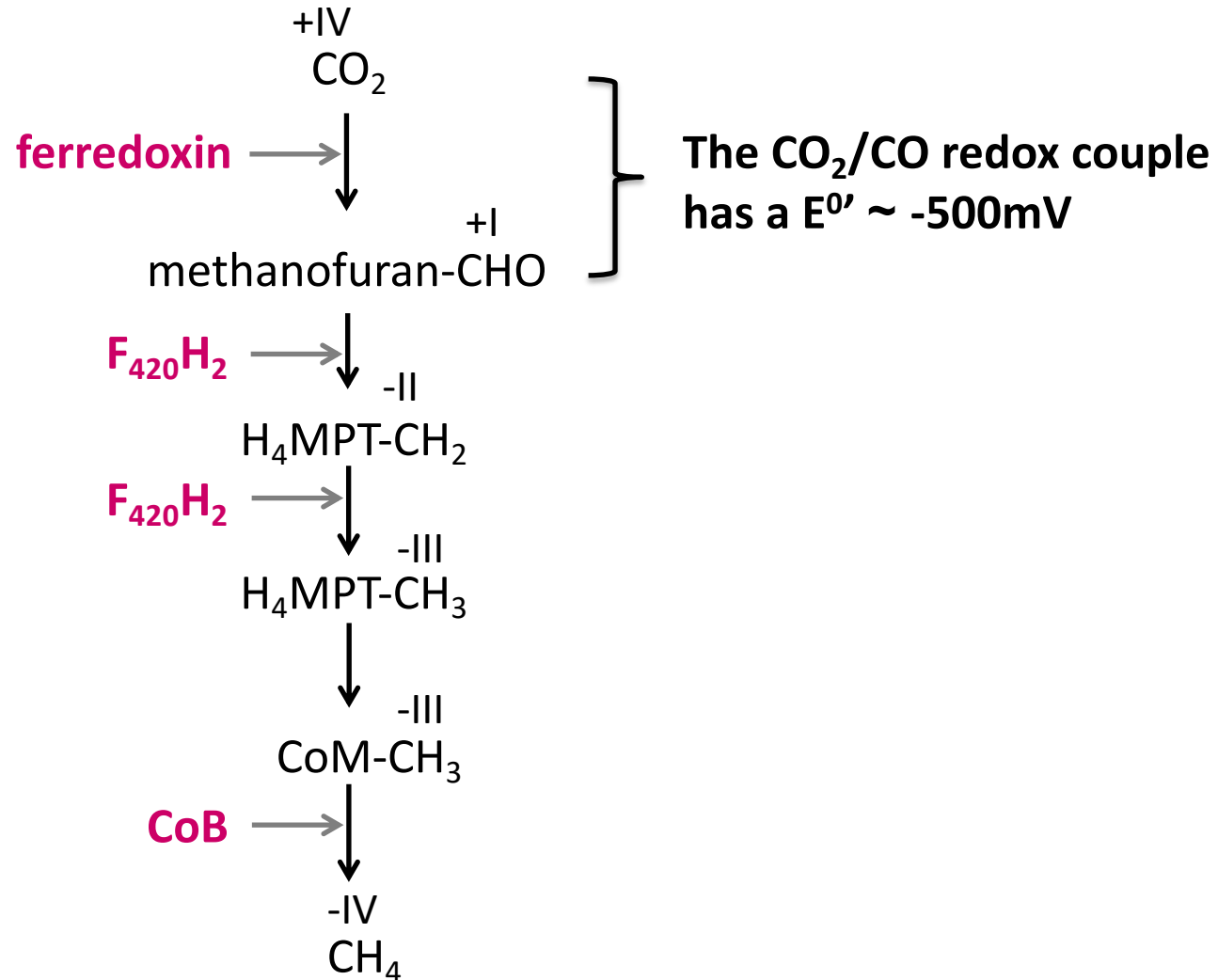
How can CO₂ be fixed without the use of ATP?



The methanogenic pathway (from H₂+CO₂)

The E^{0'} of ferredoxin is ~ -500mV

(if NADH (-320mV) is used, ATP is required to drive this reaction)



The methanogenic pathway (from H₂+CO₂)

The E^{0'} of ferredoxin is ~ -500mV

ferredoxin

+IV
CO₂

+I
methanofuran-CHO

The CO₂/CO redox couple has a E^{0'} ~ -500mV

$$E^{0'}(F_{420}H_2/F_{420}) = -360mV$$

F₄₂₀H₂

-II

H₄MPT-CH₂

F₄₂₀H₂

-III

H₄MPT-CH₃



-III

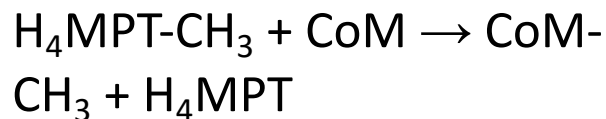
CoM-CH₃

CoB

-IV

CH₄

The only energy conserving reaction in hydrogenotrophic methanogenesis is the methyl transferase reaction:



$$\Delta G^{0'} = -30kJ/mol$$

The methanogenic pathway (from H₂+CO₂)

The E⁰ of ferredoxin is ~ -500mV

ferredoxin

+IV
CO₂

+I
methanofuran-CHO

The CO₂/CO redox couple has a E⁰ ~ -500mV

E⁰(F₄₂₀H₂/F₄₂₀)
= -360mV

F₄₂₀H₂

-II

H₄MPT-CH₂

F₄₂₀H₂

-III

H₄MPT-CH₃

2 Na⁺_{out}

-III

CoM-CH₃

E⁰(CoB/CoM-CoB)
= -120mV

CoB

CoM-S-S-CoB

-IV
CH₄

Methyl-CoM reductase
Most abundant enzyme in
the anaerobic world

The methanogenic pathway (from H₂+CO₂)

The E^{0'} of ferredoxin
is ~ -500mV

+IV
CO₂

CO₂/CO redox couple
E^{0'} ~ -500mV

A few key questions:

**Why are different cofactors used,
and how are they regenerated?**

E^{0'}(F
= -36

How is energy conserved?

**What is limiting the rate of
methanogenesis?**

E^{0'}(CoB
= -120mv

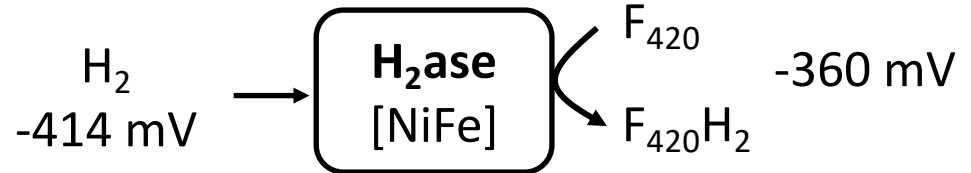
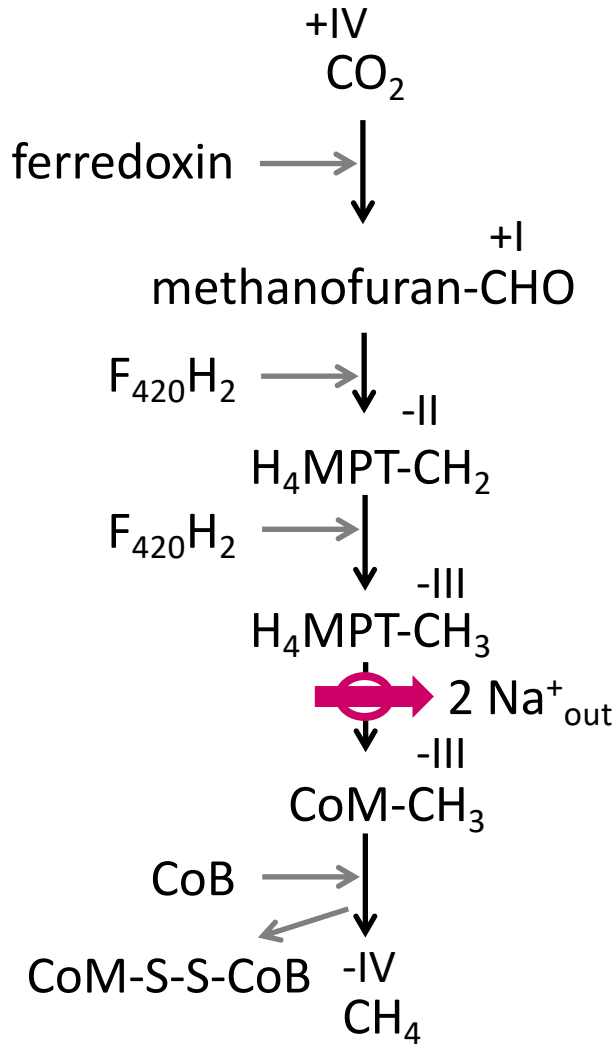
**How can CO₂ be fixed without the
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-IV
CH₄

Methyl-CoM reductase
Most abundant enzyme in
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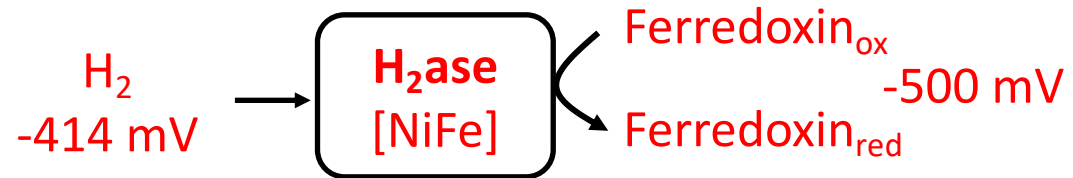
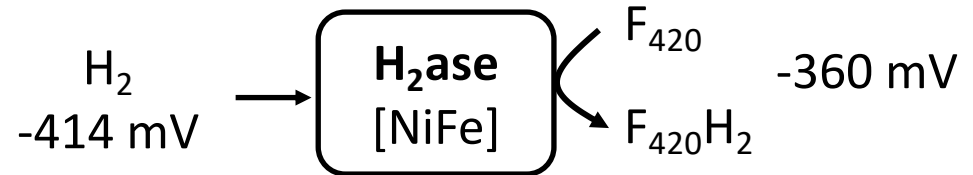
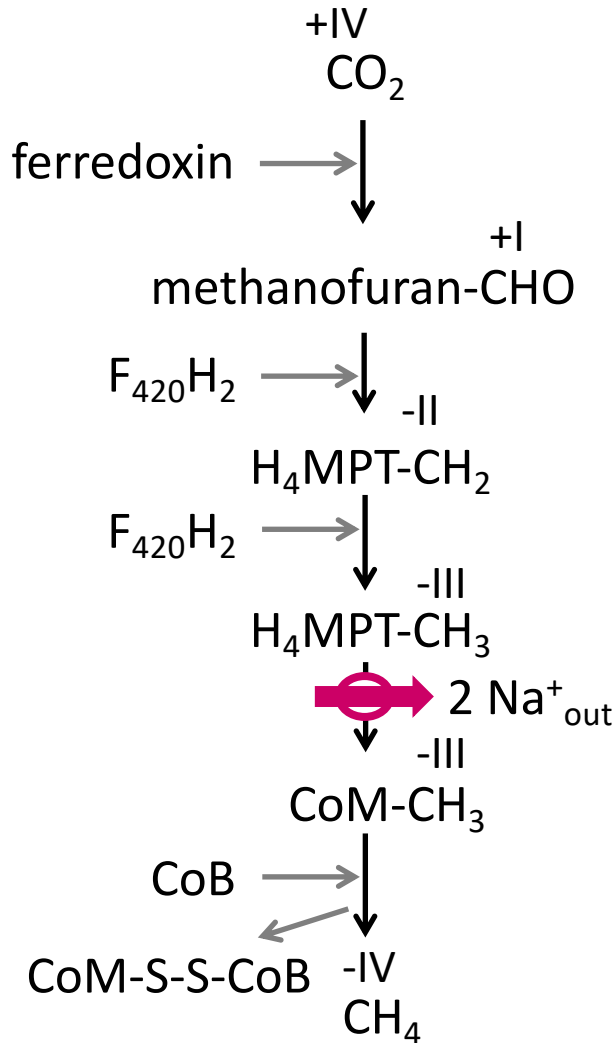
The methanogenic pathway (from H₂+CO₂)

Two hydrogenases regenerate cofactors



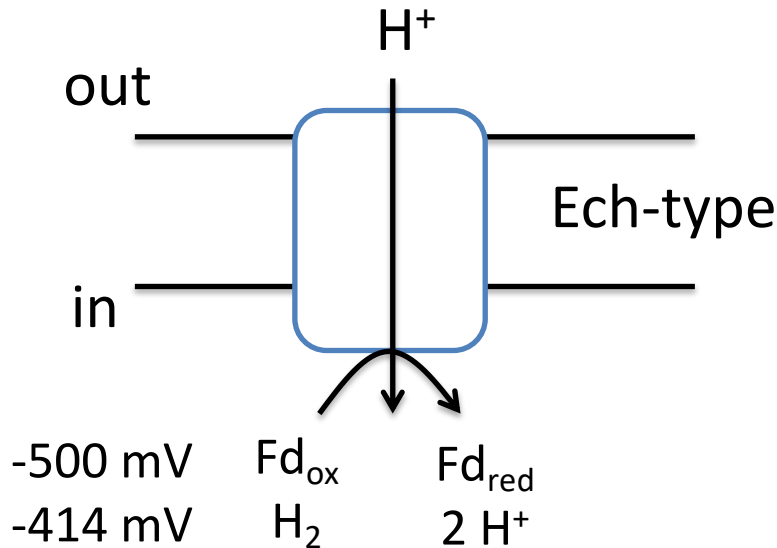
The methanogenic pathway (from H₂+CO₂)

Two hydrogenases regenerate cofactors



Thermodynamically unfavorable!!

Reducing ferredoxin with H₂ at the membrane

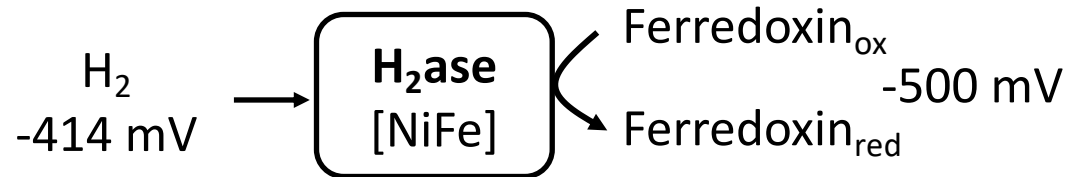


“Energy conserving hydrogenase”
[originally: *E. coli* hydrogenase type 3]

Not an option for methanogenesis from H₂+CO₂:

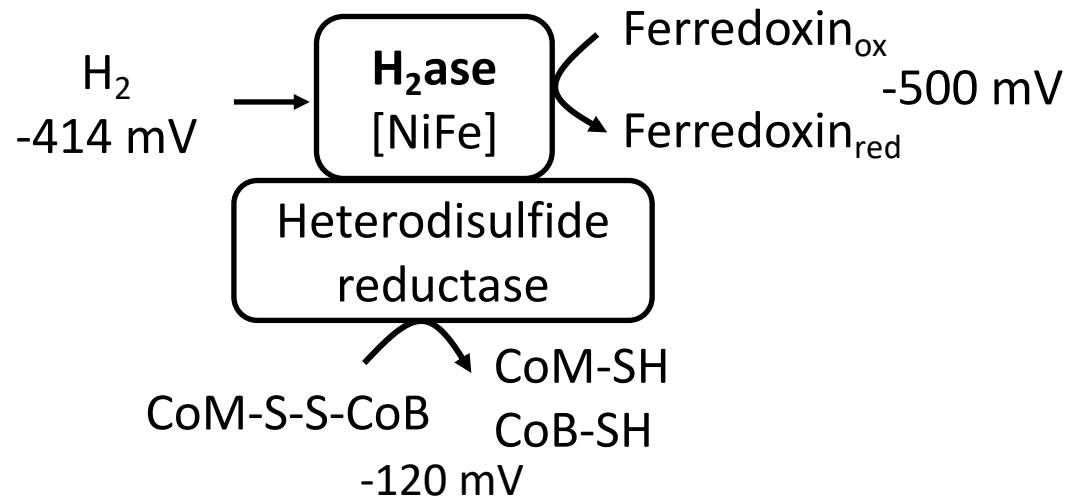
- Only 1-2 Na⁺ translocated per CH₄ produced
- 1-2 H⁺ would be needed for ferredoxin reduction
- No net energy gain of the metabolism

Reducing ferredoxin with H₂ in the cytoplasm



- **Thermodynamically unfavorable**
- **Input of ATP not possible: only 1-2 Na⁺ translocated per ferredoxin required**
- **How can these methanogens obtain reduced ferredoxin?**

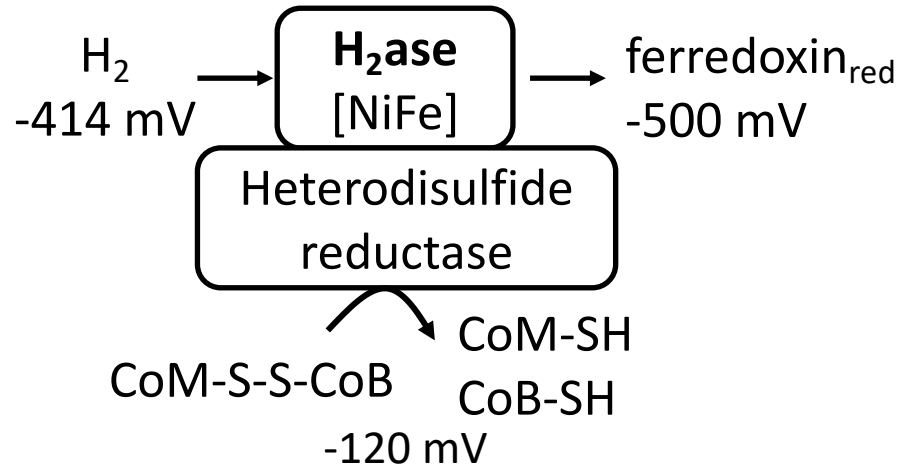
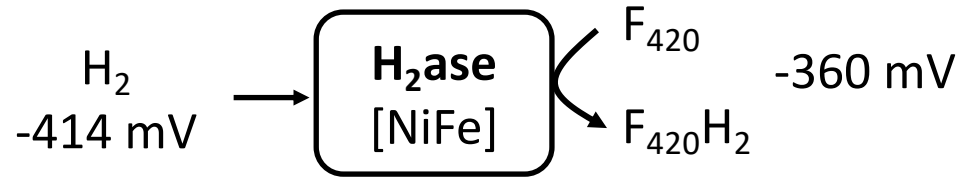
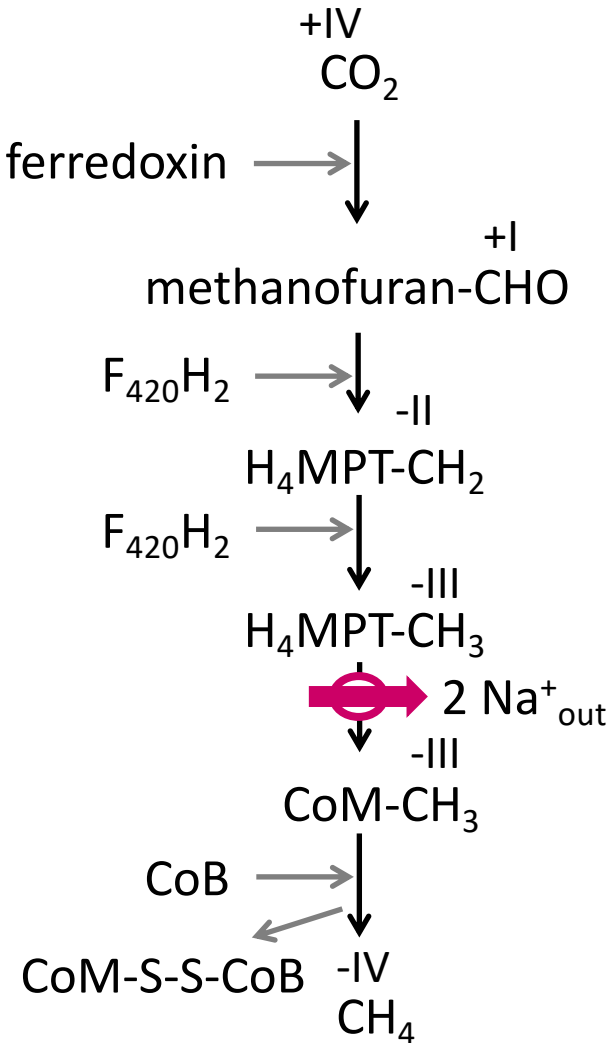
Reducing ferredoxin with H₂ in the cytoplasm



By using a thermodynamic trick: electron bifurcation
the exergonic reduction of CoM-S-S-CoB drives the
endergonic reduction of ferredoxin

The methanogenic pathway (from H_2+CO_2)

Two hydrogenases regenerate cofactors



Take home points (part II) & questions

- Cofactors need to be regenerated (closed loop)
- After closing the loop, there needs to be a net gain of ATP/translocated ions
- Redox potentials of cofactors determine the way in which they can be regenerated
- Individual enzymes can be bottlenecks for entire pathways

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Novel discoveries in anaerobic methane oxidation

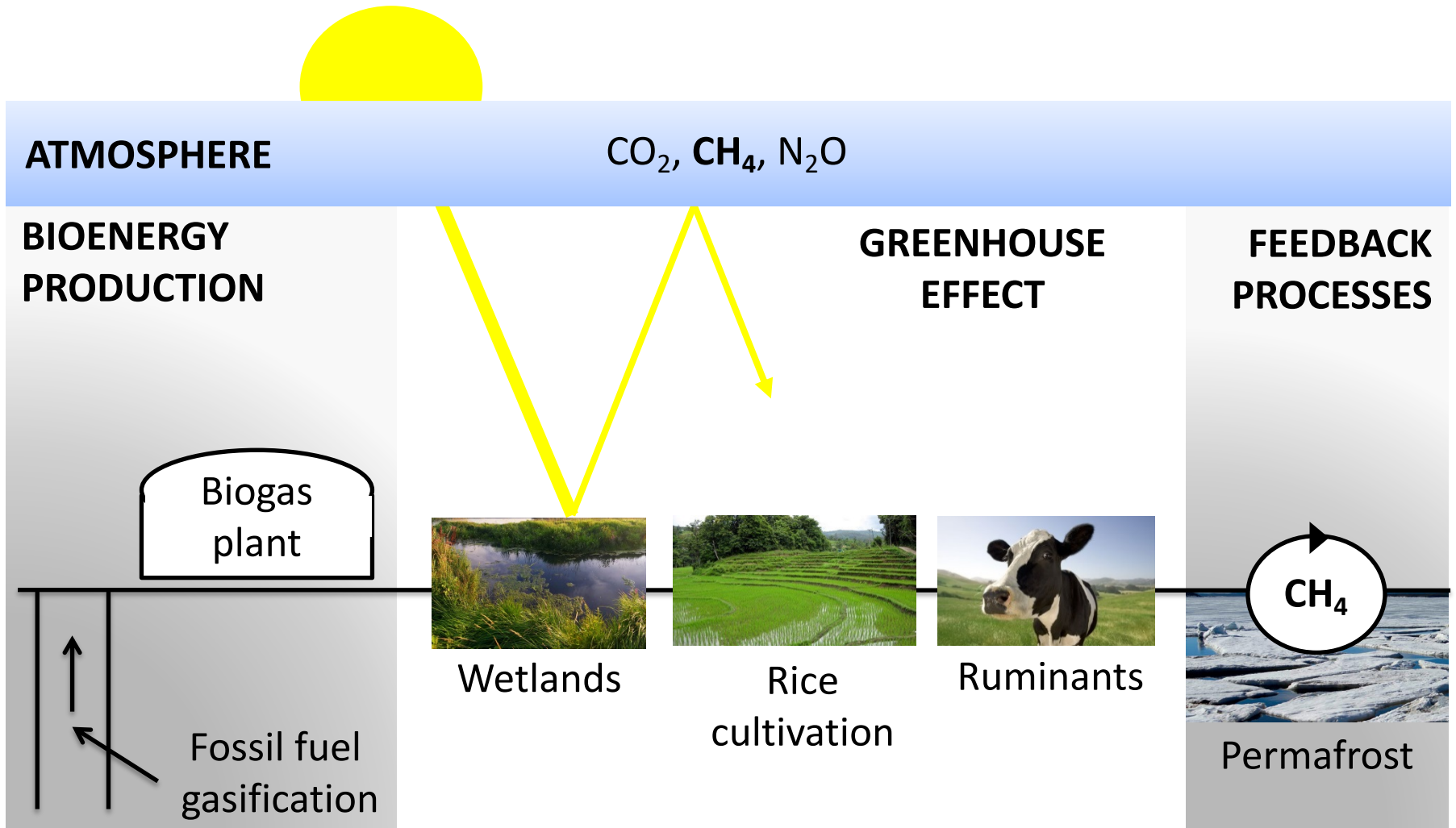
Cornelia Welte

Associate Professor

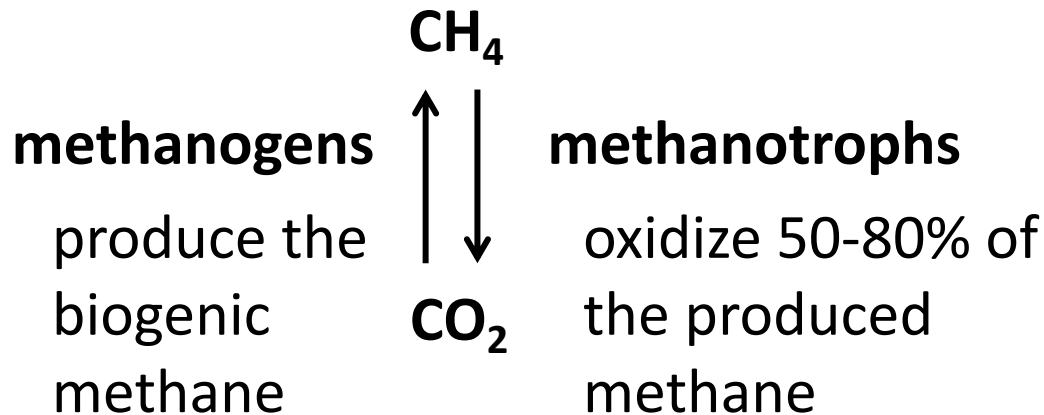
KITP Forum

March 04, 2021

The fascinating world of methane microbes

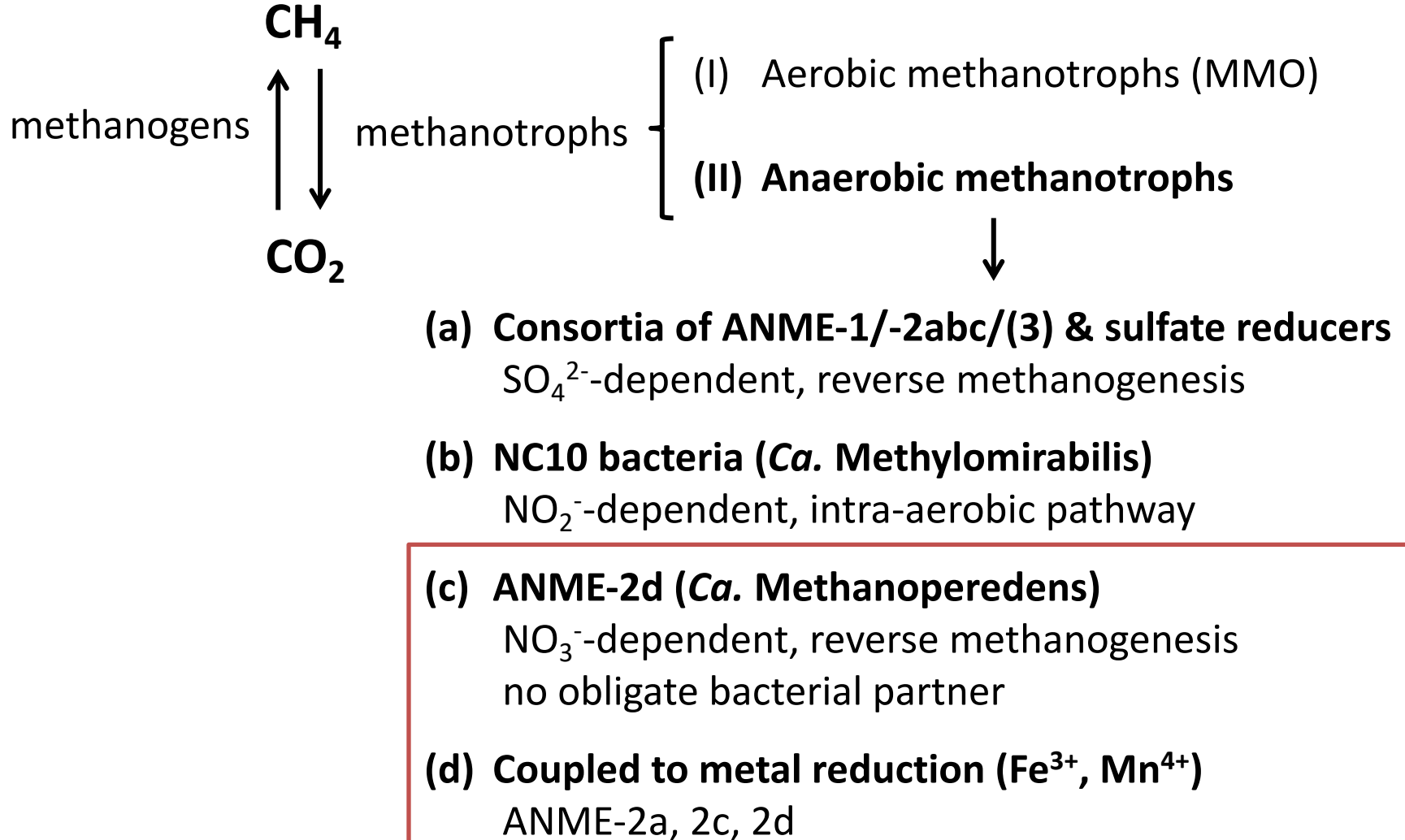


The fascinating world of methane microbes



**Understand the biochemistry,
physiology and interactions between
methanogens and methanotrophs**

Biological conversion of methane



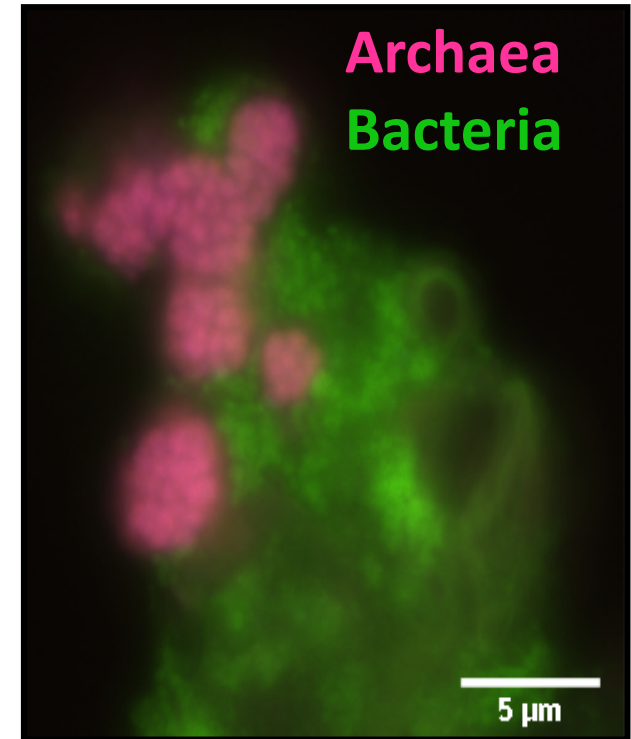
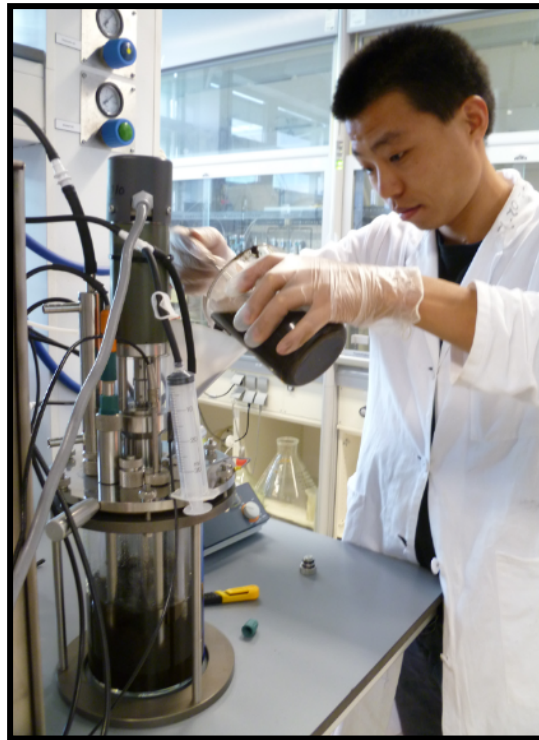
Sampling site nitrate/nitrite-dependent anaerobic oxidation of methane



High NO_3^- due to agricultural run-off / ground water

High CH_4 production in the sediment

Characterization of the enrichment culture



**Culture coupled methane oxidation to
nitrate and nitrite reduction**

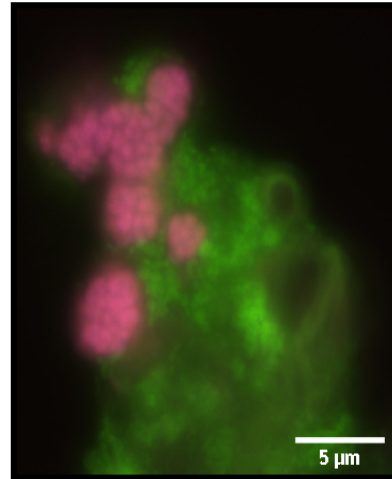
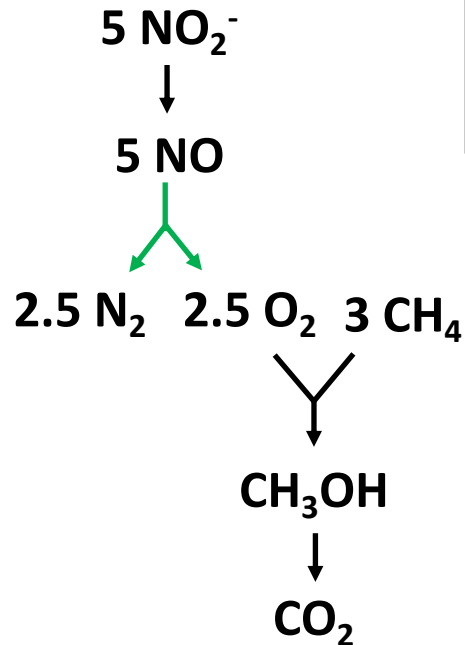
Raghoebarsing et al. (2006), Nature

Initial characterization of the enrichment culture

+ Nitrite: Bacteria

Ca. Methylospirillum oxyfera
Novel intra-aerobic pathway

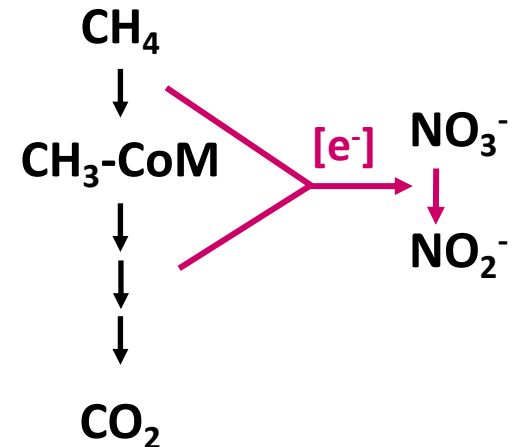
Novel NO
dismutase



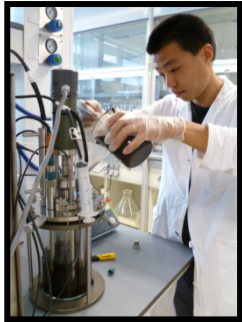
+ Nitrate: Archaea

Ca. Methanoperedens nitroreducens

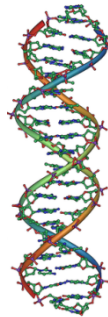
Mechanism?



Characterization of the enrichment culture



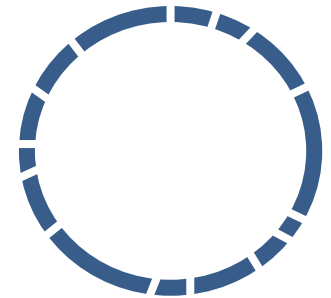
Reactor biomass



DNA
RNA



Metagenome
Metatranscriptome



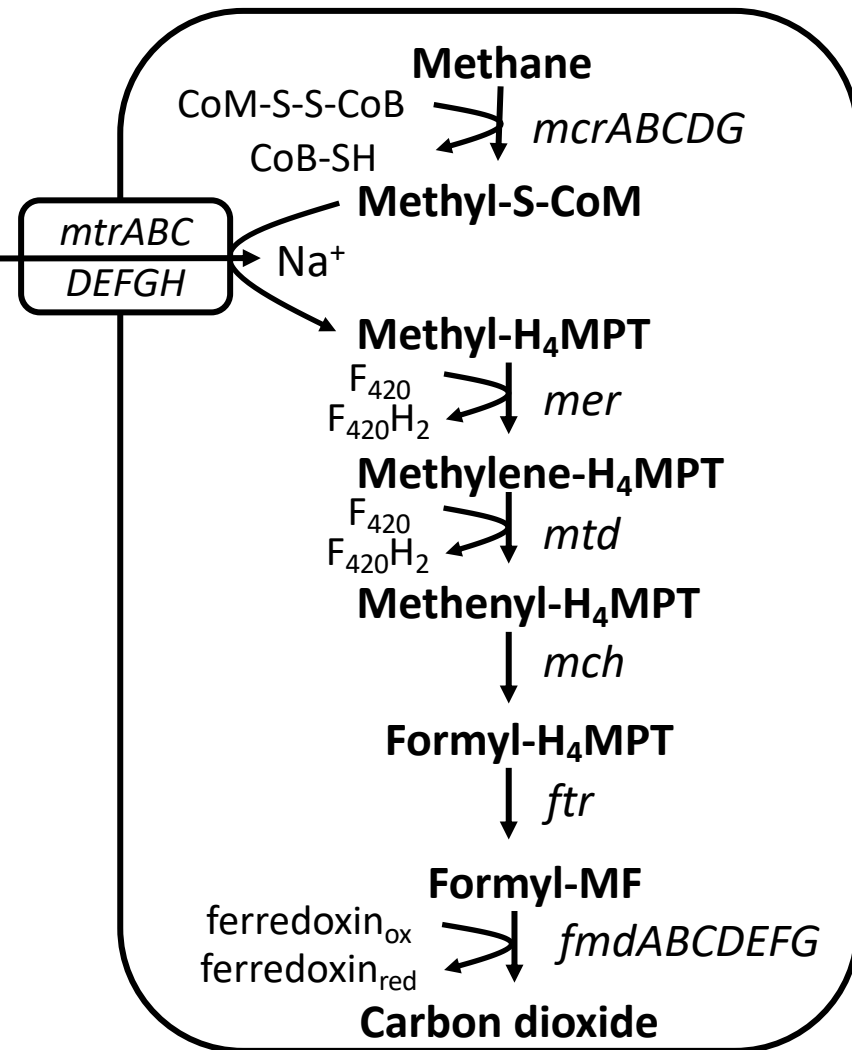
Binned
environmental
genome



A Metagenomics-Based Metabolic Model of Nitrate-Dependent Anaerobic Oxidation of Methane by *Methanoperedens*-Like Archaea

Arslan Arshad, Daan R. Speth, Rob M. de Graaf, Huub J. M. Op den Camp,
Mike S. M. Jetten and Cornelia U. Welte *

Central metabolic pathway of ANME archaea



A few considerations:

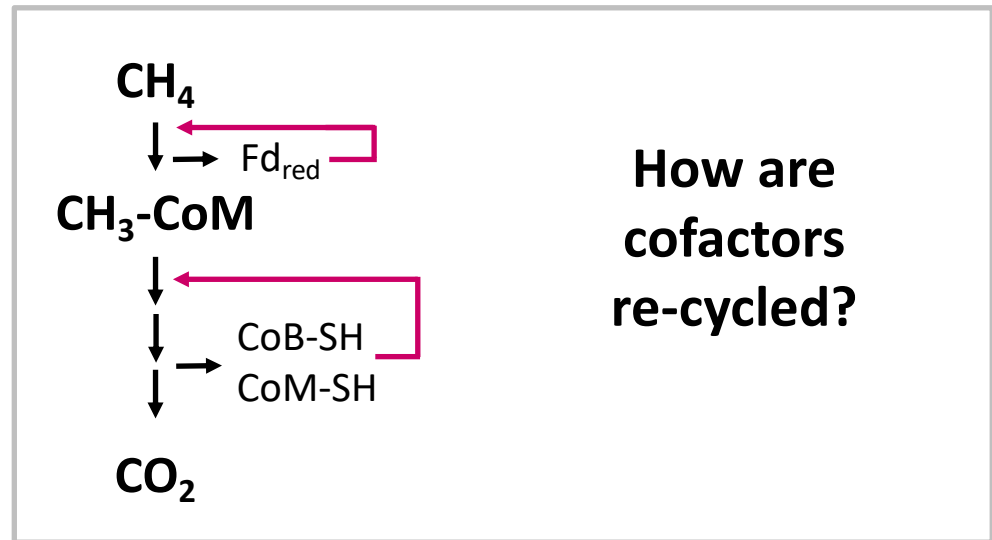
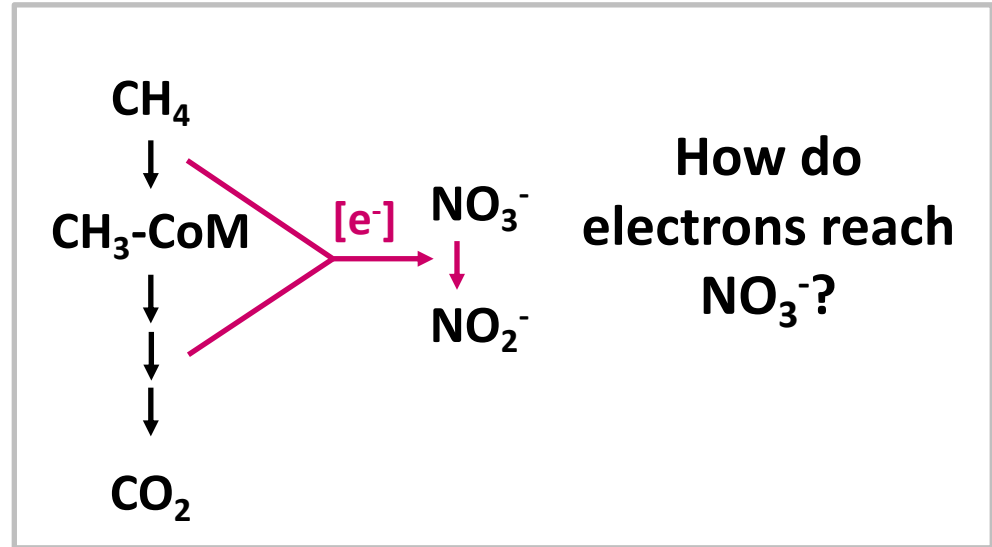
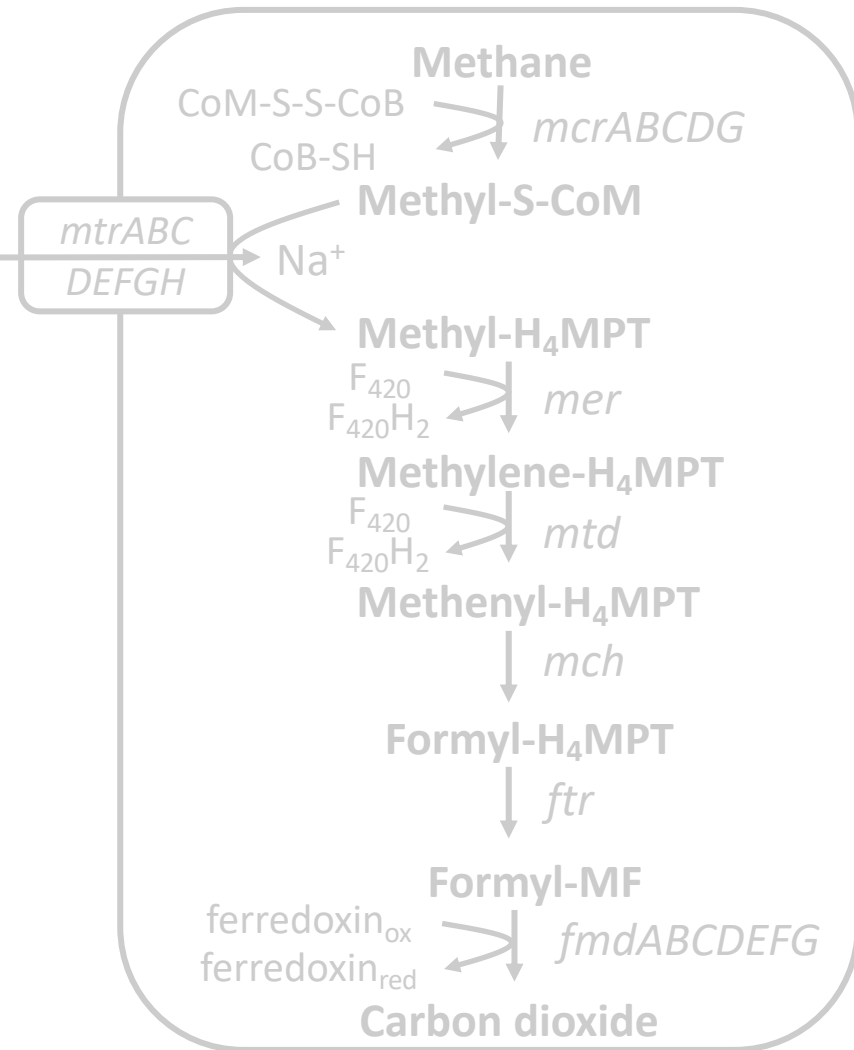
Just reverting a metabolic pathway leads to $\Delta G > 0$

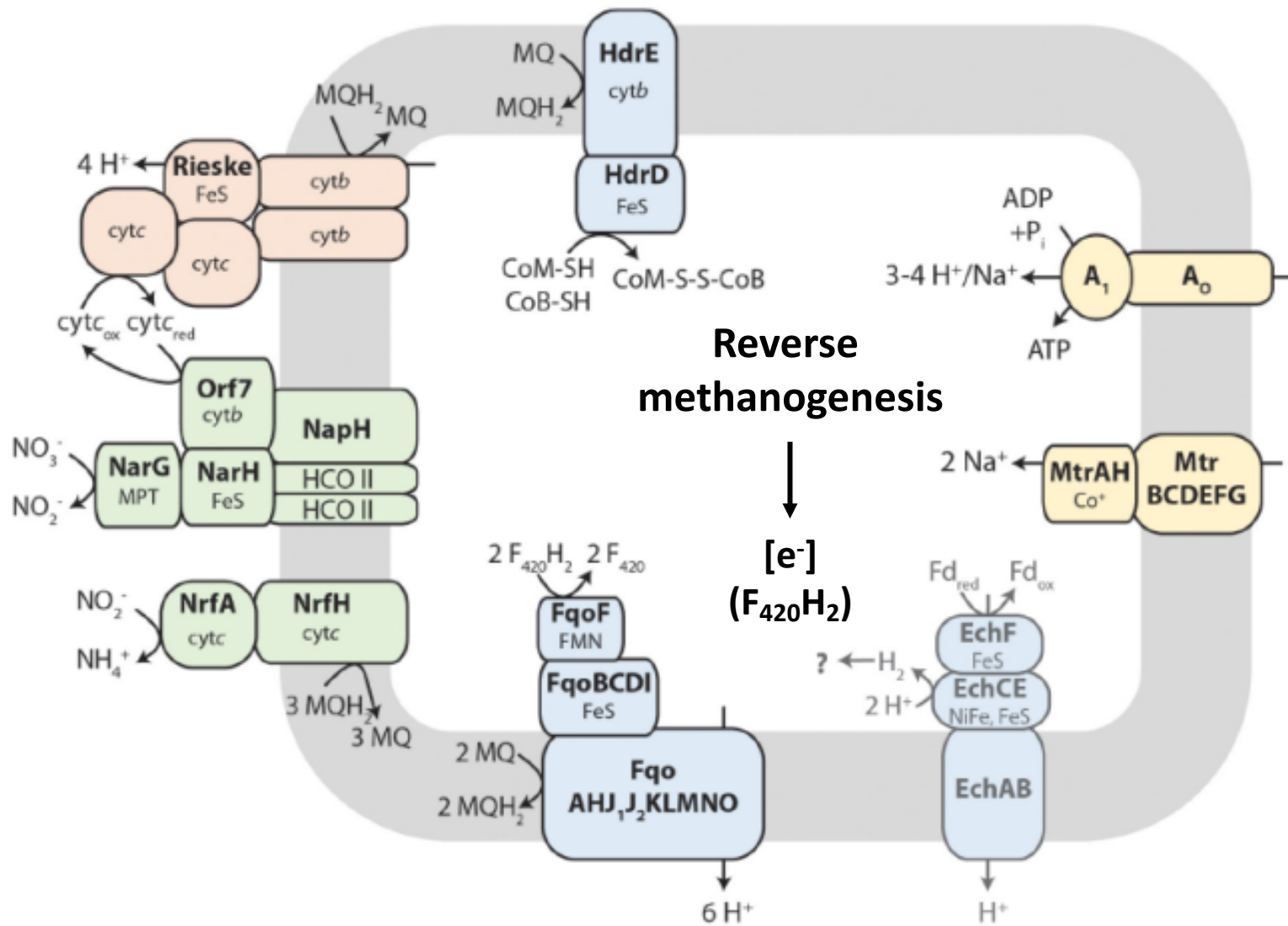
Redox-loops for the cofactors are not closed

Nitrate reduction is not yet included => needs to conserve energy to compensate for Mtr

The genome contains ca 50% hypothetical proteins!

Central metabolic pathway of ANME archaea

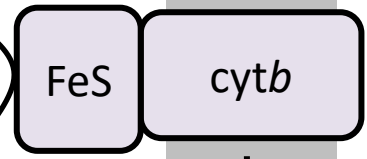




How are cofactors recycled?

Reverse
methanogenesis
↓
CoB-SH
CoM-SH

CoB-SH
CoM-SH
-120 mV
↓
CoM-S-S-CoB



Reverse
heterodisulfide
reductase

↓
[2 e⁻]
menaquinone
-80 mV
 $\Delta E^{0'} < 0$; $\Delta G^{0'} < 0$

methanophenazine
-180 mV
 $\Delta E^{0'} > 0$; $\Delta G^{0'} > 0$

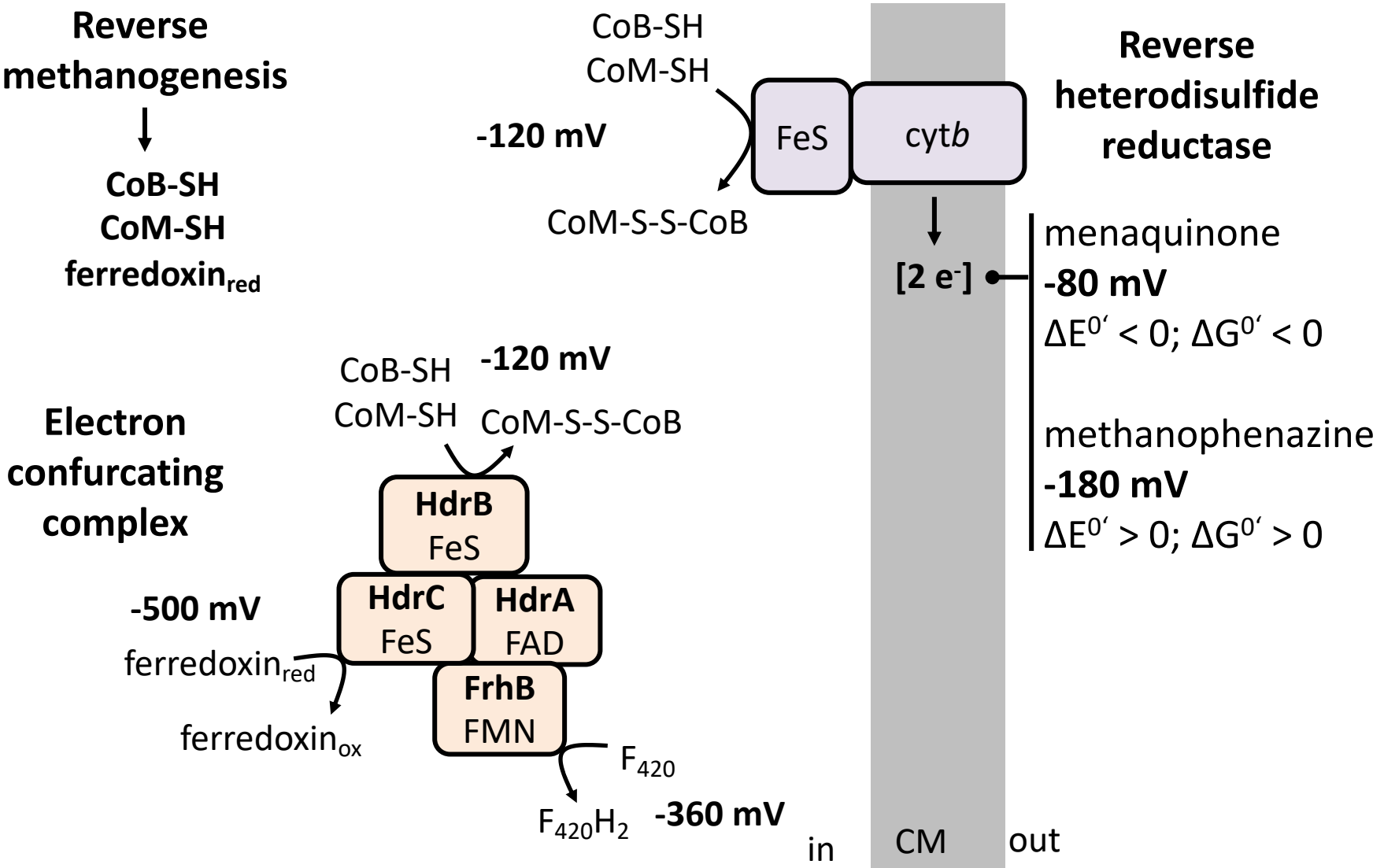
Menaquinone biosynthesis pathway present

Closely related methanogens don't use menaquinones!

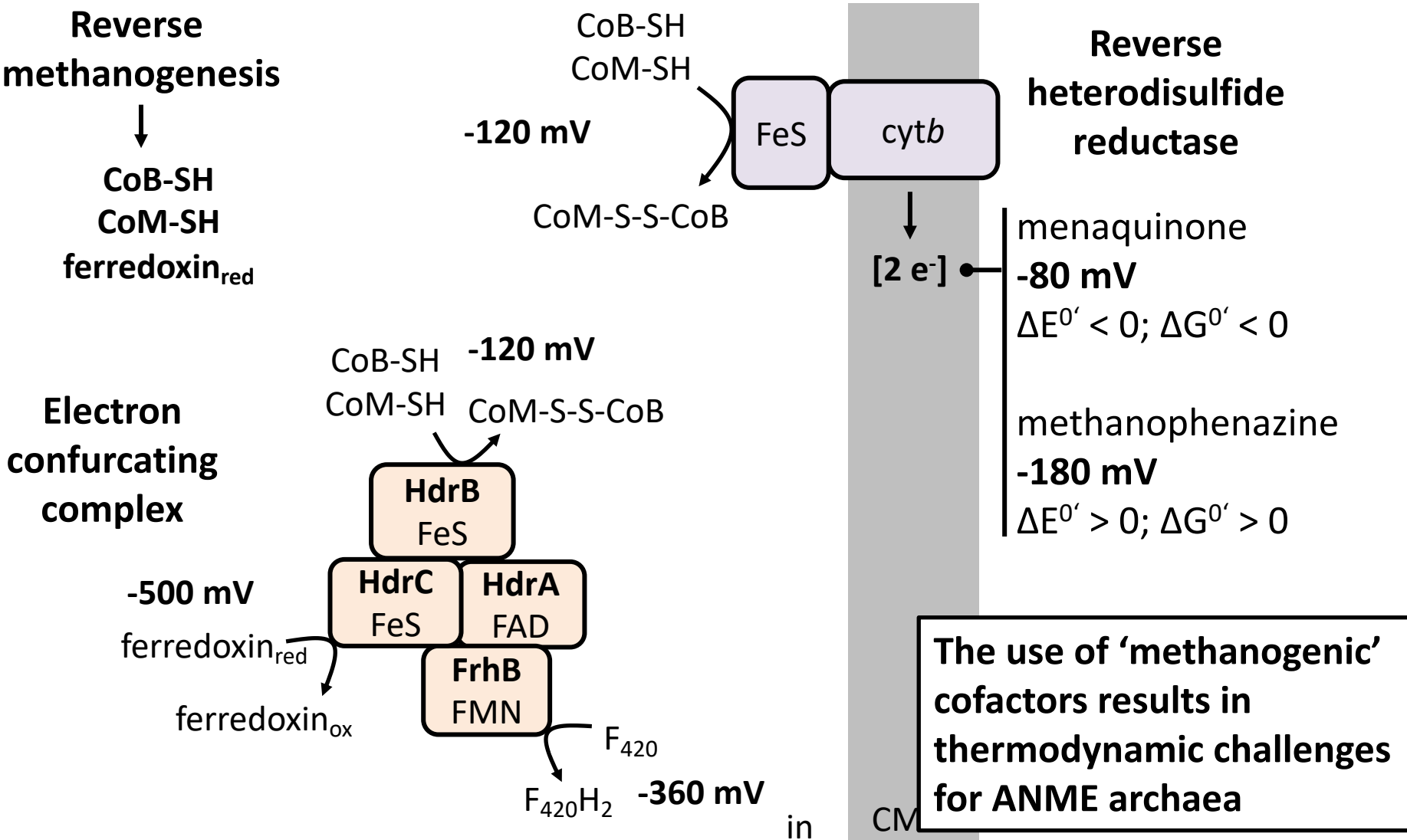
The diagram shows the chemical structure of a menaquinone, which consists of a naphthoquinone ring system with a long isoprenoid side chain. The side chain is represented as a repeating unit with a subscript 4, indicating four isoprenoid units.

in CM out

How are cofactors recycled?



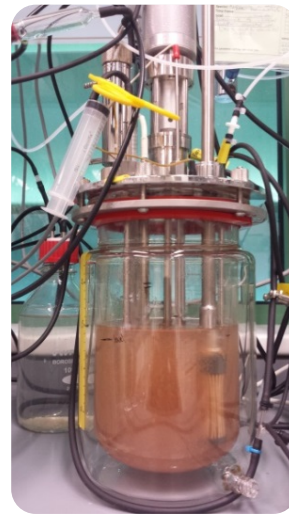
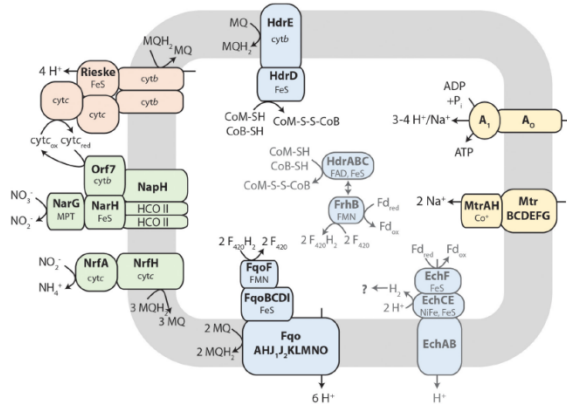
How are cofactors recycled?



Metagenome sequencing: conclusions

- (I) Complete reverse methanogenesis pathway encoded
- (II) Complex cytoplasmic and membrane-bound electron transport system
- (III) Unusual high number of *c*-type cytochromes found in archaea
- (IV) Methanotrophy is not just the reversal of methanogenesis!**

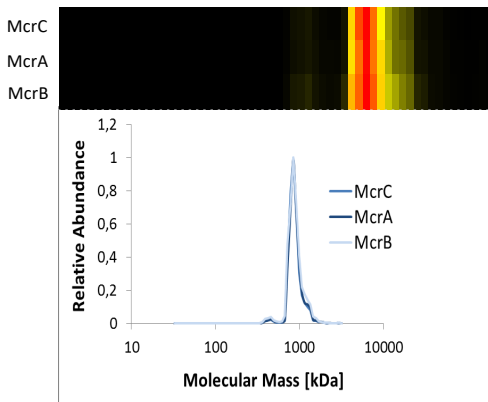
Progress made on *Ca. Methanoperedens archaea*



Physiology
Microbial interactions
 Maider Echeveste
 Dr. Julia Kurth
 Dr. Paula Dalcin Martins
 Dr. Martyna Glodowska

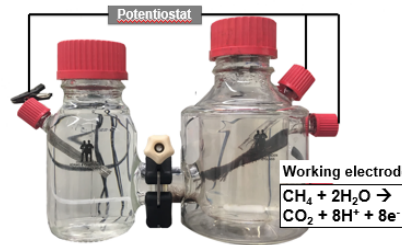
Genomic reconstruction

Dr. Arslan Arshad
 Dr. Stefanie Berger



Biochemistry & complexome analysis

Dr. Stefanie Berger

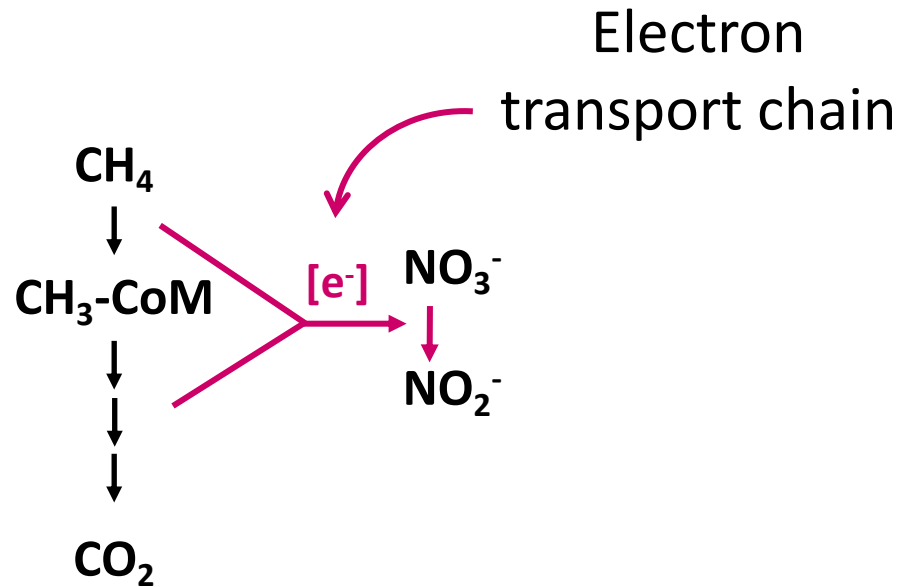


Electricity production & Extracellular electron transfer
 Heleen Ouboter



Application of ANMEs in wastewater treatment

Extracellular electron transfer by *Ca. Methanoperedens*



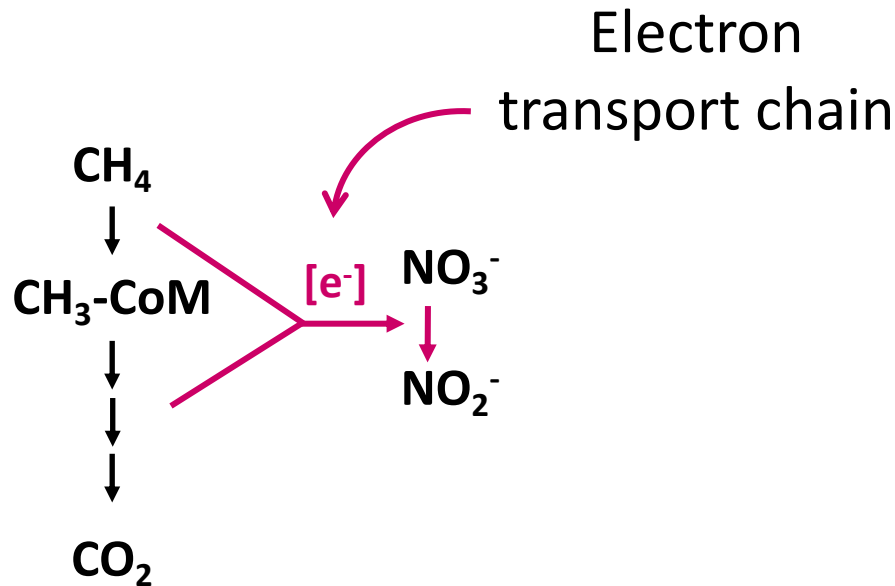
Heleen Ouboter

Can the electron acceptor be replaced by an electrode?

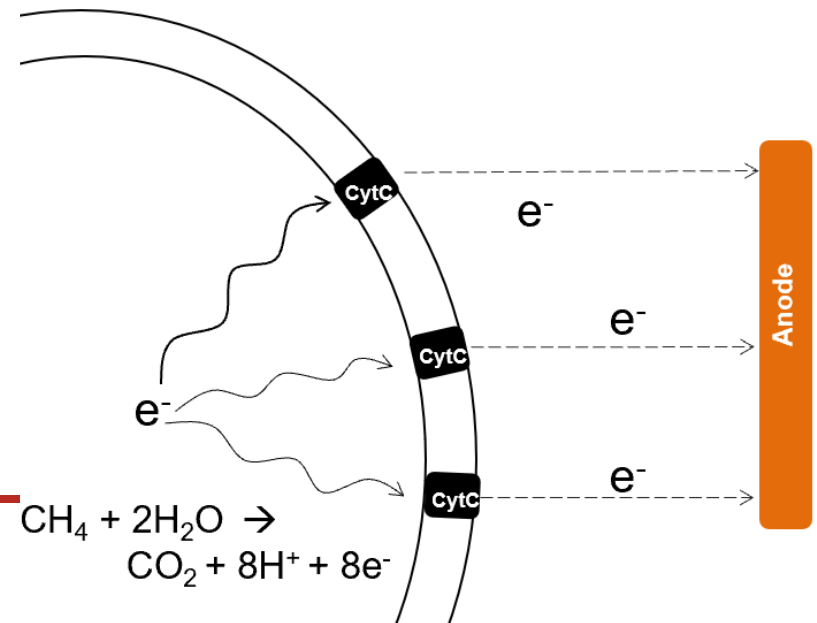
Extracellular electron transfer by *Ca. Methanoperedens*



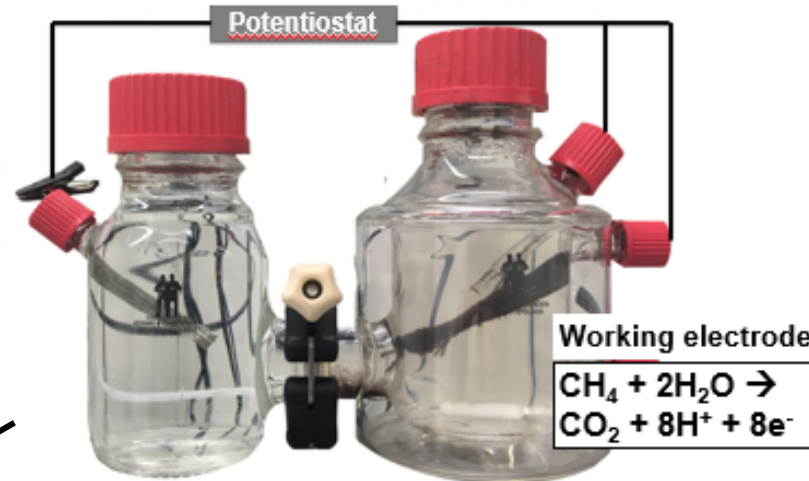
Heleen Ouboter



Can the electron acceptor be replaced by an electrode?

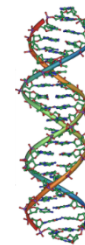


Growing *Ca. Methanoperedens* on a bioanode



Electrochemistry

Current production
Potential scans



Microbial community
bioanode & inoculum

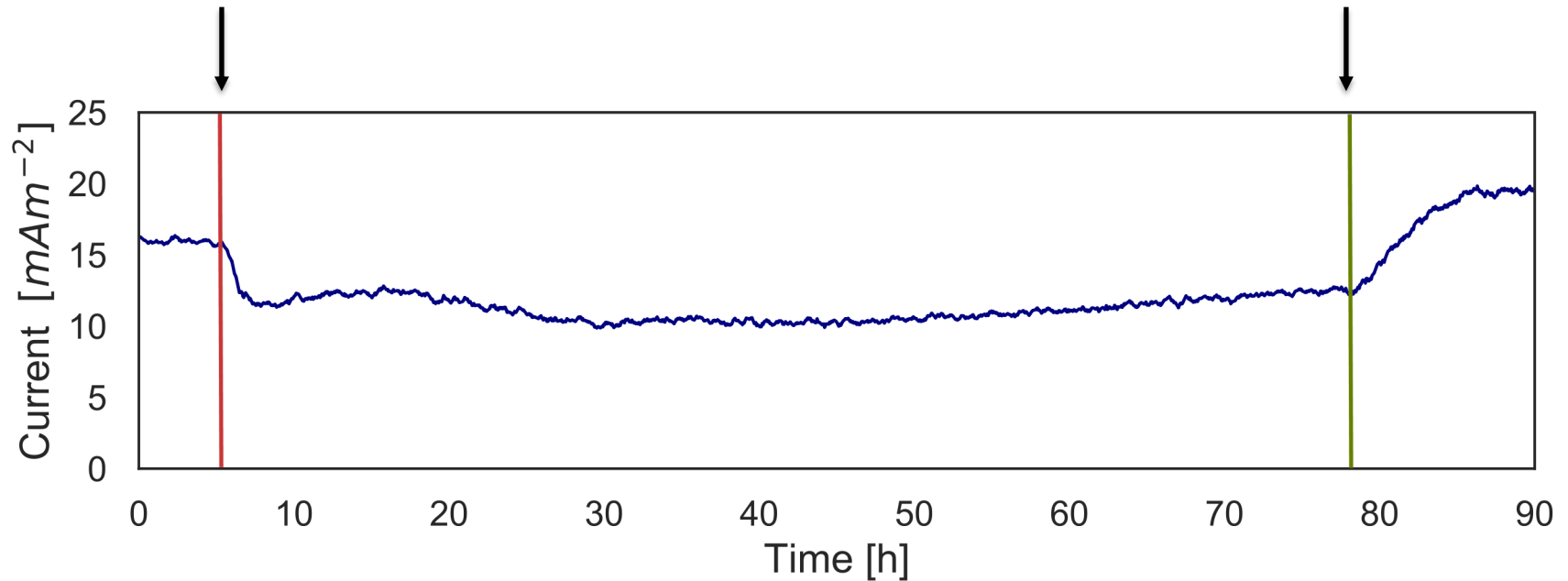
Growing *Ca. Methanoperedens* on a bioanode



Electrochemistry

Methane inflow stopped

Methane inflow re-started



Current production is methane dependent

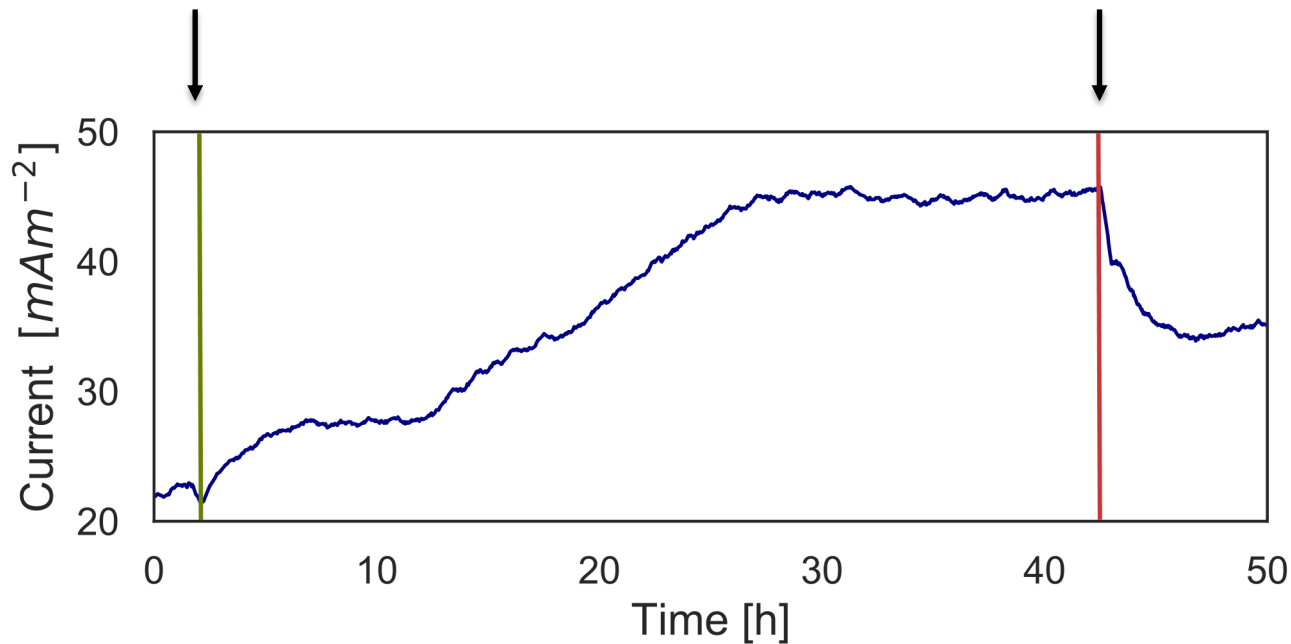
Growing *Ca. Methanoperedens* on a bioanode



Electrochemistry

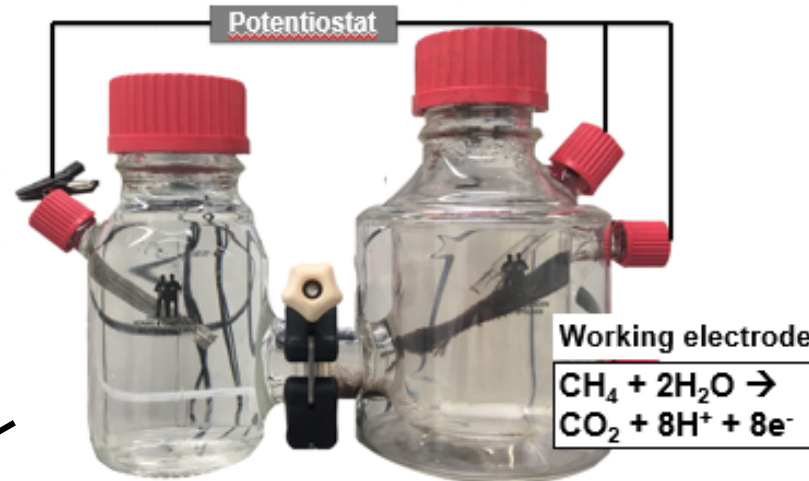
More methane added

System depressurized



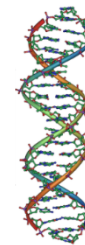
Current production is methane dependent

Growing *Ca. Methanoperedens* on a bioanode



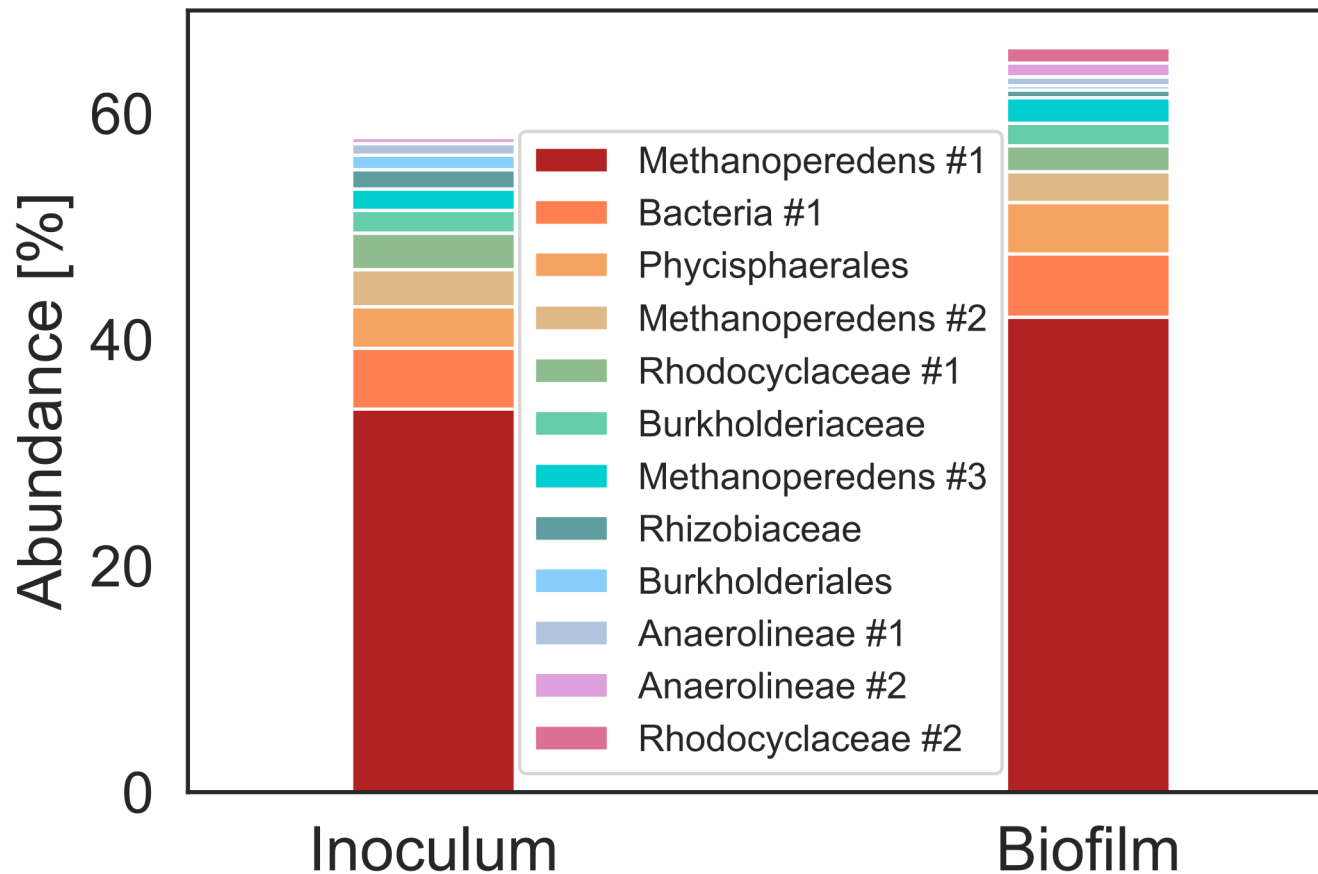
Electrochemistry

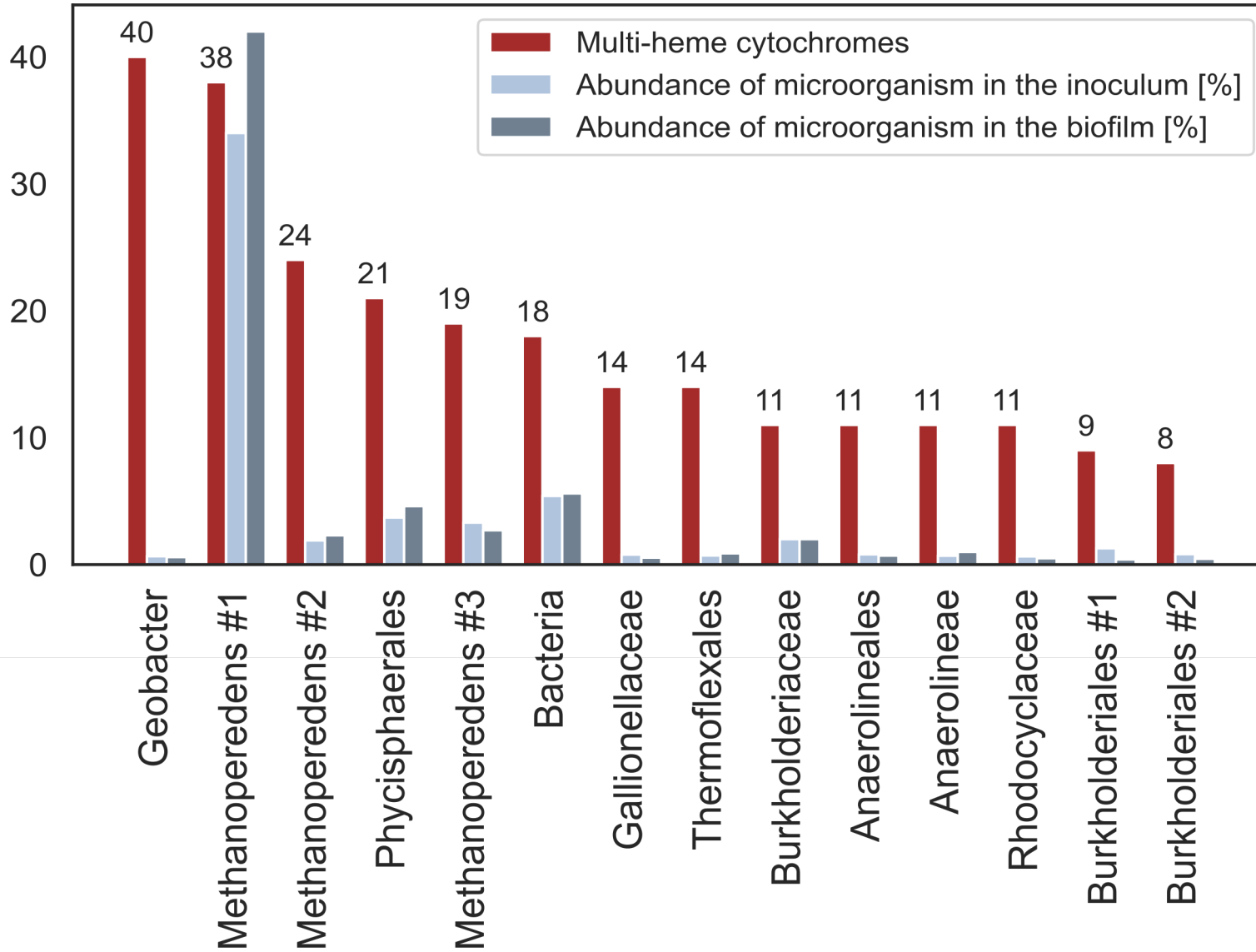
Current production
Potential scans



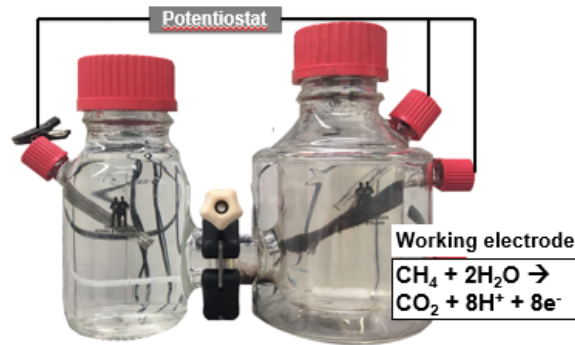
Microbial community
bioanode & inoculum

Metagenomics: *Ca.* Methanoperedens dominates both inoculum and bioanode





Growing *Ca. Methanoperedens* on a bioanode

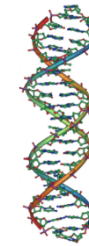


Proof-of-concept
Ca. Methanoperedens can be cultivated in bioelectrochemical system



Electrochemistry

Current production is methane dependent



Microbial community

Ca. Methanoperedens major player

Acknowledgements

Radboud University

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AIST Japan

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Wageningen University

Annemiek ter Heijne

Wetsus

Tom Sleutels

King Abdullah University of Science and Technology (KAUST)

Dario Shaw

**Thank you for
your attention!**



Bonus slides



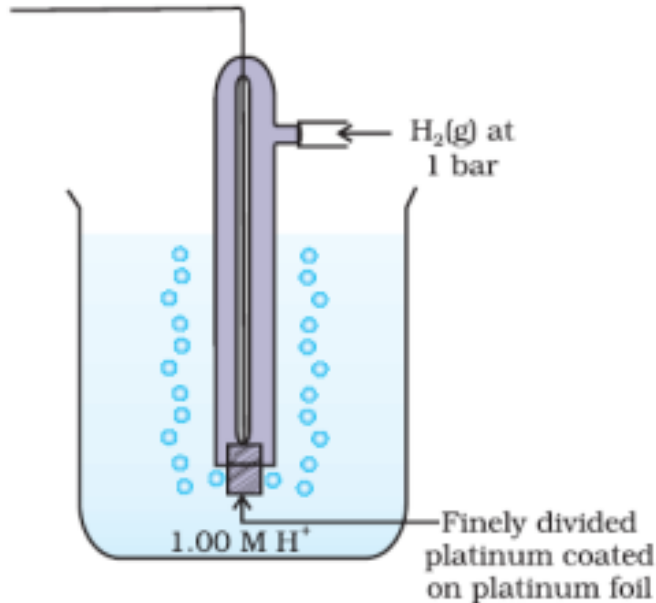
Redox potential

Wikipedia excerpt:

“Redox potential is a measure of the tendency of a chemical species to acquire electrons and thereby be reduced. Reduction potential is measured in volts (V), or millivolts (mV). Each species has its own intrinsic reduction potential; the more positive the potential, the greater the species' affinity for electrons and tendency to be reduced.”

The standard hydrogen electrode

Chemical conditions – pH 0



- 1 mol/L H⁺
- Continuous bubbling of Pt electrode
- Half cell / reference cell
- $E^0 = 0 \text{ V}$

Biological conditions – pH 7

- pH is usually not 0 (even for extremophiles – intracellular pH!)
- $E^0 (\text{H}^+/\text{H}_2) = -420 \text{ mV}$
 - ⇒ The “standard hydrogen electrode” is not the standard for biological systems
- $E^0=0$ has lost any meaning (if it ever had one); scale arbitrary but reference needed
- Whenever you think about gaseous electron donors/acceptors: constant bubbling over an electrode in a biological system...?

The redox-potential: real life

Displacement from standard conditions

$$E = E^{\circ} + 2.3 \frac{RT}{nF} \log_{10} \left\{ \frac{[\text{oxidised}]}{[\text{reduced}]} \right\} \quad \text{Nernst equation}$$

R Gas constant; $8.314 \text{ J K}^{-1} \text{ mol}^{-1}$

F Faraday's constant; $96500 \text{ J mol}^{-1} \text{ V}^{-1}$

n number of transferred electrons

Under equal concentrations of ox/red species, the second part of the equation is 0 and $E = E^{\circ}$

If the concentrations are very low, not thermodynamics but enzyme properties may become the limiting factor.

How to calculate Gibbs free energy

$$\Delta G^{0'} = -n \cdot F \cdot \Delta E^{0'}$$

“Gibbs free energy”

$\Delta G < 0$ exergonic

$\Delta G > 0$ endergonic

Number of
transferred
electrons

Faraday's constant
 $96.5 \text{ kJ mol}^{-1} \text{ V}^{-1}$

Redox potential
difference

$$\left. \begin{array}{l} E^{0'} (2\text{H}^+/\text{H}_2) = -414 \text{ mV} \\ E^{0'} (\frac{1}{2}\text{O}_2/\text{H}_2\text{O}) = +820 \text{ mV} \end{array} \right\} \Delta E^{0'} = +1234 \text{ mV} = +1.234 \text{ V}$$

$$\begin{aligned} \Delta G^{0'} &= -2 \cdot 96.5 \text{ kJ mol}^{-1} \text{ V}^{-1} \cdot 1.234 \text{ V} \\ &= -238 \text{ kJ mol}^{-1} \end{aligned}$$